

# Package ‘RRHO’

April 1, 2025

**Type** Package

**Title** Inference on agreement between ordered lists

**Version** 1.46.0

**Date** 2014-06-26

**Author** Jonathan Rosenblatt and Jason Stein

**Maintainer** Jonathan Rosenblatt <john.ros.work@gmail.com>

**Description**

The package is aimed at inference on the amount of agreement in two sorted lists using the Rank-Rank Hypergeometric Overlap test.

**License** GPL-2

**Depends** R (>= 2.10), grid

**Imports** VennDiagram

**Suggests** lattice

**biocViews** Genetics, SequenceMatching, Microarray, Transcription

**git\_url** <https://git.bioconductor.org/packages/RRHO>

**git\_branch** RELEASE\_3\_20

**git\_last\_commit** ff3a12a

**git\_last\_commit\_date** 2024-10-29

**Repository** Bioconductor 3.20

**Date/Publication** 2025-03-31

## Contents

RRHO-package . . . . .	2
HNP . . . . .	2
pvalRRHO . . . . .	3
RRHO . . . . .	4
RRHOComparison . . . . .	6
<b>Index</b>	<b>8</b>

---

RRHO-package

*Test overlap using the Rank-Rank Hypergeometric test*

---

### Description

The package is aimed at inference on the amount of agreement in two sorted lists using the Rank-Rank Hypergeometric Overlap test.

### Details

Package: RRHO  
Type: Package  
Version: 0.3  
Date: 2013-06-21  
License: GPL-2

See [RRHO](#) to get started.

### Author(s)

Jonathan Rosenblatt and Jason Stein Maintainer: Jonathan Rosenblatt <[john.ros.work@gmail.com](mailto:john.ros.work@gmail.com)>

### See Also

[RRHO](#), [RRHOComparison](#)

---

HNP

*RRHO comparison data sets.*

---

### Description

RRHO comparison data sets. See references for details.

### Usage

```
data(lists)
```

### Format

Three data frames: HNP, My and Sestan. Each is a data.frame with gene identifiers and sorting values so that they can be used as inputes to [RRHOComparison](#).

### References

Stein JL\*, de la Torre-Ubieta L\*, Tian Y, Parikshak NN, Hernandez IA, Marchetto MC, Baker DK, Lu D, Lowe JK, Wexler EM, Muotri AR, Gage FH, Kosik KS, Geschwind DH. "A quantitative framework to evaluate modeling of cortical development by neural stem cells." Manuscript in press at Neuron. (\*) Authors contributed equally to this work.

**Examples**

```
data(lists)
str(HNP) ; str(Sestan); str(My)
```

---

pvalRRHO

---

*Compute the significance of the overlap between two lists*


---

**Description**

Computes the significance of the agreements between lists as returned by [RRHO](#) using resampling.

**Usage**

```
pvalRRHO(RRHO.obj, replications, stepsize=RRHO.obj$stepsize, FUN= max)
```

**Arguments**

RRHO.obj	The output object of the <a href="#">RRHO</a> function.
replications	The number of samples to be taken from the distribution of the aggregated test statistic.
stepsize	Controls the resolution of the test: how many items between any two overlap tests (i.e., between any two <i>i</i> -s and two <i>j</i> -s.)
FUN	The function aggregating information from the whole overlap matrix into one summary statistic. Typically the <i>min</i> pvalue, or <i>max</i> on $-\log(pval)$ scale.

**Details**

The distribution of  $FUN(-\log(pval))$  is computed using resampling.

The aggregating function will typically be the max function, corresponding to the maximal  $-\log(pvalue)$ , i.e., the most significant agreement over all sublists.

The distribution is computed by resampling pairs of null sequences, computing the significances of all the overlaps as done in the reference, applying the aggregating function supplied by the user, and returning the permutation based significance.

**Value**

pval	The FWER corrected significance of observed aggregated pvalue.
FUN.ecdf	The simulated sampling distribution of the aggregated pvalues.
FUN	The matrix aggregation function used. typically max for minimal p-value.
n.items	Length of lists.
stepsize	See <a href="#">RRHO</a>
replications	The number of simulation replications.
call	The function call.

**Note**

Might take a long time to run. Depending on the number of replications, the item (gene) count and the stepsize.

Also note that the significance returned is a conservative value (by a constant of  $1/\text{replications}$ ).

**Author(s)**

Jonathan Rosenblatt

**See Also**[RRHO](#)**Examples**

```
list.length <- 100
list.names <- paste('Gene',1:list.length, sep='')
gene.list1<- data.frame(list.names, sample(list.length))
gene.list2<- data.frame(list.names, sample(list.length))
RRHO.example <- RRHO(gene.list1, gene.list2, alternative='enrichment')
pval.testing <- pvalRRHO(RRHO.example,50)
```

RRHO

*Rank-Rank Hypergeometric Overlap Test***Description**

The function tests for significant overlap between two sorted lists using the method in the reference.

**Usage**

```
RRHO(
  list1, list2,
  stepsize = defaultStepSize(list1, list2),
  labels,
  alternative,
  plots = FALSE,
  outputdir = NULL,
  BY = FALSE,
  log10.ind=FALSE)
```

**Arguments**

list1	data.frame. First column is the element (possibly gene) identifier, and the second is its value on which to sort. For differential gene expression, values are often $-\log_{10}(\text{P-value}) * \text{sign}(\text{effect})$ .
list2	data.frame. Same as list1.
stepsize	Controls the resolution of the test: how many items between any two overlap tests.
labels	Character vector with two elements: the labels of the two lists.
alternative	Either "enrichment" for a one sided test, or "two.sided" for a two sided test. See Details section.
plots	Logical. Should output plots be returned?
outputdir	Path name where plots are returned.
BY	Logical. Should Benjamini-Yekutieli FDR corrected pvalues be computed?
log10.ind	Logical. Should pvalues be reported and plotted in $-\log_{10}$ scale and not $-\log$ scale?

## Details

Following the method in the reference, the function computes the number of overlapping elements in the first  $i * stepsize$  and  $j * stepsize$  elements of each list, and return the observed significance of this overlap using a hypergeometric test (see [fisher.test](#)). The output is returned as a list of matrices including: the overlap in the first  $i * stepsize, j * stepsize$  elements and the significance of this overlap.

If `plots=TRUE` then plots of these matrices are stored in .jpg format. In the case of `alternative='two.sided'` the pvalue plots are signed, just like in [1], thus distinguishing between over and under enrichment.

## Value

`hypermat` Matrix of  $-\log(pvals)$  of the test for the first  $i, j$  elements of the lists.  
`hypermat.counts` Counts of the number of agreements in the first  $i, j$  elements of the lists.  
`hypermat.by` An optional output of the B-Y corrected p-values of `hypermat`  
`hypermat.signs` Matrix of the type of deviation from the null. Negative for underenrichment and positive for overenrichment.

## Notes

By default, pvalues are reported in (minus) the natural log scale and not in (minus) log 10 scale. This behaviour is governed by `log10.ind`.

The p-values of the two-sided hypothesis test differ from those in reference [1]. This is because the two-sided p-values suggested in [1], are based on taking either the upper or lower tail of the distribution without appropriately using both tails. This method does not correctly control the type I error rate. In the implementation here, for a two-sided test we sum the probabilities from both tails of the hypergeometric distribution. See the package vignette for a small simulation.

## Author(s)

Jonathan Rosenblatt and Jason Stein

## References

- [1] Plaisier, Seema B., Richard Taschereau, Justin A. Wong, and Thomas G. Graeber. "Rank-rank Hypergeometric Overlap: Identification of Statistically Significant Overlap Between Gene-expression Signatures." *Nucleic Acids Research* 38, no. 17(September 1, 2010)
- [2] Benjamini, Y., and D. Yekutieli. 2001. "The Control of the False Discovery Rate in Multiple Testing Under Dependency." *ANNALS OF STATISTICS* 29 (4): 1165-1188.
- [3] Stein JL(\*), de la Torre-Ubieta L(\*), Tian Y, Parikshak NN, Hernandez IA, Marchetto MC, Baker DK, Lu D, Lowe JK, Wexler EM, Muotri AR, Gage FH, Kosik KS, Geschwind DH. "A quantitative framework to evaluate modeling of cortical development by neural stem cells." Manuscript in press at *Neuron*. (\*) Authors contributed equally to this work.

## See Also

[pvalRRHO](#); [RRHOComparison](#)

**Examples**

```
list.length <- 100
list.names <- paste('Gene',1:list.length, sep='')
gene.list1<- data.frame(list.names, sample(100))
gene.list2<- data.frame(list.names, sample(100))
# Enrichment alternative
RRHO.example <- RRHO(gene.list1, gene.list2, alternative='enrichment')
image(RRHO.example$hypermat)

# Two sided alternative
RRHO.example <- RRHO(gene.list1, gene.list2, alternative='two.sided')
image(RRHO.example$hypermat)
```

RRHOComparison

*Compares two RRHO maps to a third***Description**

Comparing two RRHO maps where one of the lists is shared between the two maps as in {RRHO map 1: list1 vs list3} vs {RRHO map 2: list2 vs list3}.

**Usage**

```
RRHOComparison(list1, list2, list3,
  stepsize, plots = FALSE,
  labels, outputdir = NULL,
  log10.ind)
```

**Arguments**

list1	A data.frame from experiment 1 with two columns, column 1 is the ‘Gene Identifier’, column 2 is the signed ranking value (e.g. signed $-\log_{10}$ of p-value, or fold change).
list2	Same as list1.
list3	Same as list1.
stepsize	Integer indicating how many genes to increase by in each algorithm iteration.
labels	Character vector carrying the labels for the outputted plots.
plots	Logical. Should comparisons be plotted?
outputdir	Plot destination directory.
log10.ind	Logical. Should pvalues be reported and plotted in $-\log_{10}$ scale and not $-\log$ scale?

**Details**

The difference in {overlap between list1 and list3} compared to the {overlap between list2 and list3}. This is useful for determining if there is a statistically significant difference between two RRHO maps. In other words, this is useful for determining if the overlap between list1 and list3 is statistically different between the overlap between list2 and list3.

RRHO Difference maps are produced by calculating for each pixel the normal approximation of difference in log odds ratio and standard error of overlap between the two RRHO maps. This Z score is then converted to a P-value and corrected for multiple comparisons across pixels [3].

The function will return a RRHO of the significance of overlap between `list1` and `list3` and `list2` and `list3`. A third RRHO gives the significance of the difference between these two overlap maps.

Note that by default all pvalues are outputted in `-log` scale. This can be changed with the `log10.ind` argument.

### Value

A object including:

<code>hypermat1</code>	Pvalues of comparing <code>list1</code> to <code>list3</code> .
<code>hypermat2</code>	Pvalues of comparing <code>list2</code> to <code>list3</code> .
<code>Pdiff</code>	The pvalue of the test for a difference in difference between lists 1-3 and 2-3.
<code>Pdiff.by</code>	Pvalues, corrected for the search over all of the list using Benjamini-Yekutieli.

### Author(s)

Jason Stein and Jonathan Rosenblatt

### References

- [1] Plaisier, Seema B., Richard Taschereau, Justin A. Wong, and Thomas G. Graeber. "Rank-rank Hypergeometric Overlap: Identification of Statistically Significant Overlap between Gene-Expression Signatures." *Nucleic Acids Research* 38, no. 17 (September 1, 2010): e169-e169.
- [2] Benjamini, Y., and D. Yekutieli. "The Control of the False Discovery Rate in Multiple Testing under Dependency." *ANNALS OF STATISTICS* 29, no. 4 (2001): 1165-88.
- [3] Stein JL\*, de la Torre-Ubieta L\*, Tian Y, Parikshak NN, Hernandez IA, Marchetto MC, Baker DK, Lu D, Lowe JK, Wexler EM, Muotri AR, Gage FH, Kosik KS, Geschwind DH. "A quantitative framework to evaluate modeling of cortical development by neural stem cells." Manuscript in press at *Neuron*. (\*) Authors contributed equally to this work.

### See Also

[RRHO](#)

### Examples

```
size<- 500
list1<- data.frame(GeneIdentifier=paste('gen',1:size, sep=' '),
RankingVal=-log(runif(size)))
list2<- data.frame(GeneIdentifier=paste('gen',1:size, sep=' '),
RankingVal=-log(runif(size)))
list3<- data.frame(GeneIdentifier=paste('gen',1:size, sep=' '),
RankingVal=-log(runif(size)))
(temp.dir<- tempdir())
RRHOComparison(list1,list2,list3,
               stepsize=10,
               labels=c("list1","list2","list3"),
               plots=TRUE,
               outputdir=temp.dir,
               log10.ind=FALSE)
```

# Index

\* **datasets**

HNP, [2](#)

\* **htest**

pvalRRHO, [3](#)

RRHO, [4](#)

RRHO-package, [2](#)

fisher.test, [5](#)

HNP, [2](#)

My (HNP), [2](#)

pvalRRHO, [3](#), [5](#)

RRHO, [2-4](#), [4](#), [7](#)

RRHO-package, [2](#)

RRHOComparison, [2](#), [5](#), [6](#)

Sestan (HNP), [2](#)