

# Package ‘DEScan2’

April 10, 2025

**Type** Package

**Title** Differential Enrichment Scan 2

**Version** 1.27.0

**Date** 2023-03-09

**Maintainer** Dario Righelli <dario.righelli@gmail.com>

**Description** Integrated peak and differential caller, specifically designed for broad epigenomic signals.

**Encoding** UTF-8

**License** Artistic-2.0

**LazyData** TRUE

**biocViews** ImmunoOncology, PeakDetection, Epigenetics, Software, Sequencing, Coverage

**Depends** R (>= 3.5), GenomicRanges

**Imports** BiocParallel, BiocGenerics, ChIPpeakAnno, data.table, DelayedArray, GenomeInfoDb, GenomicAlignments, glue, IRanges, plyr, Rcpp (>= 0.12.13), rtracklayer, S4Vectors (>= 0.23.19), SummarizedExperiment, tools, utils

**LinkingTo** Rcpp, RcppArmadillo

**RoxygenNote** 7.2.3

**Suggests** BiocStyle, knitr, rmarkdown, testthat, edgeR, limma, EDASeq, RUVSeq, RColorBrewer, statmod

**VignetteBuilder** knitr

**git\_url** <https://git.bioconductor.org/packages/DEScan2>

**git\_branch** devel

**git\_last\_commit** b34d6d0

**git\_last\_commit\_date** 2024-10-29

**Repository** Bioconductor 3.21

**Date/Publication** 2025-04-10

**Author** Dario Righelli [aut, cre],  
 John Koberstein [aut],  
 Bruce Gomes [aut],  
 Nancy Zhang [aut],  
 Claudia Angelini [aut],  
 Lucia Peixoto [aut],  
 Davide Risso [aut]

## Contents

binnedCoverage . . . . .	3
binnedCovOnly . . . . .	4
binToChrCoordMatRowNames . . . . .	4
computeCoverageMovingWindowOnChr . . . . .	5
computeLambdaOnChr . . . . .	5
computeZ . . . . .	6
constructBedRanges . . . . .	7
countFinalRegions . . . . .	8
createGranges . . . . .	9
cutGRangesPerChromosome . . . . .	10
c_get_disjoint_max_win . . . . .	11
DEScan2 . . . . .	11
divideEachSampleByChromosomes . . . . .	12
evenRunMean . . . . .	13
evenRunSum . . . . .	13
finalRegions . . . . .	14
findOverlapsOverSamples . . . . .	15
findPeaks . . . . .	16
fromSamplesToChrsGRangesList . . . . .	18
generateDFofSamplesPerChromosomes . . . . .	18
get_disjoint_max_win . . . . .	19
giveUniqueNamesToPeaksOverSamples . . . . .	20
initMergedPeaksNames . . . . .	20
keepRelevantChrs . . . . .	21
rcpparma_get_disjoint_max_win . . . . .	21
readBamAsBed . . . . .	22
readBedFile . . . . .	23
readFilesAsGRangesList . . . . .	23
RleListToRleMatrix . . . . .	24
saveGRangesAsBed . . . . .	25
saveGRangesAsTsv . . . . .	26
setGRGenomeInfo . . . . .	27

## Index

28

---

binnedCoverage	<i>binnedCoverage</i>
----------------	-----------------------

---

## Description

this function computes the coverage over a binned chromosome, starting from a per base computed coverage.

## Usage

```
binnedCoverage(
  bins,
  numvar,
  mcolname,
  covMethod = c("max", "mean", "sum", "min"),
  roundingMethod = c("none", "floor", "ceiling", "round")
)
```

## Arguments

bins	a GRanges object representing a chromosome binned.
numvar	an RleList representing the per base coverage over the chr.
mcolname	the name of column where the sum have to be stored.
covMethod	a method to apply for the computing of the coverate it can be one of "max", "mean", "sum", "min". ("max" is default)
roundingMethod	a method to apply to round the computations it can be one of "none", "floor", "ceiling", "round". It's useful only when using covMethod="mean". ("none" is default)

## Value

the bins GRanges with the mcolname attached

## Examples

```
## dividing one chromosome in bins of 50 bp each
seqinfo <- GenomeInfoDb::Seqinfo(genome="mm9")
bins <- GenomicRanges::tileGenome(
  seqlengths=GenomeInfoDb::seqlengths(seqinfo)[1],
  tilewidth=50,
  cut.last.tile.in.chrom=TRUE)
gr <- GenomicRanges::GRanges(seqnames = S4Vectors::Rle("chr1", 100),
  ranges=IRanges::IRanges(start = seq(from=10, to=1000, by=10),
  end=seq(from=20, to=1010, by = 10)))
cov <- GenomicRanges::coverage(x=gr)
(binnedMaxCovGR <- binnedCoverage(bins, cov, "binned_cov"))
(binnedMeanCovGR <- binnedCoverage(bins, cov, "binned_cov",
```

```

covMethod="mean", roundingMethod="floor"))
(binnedSumCovGR <- binnedCoverage(bins, cov, "binned_cov", covMethod="sum"))

```

---

binnedCovOnly	<i>binnedCovOnly</i>
---------------	----------------------

---

### Description

it's useful just to coerce the bin coverage to an Rle object

### Usage

```
binnedCovOnly(bins, numvar, mcolname)
```

### Arguments

bins	a GRanges object representing a chromosome binned
numvar	an RleList representing the per base coverage over the chr
mcolname	the name of column where the sum have to be stored

### Value

an Rle within the per bin computed coverage

---

binToChrCoordMatRowNames	<i>binToChrCoordMatRowNames</i>
--------------------------	---------------------------------

---

### Description

computes the starting range of the bins for the binMatrix, taking in input the length of the chromosome of the matrix.

### Usage

```
binToChrCoordMatRowNames(binMatrix, chrLength, binWidth = 50)
```

### Arguments

binMatrix	a matrix where each row represents a bin.
chrLength	the length of the chromosome of the binMatrix.
binWidth	the width of the bin.

### Value

the binMatrix with start range as rownames.

---

```
computeCoverageMovingWindowOnChr
    computeCoverageMovingWindowOnChr
```

---

### Description

computes the coverage on a chromosome with a set of moving windows of dimensions minWinWidth:maxWinWidth

### Usage

```
computeCoverageMovingWindowOnChr(
  chrBedGRanges,
  minWinWidth = 50,
  maxWinWidth = 1000,
  binWidth = 50,
  verbose = TRUE
)
```

### Arguments

chrBedGRanges a GRanges to compute the coverage  
 minWinWidth the minimum width of the window to use for the coverage  
 maxWinWidth the maximum width of the window to use for the coverage  
 binWidth the dimension of the bin in base number

### Value

RleList where each element is a window within the Rle of its coverage

---

```
computeLambdaOnChr    computeLambdaOnChr
```

---

### Description

computes the lambdas on a chromosome for the winVector windows and other two windows (min/maxCompWinWidth) to compare with

**Usage**

```
computeLambdaOnChr(
  chrGRanges,
  winVector = seq_len(20),
  minChrRleWComp,
  minCompWinWidth = 5000,
  maxChrRleWComp,
  maxCompWinWidth = 10000,
  verbose = TRUE
)
```

**Arguments**

chrGRanges	the GRanges representing the reads of the chromosome.
winVector	the of width of the windows used to compute the coverage.
minChrRleWComp	and Rle object within coverage of window of width minCompWinWidth.
minCompWinWidth	the width of the window used for the coverage of minChrRleWComp in bases.
maxChrRleWComp	and Rle object within coverage of window of width minCompWinWidth.
maxCompWinWidth	the width of the window used for the coverage of maxChrRleWComp in bases.
verbose	verbose flag.
binSize	the size of the bin in bases.

**Value**

an RleList where each element is a window of winVector, within an Rle representing the lambda computed for that window.

---

computeZ

*computeZ*

---

**Description**

Computes Z-Scores returning the z matrix.

**Usage**

```
computeZ(
  lambdaChrRleList,
  runWinRleList,
  chrLength,
  minCount = 0.1,
  binSize = 50,
  verbose = FALSE
)
```

**Arguments**

lambdaChrRleList	an RleList of lambda values computed by computeLambdaOnChr function each element of the list is an Rle representing the lambda for the moving window in the list position.
runWinRleList	an RleList of coverage values computed. by computeCoverageMovingWindowOnChr function each element of the list is an Rle representing the coverage for the moving window in the list position.
chrLength	the length of the chr in analysis.
minCount	A small constant (usually no larger than one) to be added to the counts prior to the log transformation to avoid problems with log(0).
binSize	the size of the bin.
verbose	verbose output.

**Value**

z a matrix of z scores for each window (column) and bin (row). where the rownames represent the starting base of each bin.

---

constructBedRanges      *constructBedRanges*

---

**Description**

Constructs a GRanges object from a bam/bed/bed.zip file in a consistent way.

**Usage**

```
constructBedRanges(
  filename,
  filetype = c("bam", "bed", "bed.zip", "narrow", "broad"),
  genomeName = NULL,
  onlyStdChrs = FALSE,
  arePeaks = FALSE,
  verbose = FALSE
)
```

**Arguments**

filename	the complete file path of a bam?bed file.
filetype	the file type bam/bed/bed.zip/narrow/broad.
genomeName	the name of the genome used to map the reads (i.e. "mm9"). N.B. if NOT NULL the GRanges Seqinfo will be forced to genomeName Seqinfo (needs Internet access, but strongly suggested!)
onlyStdChrs	flag to keep only standard chromosome.
arePeaks	flag indicating if the file contains peaks.
verbose	flag to obtain verbose output.

**Value**

a GRanges object.

**Examples**

```
files <- list.files(system.file("extdata/bam/", package="DEScan2"),
                   pattern="bam$", full.names=TRUE)
bgr <- constructBedRanges(files[1], filetype="bam", genomeName="mm9",
                          onlyStdChrs=TRUE)
bgr
```

---

countFinalRegions	<i>countFinalRegions</i>
-------------------	--------------------------

---

**Description**

count reads falling within the final regions.

**Usage**

```
countFinalRegions(
  regionsGRanges,
  readsFilePath = NULL,
  fileType = c("bam", "bed"),
  minCarriers = 2,
  genomeName = NULL,
  onlyStdChrs = FALSE,
  carrierscolname = "k-carriers",
  ignStrandSO = TRUE,
  modeSO = "Union",
  saveFlag = FALSE,
  savePath = "finalRegions",
  verbose = TRUE
)
```

**Arguments**

regionsGRanges	a GRanges objects representing the peaks to compute the coverage, with a "k-carriers" mcols. (typically generated by finalRegions function).
readsFilePath	the filepath of bam or bed files necessary to compute the coverage.
fileType	the file type of the input files.
minCarriers	minimum number of carriers (samples).
genomeName	code name of the genome of reads files (i.e. "mm9").
onlyStdChrs	a flag indicating if to keep only the standard chromosomes



carrierscolname	character describing the name of the column within the carriers number (default is "k-carriers").
ignStrandSO	a flag indicating if to ignore the reads strand. (see GenomicAlignments::summarizeOverlaps).
modeSO	the mode to use, default is "Union". (see GenomicAlignments::summarizeOverlaps).
saveFlag	a flag indicating if to save the results.
savePath	the path where to store the results.
verbose	verbose output.

### Value

A SummarizedExperiment object containing as assays the read counts matrix with regions as rows and samples as columns, and as rowRanges the GRanges object representing the peaks used as rows in the matrix.

### Examples

```
filename <- system.file("extdata/regions/regions.rds", package="DEScan2")
regionsGR <- readRDS(file=filename)
reads.path <- system.file("extdata/bam", package="DEScan2")
finalRegionsSE <- countFinalRegions(regionsGRanges=regionsGR,
  readsFilePath=reads.path, fileType="bam", minCarriers=1,
  genomeName="mm9", onlyStdChrs=TRUE, ignStrandSO=TRUE, saveFlag=FALSE,
  verbose=TRUE)
library("SummarizedExperiment")
assay(finalRegionsSE) ## matrix of counts
rowRanges(finalRegionsSE) ## the GRanges of the input regions
```

---

createGranges	<i>createGranges</i>
---------------	----------------------

---

### Description

a simplified wrapper function to create a GRanges object.

### Usage

```
createGranges(chrSeqInfo, starts, widths, mcolname = NULL, mcolvalues = NULL)
```

### Arguments

chrSeqInfo	a seqinfo object.
starts	the start ranges.
widths	the width of each range.
mcolname	the name for the mcol attribute.
mcolvalues	the values for the mcol attribute.

**Value**

a GRanges object.

**Examples**

```
chrSeqInfo <- GenomeInfoDb::Seqinfo(genome="mm9")["chr1"]
starts=sample(seq_len(100), 10)
widths=starts+10;
mcolname <- "z-score";
mcolvalues <- sample(seq_len(100), 10)
chrGR <- createGranges(chrSeqInfo=chrSeqInfo, starts=starts, widths=widths,
                      mcolname=mcolname, mcolvalues=mcolvalues)
```

---

cutGRangesPerChromosome

*cutGRangesPerChromosome*

---

**Description**

takes in input a GRanges object, producing a LIST of GRanges, one for each chromosome.

**Usage**

```
cutGRangesPerChromosome(GRanges)
```

**Arguments**

GRanges            a GRanges object.

**Value**

a named list of GRanges, one for each chromosome.

**Examples**

```
library("GenomicRanges")
gr <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))
(grchrlist <- cutGRangesPerChromosome(gr))
```

---

```
c_get_disjoint_max_win
      c_get_disjoint_max_win
```

---

**Description**

just a wrapper for the C function. Useful to modify indexes and colnames.

**Usage**

```
c_get_disjoint_max_win(
  z0,
  sigwin = 10,
  nmax = 9999999,
  zthresh = 10,
  verbose = FALSE
)
```

**Arguments**

z0	the z matrix.
sigwin	the sigwin.
nmax	the nmax.
zthresh	peaks lower than this value will not be kept.
verbose	verbose flag.

**Value**

a matrix

---

DEScan2	<i>DEScan2</i>
---------	----------------

---

**Description**

integrated peak and differential caller, specifically designed for broad epigenomic signals.

**Author(s)**

some authors

---

divideEachSampleByChromosomes
*divideEachSampleByChromosomes*

---

**Description**

taken in input a grangeslist of samples, generate a list of samples where each element has a GRanges-List each element of the GRangesList represents a single chromosome.

**Usage**

```
divideEachSampleByChromosomes(samplesGRangesList)
```

**Arguments**

`samplesGRangesList`  
a GRangesList of samples.

**Value**

list of samples where each element is a list of chromosomes and each of these elements is a GRanges.

**Examples**

```
library("GenomicRanges")
gr1 <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))
gr2 <- GRanges(
  seqnames=Rle(c("chr1", "chr4", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr4=12, chr3=13))
sgr1 <- GRangesList(gr1, gr2)
names(sgr1) <- c("samp1", "samp2")
(sampChrGr1 <- divideEachSampleByChromosomes(sgr1))
```

---

evenRunMean	<i>evenRunMean</i>
-------------	--------------------

---

**Description**

this function computes a running mean over  $x$  with a window width  $k$  (modified from S4Vectors package to work on even  $k$ , see evenRunSum).

**Usage**

```
evenRunMean(x, k, endrule = c("drop", "constant"), na.rm = FALSE)
```

**Arguments**

$x$	an Rle object, typically a coverage object.
$k$	window dimension for the running sum over $x$ .
endrule	refer to S4Vectors::runMean.
na.rm	refer to S4Vectors::runMean.

**Value**

an Rle within the running mean over  $x$  with a win of length  $k$ .

---

evenRunSum	<i>evenRunSum</i>
------------	-------------------

---

**Description**

this function computes a running sum over  $x$  with a window width  $k$  (modified from S4Vectors package to work on even  $k$ , in such a case it adds a length at the end of the output Rle).

**Usage**

```
evenRunSum(x, k, endrule = c("drop", "constant"), na.rm = FALSE)
```

**Arguments**

$x$	an Rle object, typically a coverage object.
$k$	window dimension for the running sum over $x$ .
endrule	refer to S4Vectors::runSum.
na.rm	refer to S4Vectors::runSum.

**Value**

an Rle within the running sum over  $x$  with a win of length  $k$ .

---

finalRegions	<i>finalRegions</i>
--------------	---------------------

---

### Description

Align peaks to form common regions then filter regions for presence in multiple replicates taking in input a GRangesList where each element is a sample of called peaks.

### Usage

```
finalRegions(
  peakSamplesGRangesList,
  zThreshold = 20,
  minCarriers = 2,
  saveFlag = TRUE,
  outputFolder = "overlappedPeaks",
  verbose = FALSE,
  scorecolname = "z-score",
  coverageFlag = FALSE,
  BPPARAM = BiocParallel::bpparam()
)
```

### Arguments

peakSamplesGRangesList	named GRangesList where each element is a sample of called peaks. A score mcols values is needed for each GRanges. The scorecolname param can be used as reference name for the score. (typically returned by findPeaks function).
zThreshold	a minimum threshold for the z score. All peaks lesser than this value will be ignored.
minCarriers	a threshold of minimum samples (carriers) for overlapped regions.
saveFlag	a flag for saving results in a tsv file.
outputFolder	the directory name to store the bed file.
verbose	verbose output.
scorecolname	character describing the name of the column within the peaks score.
coverageFlag	boolean indicating if to compute the scores in a coverage mode (sum of the reads of merged peak) or in a score mode (a normalized score across the merged peaks)
BPPARAM	object of class bpparamClass that specifies the back-end to be used for computations. See <a href="#">bpparam</a> for details.

### Value

a GRanges of selected overlapping peaks with z-score, n-peaks, k-carriers as mcols object.

**Examples**

```

peak.path <- system.file("extdata/peaks/RData/peaksGRL_all_files.rds",
                        package="DEScan2")
gr1 <- readRDS(peak.path)
gr1

regionsGR <- finalRegions(peakSamplesGRangesList=gr1, zThreshold=1,
                          minCarriers=3, saveFlag=FALSE, verbose=TRUE)

```

---

```
findOverlapsOverSamples
```

```
findOverlapsOverSamples
```

---

**Description**

given in input a GRangesList where each element is a sample computes the coverage extending a both direction window of prefixed length.

**Usage**

```

findOverlapsOverSamples(
  samplePeaksGRangesList,
  extendRegions = 200,
  minOverlap = 0L,
  maxGap = -1L,
  zThresh = 10,
  verbose = FALSE,
  scorecolname = "z-score",
  coverageFlag = FALSE
)

```

**Arguments**

samplePeaksGRangesList	given a granges list of samples finds the overlapping regions between them.
extendRegions	the number of bases to extend each region at its start and end.
minOverlap	the minimum overlap each peak needs to have. (see ChipPeakAnno::findOverlapsOfPeaks)
maxGap	the maximum gap admissible between the peaks. (see ChipPeakAnno::findOverlapsOfPeaks)
zThresh	a threshold value on z-score/scorecolname
verbose	verbose flag
scorecolname	character describing the name of the column within the peaks score.
coverageFlag	boolean indicating if to compute the scores in a coverage mode (sum of the reads of merged peak) or in a score mode (a normalized score across the merged peaks)

**Value**

a GRanges of peaks overlapped and unique between samples.

**Examples**

```
(peaks.file <- system.file("extdata/peaks/RData/peaksGRL_all_files.rds",
                          package="DEScan2"))
peaksGRLFiles <- readRDS(peaks.file)
(overlPeaks <- findOverlapsOverSamples(peaksGRLFiles))
```

---

 findPeaks

*findPeaks*


---

**Description**

This function calls peaks from bed or bam inputs using a variable window scan with a poisson model using the surrounding maxCompWinWidth (10kb) as background.

**Usage**

```
findPeaks(
  files,
  filetype = c("bam", "bed"),
  genomeName = NULL,
  binSize = 50,
  minWin = 50,
  maxWin = 1000,
  zthresh = 10,
  minCount = 0.1,
  minCompWinWidth = 5000,
  maxCompWinWidth = 10000,
  outputFolder = "Peaks",
  save = TRUE,
  force = TRUE,
  verbose = FALSE,
  sigwin = 10,
  onlyStdChrs = TRUE,
  chr = NULL,
  BPPARAM = BiocParallel::bpparam()
)
```

**Arguments**

files	Character vector containing paths of files to be analyzed.
filetype	Character, either "bam" or "bed" indicating format of input file.
genomeName	the code of the genome to use as reference for the input files. (cfr. constructBedRanges function parameters)



binSize	Integer size in bases of the minimum window for scanning, 50 is the default.
minWin	Integer indicating the minimum window size in bases notation.
maxWin	Integer indicating the maximum window size in bases notation.
zthresh	Cutoff value for z-scores. Only windows with greater z-scores will be kept, default is 10.
minCount	A small constant (usually no larger than one) to be added to the counts prior to the log transformation to avoid problems with log(0).
minCompWinWidth	minimum bases width of a comparing window for Z-score.
maxCompWinWidth	maximum bases width of a comparing window for Z-score.
outputFolder	A string, Name of the folder to save the Peaks (optional) if the directory doesn't exist, it will be created. (Default is "Peaks")
save	Boolean, if TRUE files will be saved in a <code>"/Peaks/chr*"</code> directory created (if not already present) in the current working directory.
force	a boolean flag indicating if to force output overwriting.
verbose	if to show additional messages
sigwin	an integer value used to compute the length of the signal of a peak (default value is 10).
onlyStdChrs	a flag to work only with standard chromosomes. (cfr. <code>constructBedRanges</code> function parameters).
chr	if not NULL, a character like <code>"chr#"</code> indicating the chromosomes to use.
BPPARAM	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See <a href="#">bpparam</a> for details.

### Value

A `GRangesList` where each element is a sample. Each `GRanges` represents the founded peaks and attached the z-score of the peak as `mcols`.

### Examples

```
bam.files <- list.files(system.file("extdata/bam", package = "DEScan2"),
                      full.names = TRUE)

peaks <- findPeaks(files=bam.files[1], filetype="bam",
                  genomeName="mm9",
                  binSize=50, minWin=50, maxWin=1000,
                  zthresh=5, minCount=0.1, sigwin=10,
                  minCompWinWidth=5000, maxCompWinWidth=10000,
                  save=FALSE,
                  onlyStdChrs=TRUE,
                  chr=NULL,
                  verbose=FALSE)

head(peaks)
```

---

```
fromSamplesToChrsGRangesList
  fromSamplesToChrsGRangesList
```

---

**Description**

converts a GRangesList orgnized per samples to a GRangesList organized per Chromosomes where each element is a GRangesList of samples.

**Usage**

```
fromSamplesToChrsGRangesList(samplesGRangesList)
```

**Arguments**

```
samplesGRangesList
  a GRangesList of samples. Typically generaed by findPeaks function.
```

**Value**

A GRangesList of chromosomes where each element is a GRanges list of samples.

**Examples**

```
library("GenomicRanges")
gr1 <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))
gr2 <- GRanges(
  seqnames=Rle(c("chr1", "chr4", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr4=12, chr3=13))
sgr1 <- GRangesList(gr1, gr2)
names(sgr1) <- c("samp1", "samp2")
(chrGr1SampGr <- fromSamplesToChrsGRangesList(sgr1))
```

---

```
generatedDFofSamplesPerChromosomes
  generateDFofSamplesPerChromosomes
```

---

**Description**

generates a dataframe where each row is a sample (1st col) and a string with its chromosomes separated by ";" (2nd col) (useful to fromSamplesToChromosomesGRangesList function).

**Usage**

```
generateDFofSamplesPerChromosomes(samplesChrGRList)
```

**Arguments**

`samplesChrGRList`  
a GRangesList of samples each divided by chromosome.

**Value**

a dataframe where each row is a sample (1st col) and a string with its chromosomes separated by ";" (2nd col).

---

`get_disjoint_max_win`    *get\_disjoint\_max\_win*

---

**Description**

find significant z score windows keeping the max value without intersections

**Usage**

```
get_disjoint_max_win(  
  z0,  
  sigwin = 20,  
  nmax = Inf,  
  zthresh = -Inf,  
  two_sided = FALSE,  
  verbose = FALSE  
)
```

**Arguments**

`z0`                    Matrix containing z scores with bins as rows and windows size as columns.  
`sigwin`                Integer indicating how many bins per fragment.  
`nmax`                 Integer indicating the maximum number of windows to return.  
`zthresh`              Integer indicating the minimum z-score considered significant.  
`two_sided`            not used argument.  
`verbose`              verbose flag.

**Value**

a matrix of integer containing founded peaks

---

giveUniqueNamesToPeaksOverSamples  
*giveUniqueNamesToPeaksOverSamples*

---

**Description**

given a GRangesList of samples assigns unique names to the peaks of each sample.

**Usage**

giveUniqueNamesToPeaksOverSamples(samplePeaksGRangelist)

**Arguments**

samplePeaksGRangelist  
 a GRangesList of peaks, one GRanges for each sample.

**Value**

a GRangesList of samples within renamed peaks for each element.

---

initMergedPeaksNames *initMergedPeaksNames*

---

**Description**

given a GRanges of merged peaks assigns them new names.

**Usage**

initMergedPeaksNames(mergedGRanges)

**Arguments**

mergedGRanges A GRanges object. (Typically Generated in findOverlapsOverSamples function  
 )

**Value**

a granges of renamed peaks.

---

keepRelevantChrs	<i>keepRelevantChrs</i>
------------------	-------------------------

---

### Description

subselect a list of GRanges created with cutGRangesPerChromosome returning only the relevant chromosomes GRanges.

### Usage

```
keepRelevantChrs(chrGRangesList, chr = NULL)
```

### Arguments

`chrGRangesList` where each element is a chromosome, typically created with cutGRangesPerChromosome.

`chr` a character vector of chromosomes names of the form "chr#".

### Value

the input chrGRangesList with only the relevant chromosomes.

### Examples

```
library("GenomicRanges")
gr1 <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))
grlc <- cutGRangesPerChromosome(gr1)
(grlChr <- keepRelevantChrs(grlc, c("chr1", "chr3")))
```

---

rcpparma_get_disjoint_max_win	<i>rcpparma_get_disjoint_max_win</i> Computes the disjoint max_win matrix.
-------------------------------	--

---

### Description

rcpparma\_get\_disjoint\_max\_win Computes the disjoint max\_win matrix.

**Usage**

```
rcpparma_get_disjoint_max_win(
  z0,
  sigwin = 10L,
  zthresh = 10,
  nmax = 9999999L,
  verbose = TRUE
)
```

**Arguments**

<code>z0</code>	a matrix.
<code>sigwin</code>	sigwin.
<code>zthresh</code>	zthresh.
<code>nmax</code>	nmax.
<code>verbose</code>	verbose.

**Value**

a matrix of three columns (`bin_idx`, `win_idx`, `z_val`) idxs in C style.

---

readBamAsBed	<i>readBamAsBed</i>
--------------	---------------------

---

**Description**

read a bam file into a bed like format. forcing UCSC format for chromosomes names.

**Usage**

```
readBamAsBed(file)
```

**Arguments**

<code>file</code>	Character indicating path to bam file.
-------------------	--

**Value**

GRanges object.

**Examples**

```
files <- list.files(system.file("extdata/bam", package="DEScan2"),
  full.names=TRUE)
gr <- readBamAsBed(files[1])
```

---

readBedFile	<i>readBedFile</i>
-------------	--------------------

---

**Description**

read a bed file into a GenomicRanges like format. forcing UCSC format for chromosomes names.

**Usage**

```
readBedFile(filename, arePeaks = FALSE)
```

**Arguments**

filename	the bed filename.
arePeaks	a flag indicating if the the bed file represents peaks.

**Value**

GRanges object

**Examples**

```
bedFile <- list.files(system.file("extdata/bed", package="DEScan2"),
                      full.names=TRUE)
gr <- readBedFile(bedFile)
```

---

readFilesAsGRangesList	<i>readFilesAsGRangesList</i>
------------------------	-------------------------------

---

**Description**

Takes in input the path of bam/bed files to process and stores them in a GRangesList object, named with filePath/filenames. (for lazy people)

**Usage**

```
readFilesAsGRangesList(
  filePath,
  fileType = c("bam", "bed", "bed.zip", "narrow", "broad"),
  genomeName = NULL,
  onlyStdChrs = TRUE,
  arePeaks = TRUE,
  verbose = TRUE
)
```

**Arguments**

filePath	the path of input files.
fileType	the type of the files (bam/bed/bed.zip/narrow/broad).
genomeName	the genome code to associate to the files. (recommended) (i.e. "mm9", "hg17")
onlyStdChrs	a flag to keep only standard chromosomes.
arePeaks	a flag indicating if the files contain peaks.
verbose	verbose output flag.

**Value**

a GRangesList object

**Examples**

```
files.path <- system.file("extdata/bam", package="DEScan2")
grl <- readFilesAsGRangesList(filePath=files.path, fileType="bam",
                              genomeName="mm9", onlyStdChrs=TRUE,
                              verbose=TRUE)

class(grl)
names(grl)
grl
```

---

RleListToRleMatrix     *RleListToRleMatrix*

---

**Description**

a wrapper to create a RleMatrix from a RleList object.

**Usage**

```
RleListToRleMatrix(RleList, dimnames = NULL)
```

**Arguments**

RleList	an RleList object with all elements of the same length.
dimnames	the names for dimensions of RleMatrix (see DelayedArray pkg).

**Value**

a RleMatrix from DelayedArray package.



## Examples

```
library("DelayedArray")
lengths <- c(3, 1, 2)
values <- c(15, 5, 20)
e11 <- S4Vectors::Rle(values=values, lengths=lengths)

e12 <- S4Vectors::Rle(values=sort(values), lengths=lengths)

rleList <- IRanges::RleList(e11, e12)
names(rleList) <- c("one", "two")
(rleMat <- RleListToRleMatrix(rleList))
```

---

saveGRangesAsBed	<i>saveGRangesAsBed</i>
------------------	-------------------------

---

## Description

save a GRanges object as bed file.

## Usage

```
saveGRangesAsBed(
  GRanges,
  filepath = tempdir(),
  filename = tempfile(),
  force = FALSE,
  verbose = FALSE
)
```

## Arguments

GRanges	the GRanges object.
filepath	the path to store the files.@
filename	the name to give to the files.
force	force overwriting.
verbose	verbose output flag.

## Value

none

**Examples**

```
library("GenomicRanges")
gr <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))

saveGRangesAsBed(GRanges=gr, filepath=tempdir(), filename=tempfile(),
  verbose=TRUE)
```

---

saveGRangesAsTsv	<i>saveGRangesAsTsv</i>
------------------	-------------------------

---

**Description**

save a GRanges object as tsv file.

**Usage**

```
saveGRangesAsTsv(
  GRanges,
  filepath = tempdir(),
  filename = tempfile(),
  col.names = NA,
  row.names = TRUE,
  sep = "\t",
  force = FALSE,
  verbose = FALSE
)
```

**Arguments**

GRanges	the GRanges object.
filepath	the path to store the files.
filename	the name to give to the files.
col.names	a logical value indicating whether the column names are to be written in the file, or a character vector indicating the column names, or NA for writing column names for writing a TAB for the column name of the row names, default is NA (see <a href="#">write.table</a> ).
row.names	a logical value indicating whether the row names are to be written in the file, or a character vector indicating the row names (see <a href="#">write.table</a> ).
sep	the column separator character (default is <code>"\t"</code> ).
force	force overwriting.
verbose	verbose output flag.

**Value**

none

**Examples**

```
gr <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))
saveGRangesAsTsv(gr, verbose=TRUE)
```

---

setGRGenomeInfo	<i>setGRGenomeInfo</i> given a genome code (i.e. "mm9", "mm10", "hg19", "hg38") retrieve the SeqInfo of that genome and assigns it to the input GRanges. Finally filters out those Infos not necessary to the GRanges.
-----------------	--

---

**Description**

setGRGenomeInfo given a genome code (i.e. "mm9", "mm10", "hg19", "hg38") retrieve the SeqInfo of that genome and assigns it to the input GRanges. Finally filters out those Infos not necessary to the GRanges.

**Usage**

```
setGRGenomeInfo(GRanges, genomeName = NULL, verbose = FALSE)
```

**Arguments**

GRanges	a GRanges object.
genomeName	a genome code (i.e. "mm9")
verbose	verbose output

**Value**

a GRanges object with the seqinfo of the genome code

**Examples**

```
library("GenomicRanges")
gr <- GRanges(
  seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
  ranges=IRanges(1:10, end=10),
  strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
  seqlengths=c(chr1=11, chr2=12, chr3=13))
mm9gr <- setGRGenomeInfo(GRanges=gr, genomeName="mm9", verbose=TRUE)
```

# Index

- \* **internal.**
  - computeZ, 6
- \* **internal**
  - binnedCovOnly, 4
  - binToChrCoordMatRowNames, 4
  - c\_get\_disjoint\_max\_win, 11
  - computeCoverageMovingWindowOnChr, 5
  - computeLambdaOnChr, 5
  - evenRunMean, 13
  - evenRunSum, 13
  - generateDFofSamplesPerChromosomes, 18
  - get\_disjoint\_max\_win, 19
  - giveUniqueNamesToPeaksOverSamples, 20
  - initMergedPeaksNames, 20
  - rcpparma\_get\_disjoint\_max\_win, 21
- binnedCoverage, 3
- binnedCovOnly, 4
- binToChrCoordMatRowNames, 4
- bpparam, 14, 17
  
- c\_get\_disjoint\_max\_win, 11
- computeCoverageMovingWindowOnChr, 5
- computeLambdaOnChr, 5
- computeZ, 6
- constructBedRanges, 7
- countFinalRegions, 8
- createGranges, 9
- cutGRangesPerChromosome, 10
  
- DEScan2, 11
- divideEachSampleByChromosomes, 12
  
- evenRunMean, 13
- evenRunSum, 13
  
- finalRegions, 14
- findOverlapsOverSamples, 15
  
- findPeaks, 16
- fromSamplesToChrsGRangesList, 18
  
- generateDFofSamplesPerChromosomes, 18
- get\_disjoint\_max\_win, 19
- giveUniqueNamesToPeaksOverSamples, 20
  
- initMergedPeaksNames, 20
  
- keepRelevantChrs, 21
  
- rcpparma\_get\_disjoint\_max\_win, 21
- readBamAsBed, 22
- readBedFile, 23
- readFilesAsGRangesList, 23
- RleListToRleMatrix, 24
  
- saveGRangesAsBed, 25
- saveGRangesAsTsv, 26
- setGRGenomeInfo, 27
  
- write.table, 26