

Package ‘TimerQuant’

April 17, 2025

Type Package
Title Timer Quantification
Version 1.38.0
Date 2015-05-22
Author Joseph Barry
Maintainer Joseph Barry <joseph.barry@embl.de>
Depends shiny
Suggests BiocStyle, reshape2, knitr, shinyBS
Imports ggplot2, grid, gridExtra, deSolve, dplyr, locfit
VignetteBuilder knitr
Description Supplementary Data package for tandem timer methods paper
by Barry et al. (2015) including TimerQuant shiny applications.
biocViews ExperimentData, Danio_rerio_Data, HighThroughputImagingData,
Tissue
License Artistic-2.0
LazyLoad yes
NeedsCompilation no
git_url <https://git.bioconductor.org/packages/TimerQuant>
git_branch RELEASE_3_21
git_last_commit b916b21
git_last_commit_date 2025-04-15
Repository Bioconductor 3.21
Date/Publication 2025-04-17

Contents

analyticSolutions	2
fitCV	3
FRETdata	4

genRatioHeatmap	4
genTimeSteadyStateHeatmap	5
getBreaks10	6
getSpacedSeq	6
maturationData	7
plotPrimordiumProfile	7
profileGradients	8
ratioSteadyState	8
ratioTimeDependent	9
runShinyApps	10
signal	11
simulatedSignal	11

Index	13
--------------	-----------

analyticSolutions	<i>Analytic model solutions for fluorescence intensity</i>
-------------------	--

Description

Time-dependent and steady-state analytic solution to one-step model for number of mature fluorophores. Where f is given as a parameter the returned value is transformed from a molecular population into a fluorescence intensity. For the function names, 0 refers to the dark population of non-mature fluorophores, and 1 to the mature, fluorescent population. 'ss' indicates steady-state solutions. tss is the time required to reach steady-state.

Usage

```
tss(m, k)
x0ss(p, m, k)
x1(p, m, k, t, f=1)
x1ss(p, m, k, f=1)
x1fretFP1(p, m1, m2, k, t, E=0, f=1)
x1fretFP1ss(p, m1, m2, k, E=0, f=1)
```

Arguments

p	Protein production rate (molecules produced per unit time).
m	Maturation rate of fluorophore, which can be for either FP1 or FP2 (convert to maturation time with $\log(2)/m$).
m1	Maturation rate of FP1.
m2	Maturation rate of FP2.
k	Protein degradation rate (convert to half-life with $\log(2)/k$).
t	Time (must be non-negative).
E	FRET coefficient representing energy transfer from FP1 to FP2.
f	Proportionality factor relating intensity to the number of molecules. When equal to one then the readout is number of molecules directly.

Value

A numeric specifying the model solution for the given parameters.

Author(s)

Joseph D. Barry

Examples

```
t0 <- seq(0.001, 1000, by=0.1)
plot(t0, x1(p=10, m=log(2)/5, k=log(2)/100, t=t0), type="l", col="darkgreen",
      lwd=2, xlab="Time (min)", ylab="Number of mature fluorophores", cex.lab=1.4)
```

fitCV	<i>fitCV</i>
-------	--------------

Description

Fits a smoothing line to coefficient of variation profiles.

Usage

```
fitCV(x, scaleLog10)
```

Arguments

x	A dataframe with columns Time (FP maturation time) and CV (coefficient of variation of timer signal).
scaleLog10	A logical indicating whether the points are spaced on the log10 scale or not.

Value

A dataframe containing fitted values and the minimum CV of the profile.

Author(s)

Joseph D. Barry

Examples

```
if (interactive()) vignette(topic="genPaperFigures", package="TimerQuant")
```

FRETdata

*FRET Data***Description**

A three-dimensional array of dimensions FRET value x tFT x assay type containing FRET readouts.

Usage

FRETdata

genRatioHeatmap

*generate ratio heatmap***Description**

Visualize timer ratios as a function of either FP1 or FP2 maturation time, and protein half-life.

Usage

```
genRatioHeatmap(tRangeFP, Tfixed, TA, TB, channel, E, f=1, n, ramp)
```

Arguments

tRangeFP	Vector containing two numerics specifying the range of maturation times to display for the chosen fluorescence channel.
Tfixed	The maturation time of the FP that will remain fixed.
TA	tFT half-life in location A.
TB	tFT half-life in location B.
channel	Integer specifying fluorescence channel to be varied (1 or 2).
E	FRET value representing transfer from FP1 to FP2.
f	$f=f_2/f_1$, the ratio of prefactors relating the number of molecules to the fluorescence intensity.
n	Integer specifying the number of data points. Choose a higher n for a higher pixel density.
ramp	Colour ramp, see colorRampPalette for more details.

Value

Returns a ggplot2 heatmap.

Author(s)

Joseph D. Barry

Examples

```
if (interactive()) vignette(topic="genPaperFigures", package="TimerQuant")
```

```
genTimeSteadyStateHeatmap
```

Time to reach steady-state

Description

Visualize the time to reach steady-state as a function of FP2 maturation time and protein half-life.

Usage

```
genTimeSteadyStateHeatmap(tRangeFP2, tRangeHlife, n, ramp)
```

Arguments

tRangeFP2	Vector containing two numerics specifying the range of FP2 maturation times.
tRangeHlife	Vector containing two numerics specifying the range of protein half-lives.
n	Integer specifying the number of data points. Choose a higher n for a higher pixel density.
ramp	Colour ramp, see colorRampPalette for more details.

Value

Returns a ggplot2 heatmap.

Author(s)

Joseph D. Barry

Examples

```
if (interactive()) vignette(topic="genPaperFigures", package="TimerQuant")
```

getBreaks10	<i>Get log10 breaks</i>
-------------	-------------------------

Description

Return breaks for each half-decade on the log10 scale, e.g. 1, 5, 10, 50, ...

Usage

```
getBreaks10(x)
```

Arguments

x	A vector of numbers. Breaks will be calculated across the range of x.
---	---

Value

A sequence of breaks useful for ticks or labels on the log10 scale.

Author(s)

Joseph D. Barry

Examples

```
getBreaks10(c(1, 100))
```

getSpacedSeq	<i>Get Spaced Sequence</i>
--------------	----------------------------

Description

Return points nicely spaced for on the log10 scale.

Usage

```
getSpacedSeq(x, n)
```

Arguments

x	A vector of two numbers containing the minimum and maximum of the desired sequence.
n	The desired length of the sequence to be returned.

Value

A sequence of numbers with appropriate spacing for the log10 scale.

Author(s)

Joseph D. Barry

Examples

```
getSpacedSeq(c(1, 1000), n=10)
```

maturationData	<i>Maturation Data</i>
----------------	------------------------

Description

A five-dimensional array of dimensions time (in minutes) x data columns x sample x view x tFT (identified by the RFP since sfGFP is present for all) containing fluorescence intensity readouts for the fluorophore maturation curves.

Usage

```
maturationData
```

plotPrimordiumProfile	<i>Plot Primordium Profile</i>
-----------------------	--------------------------------

Description

Visualizes primordium signal as a function of position with median and median absolute deviation across samples.

Usage

```
plotPrimordiumProfile(x, add, ylab, lwd, cex.lab, cex.axis, xlim, ylim, main,
  col, lty, alpha)
```

Arguments

x	A matrix of data where rows are samples and columns are sequential positions.
add	A logical indicating whether or not to add to the existing plot.
ylab	The y-axis label.
lwd	Integer specifying width of lines.
cex.lab	Integer specifying size of labels.
cex.axis	Integer specifying size of axis labels.
xlim	An optional vector of length 2 specifying the limits for the x-axis.
ylim	An optional vector of length 2 specifying the limits for the y-axis.

main	Plot title.
col	Line colour.
lty	Style of line
alpha	A numeric between zero and one specifying the level of transparency for the shaded region.

Value

Produces a plot of signal vs position summarizing across multiple primordium samples.

Author(s)

Joseph D. Barry

Examples

```
if (interactive()) vignette(topic="genPaperFigures", package="TimerQuant")
```

profileGradients	<i>Profile Gradients</i>
------------------	--------------------------

Description

A three-dimensional array of dimensions tFT x sample x position containing ratio readouts for migrating posterior lateral line primordia.

Usage

```
profileGradients
```

ratioSteadyState	<i>analytic function ratioSteadyState</i>
------------------	---

Description

Steady-state analytic solution to one-step model for the ratio of mature to non-mature fluorophores.

Usage

```
ratioSteadyState(T1, T2, halfLife, E=0, f=1)
```

Arguments

T1	Maturation time of fluorescent protein 1 (FP1, fast maturing).
T2	Maturation time of fluorescent protein 2 (FP2, slow maturing).
halfLife	Protein half-life.
E	FRET value representing transfer from FP1 to FP2.
f	$f=f_2/f_1$, the ratio of prefactors relating the number of molecules to fluorescence intensity for each fluorescence channel.

Value

A numeric specifying the model steady-state solution for the given parameters.

Author(s)

Joseph D. Barry

Examples

```
halfLifeSeq <- seq(1, 2000, by=0.1)
plot(halfLifeSeq, ratioSteadyState(T1=5, T2=100, halfLife=halfLifeSeq),
     type="l", lwd=2, ylim=c(0, 1), xlab="tFT half-life (min)",
     ylab="Steady-state ratio", cex.lab=1.4, log="x", col="red")
```

ratioTimeDependent	<i>analytic function ratioTimeDependent</i>
--------------------	---

Description

Steady-state analytic solution to one-step model for the ratio of mature to non-mature fluorophores.

Usage

```
ratioTimeDependent(T1, T2, halfLife, t, E=0, f=1)
```

Arguments

T1	Maturation time of fluorescent protein 1 (FP1, fast maturing).
T2	Maturation time of fluorescent protein 2 (FP2, slow maturing).
halfLife	Protein half-life.
t	Time, which must be non-negative.
E	FRET value representing energy transfer from FP1 to FP2.
f	$f=f_2/f_1$, the ratio of prefactors relating the number of molecules to fluorescence intensity.

Value

A numeric specifying the model time-dependent solution for the given parameters.

Author(s)

Joseph D. Barry

Examples

```
tSeq <- seq(0.1, 300, by=0.1)
plot(tSeq, ratioTimeDependent(T1=5, T2=100, halfLife=30, t=tSeq, E=0, f=1), type="l", lwd=2,
     xlab="time (min)", ylab="ratio", cex.lab=1.4, col="black", ylim=c(0, 0.3))
points(tSeq, ratioTimeDependent(T1=5, T2=100, halfLife=30, t=tSeq, E=0.4, f=1), type="l", lwd=2,
      col="red")
abline(h=ratioSteadyState(T1=5, T2=100, halfLife=30, E=0, f=1), lty=2, col="black")
abline(h=ratioSteadyState(T1=5, T2=100, halfLife=30, E=0.4, f=1), lty=2, col="red")
```

runShinyApps

Run R-shiny applications

Description

Wrapper functions that run shiny apps located in extdata subdirectory of R package.

Usage

```
runChooseFP2App()
runTimerModellingApp()
```

Author(s)

Joseph D. Barry

Examples

```
runChooseFP2App()

runTimerModellingApp()
```

`signal`*Signal*

Description

Computes timer signal (without additive noise) for a set of model parameters.

Usage

```
signal(T1, T2, TA, TB, E=0)
```

Arguments

T1	Maturation time of fluorescent protein 1 (fast maturing).
T2	Maturation time of fluorescent protein 2 (slow maturing).
TA	Minimum protein half-life.
TB	Maximum protein half-life.
E	FRET value representing transfer from FP1 to FP2.

Value

A numeric specifying the timer signal.

Author(s)

Joseph D. Barry

Examples

```
signal(T1=5, T2=60, TA=30, TB=180, E=0)
signal(T1=5, T2=60, TA=30, TB=180, E=0.5)
```

`simulatedSignal`*Simulated Timer Signal*

Description

Additive error model for timer signal.

Usage

```
simulatedRatio(T1, T2, hLife, sigmaAdd, p, E)
simulatedSignal(T1, T2, TA, TB, sigmaAdd, p, E)
simulatedSignalN(T1, T2, TA, TB, sigmaAdd, N, p, E)
```

Arguments

T1	Maturation time of fluorescent protein 1 (FP1, fast maturing).
T2	Maturation time of fluorescent protein 2 (FP2, slow maturing).
hLife	Protein half-life.
TA	Minimum protein half-life.
TB	Maximum protein half-life.
sigmaAdd	Standard deviation of normal distribution from which noise terms are drawn.
p	Protein production rate (molecules produced per unit time).
E	FRET value representing energy transfer from FP1 to FP2.
N	Number of simulation realizations.

Value

Returns simulated values for ratios or timer signal.

Author(s)

Joseph D. Barry

Examples

```
if (interactive()) vignette(topic="genPaperFigures", package="TimerQuant")
```

Index

- * **FRETdata**
 - FRETdata, [4](#)
- * **datasets**
 - maturationData, [7](#)
 - profileGradients, [8](#)
- * **fitCV**
 - fitCV, [3](#)
- * **genRatioHeatmap**
 - genRatioHeatmap, [4](#)
- * **genTimeSteadyStateHeatmap**
 - genTimeSteadyStateHeatmap, [5](#)
- * **getBreaks10**
 - getBreaks10, [6](#)
- * **getSpacedSeq**
 - getSpacedSeq, [6](#)
- * **plotPrimordiumProfile**
 - plotPrimordiumProfile, [7](#)
- * **ratioSteadyState**
 - ratioSteadyState, [8](#)
- * **ratioTimeDependent**
 - ratioTimeDependent, [9](#)
- * **runShinyApps**
 - runShinyApps, [10](#)
- * **signal**
 - signal, [11](#)
- * **simulatedSignal**
 - simulatedSignal, [11](#)
- * **x1**
 - analyticSolutions, [2](#)

[analyticSolutions, 2](#)

[fitCV, 3](#)

[FRETdata, 4](#)

[genRatioHeatmap, 4](#)

[genTimeSteadyStateHeatmap, 5](#)

[getBreaks10, 6](#)

[getSpacedSeq, 6](#)

[maturationData, 7](#)

[plotPrimordiumProfile, 7](#)

[profileGradients, 8](#)

[ratioSteadyState, 8](#)

[ratioTimeDependent, 9](#)

[runChooseFP2App \(runShinyApps\), 10](#)

[runShinyApps, 10](#)

[runTimerModellingApp \(runShinyApps\), 10](#)

[signal, 11](#)

[simulatedRatio \(simulatedSignal\), 11](#)

[simulatedSignal, 11](#)

[simulatedSignalN \(simulatedSignal\), 11](#)

[tss \(analyticSolutions\), 2](#)

[x0ss \(analyticSolutions\), 2](#)

[x1 \(analyticSolutions\), 2](#)

[x1fretFP1 \(analyticSolutions\), 2](#)

[x1fretFP1ss \(analyticSolutions\), 2](#)

[x1ss \(analyticSolutions\), 2](#)