

# Package ‘GOstats’

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**Title** Tools for manipulating GO and microarrays

**Version** 2.54.0

**Description** A set of tools for interacting with GO and microarray data. A variety of basic manipulation tools for graphs, hypothesis testing and other simple calculations.

**Depends** R (>= 2.10), Biobase (>= 1.15.29), Category (>= 2.43.2), graph

**Imports** methods, stats, stats4, AnnotationDbi (>= 0.0.89), GO.db (>= 1.13.0), RBGL, annotate (>= 1.13.2), AnnotationForge, Rgraphviz

**Suggests** hgu95av2.db (>= 1.13.0), ALL, multtest, genefilter, RColorBrewer, xtable, SparseM, GSEABase, geneplotter, org.Hs.eg.db, RUnit, BiocGenerics

**License** Artistic-2.0

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GOstats-package	<i>Tools for manipulating GO and microarrays.</i>
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### Description

A set of tools for interacting with GO and microarray data. A variety of basic manipulation tools for graphs, hypothesis testing and other simple calculations.

### Details

Package: GOstats  
 Version: 1.7.4  
 Date: 23-08-2006  
 biocViews: Statistics, Annotation, GO, MultipleComparisons  
 Depends: graph (>= 1.9.25), GO, annotate, RBGL, xtable, Biobase, genefilter, multtest, Category (>= 1.3.7), methods  
 Imports: methods, Category  
 Suggests: hgu95av2.db (>= 1.6.0)  
 License: Artistic

#### Index:

ALL	Acute Lymphoblastic Leukemia Data from the Ritz Laboratory
GOstats-defunct	Defunct Functions in GOstats Package
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dropECode	Drop GO labels for specified Evidence Codes
getEvidence	Get the Evidence codes for a set of GO terms.
getGOTerm	Functions to Access GO data.
getOntology	Get GO terms for a specified ontology
hasGOannotate	Check for GO annotation

idx2dimnames	Index to Dimnames
makeGOGraph	Construct a GO Graph
notConn	Find genes that are not connected to the others.
oneGOGraph	Construct the GO graph given a set of leaves.
shortestPath	Shortest Path Analysis
simLL	Functions to compute similarities between GO graphs and also between Entrez Gene IDs based on their induced GO graphs.
triadCensus	Triad Functions

Further information is available in the following vignettes:

GOstats	Using GOstats (source, pdf)
GOusage	Basic GO Usage (source, pdf)
GOvis	Visualizing Data Using GOstats (source, pdf)

### Author(s)

R. Gentleman with contributions from S. Falcon

Maintainer: R. Gentleman <rgentlem@fhcrc.org>

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compCorrGraph	<i>A function to compute a correlation based graph from Gene Expression Data</i>
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---

### Description

Given a set of gene expression data (an instance of the ExpressionSet class) this function computes a graph based on correlations between the probes.

### Usage

```
compCorrGraph(eSet, k = 1, tau = 0.6)
```

### Arguments

eSet	An instance of the ExpressionSet class.
k	The power to raise the correlations to.
tau	The lower cutoff for absolute correlations.

### Details

Zhou et al. describe a method of computing a graph between probes (genes) based on estimated correlations between probes. This function implements some of their methods.

Pearson correlations between probes are computed and then these are raised to the power k. Any of the resulting estimates that are less than tau in absolute value are set to zero.

**Value**

An instance of the graph class. With edges and edge weights determined by applying the algorithm described previously.

**Author(s)**

R. Gentleman

**References**

Zhou et al., Transitive functional annotation by shortest-path analysis of gene expression data.

**See Also**

[compGdist](#)

**Examples**

```
## Create an ExpressionSet to work with
set.seed(123)
exprMat <- matrix(runif(50 * 5), nrow=50)
genData <- new("ExpressionSet", exprs=exprMat)

corrG = compCorrGraph(genData)
```

---

compGdist

*A function to compute the distance between pairs of nodes in a graph.*

---

**Description**

Given a graph, `g`, and a set of nodes in the graph, `whNodes`, Dijkstra's shortest path algorithm is used to compute the distance between all pairs of nodes in `whNodes`.

**Usage**

```
compGdist(g, whNodes, verbose = FALSE)
```

**Arguments**

<code>g</code>	An instance of the graph class.
<code>whNodes</code>	A vector of labels of the nodes in <code>g</code> for which distances are to be computed.
<code>verbose</code>	If TRUE then output reporting the progress will be reported.

**Details**

This function can be quite slow, computation of the pairwise distances is not especially fast and if `whNodes` is long then there are many of them to compute.

**Value**

A matrix containing the pairwise distances. It might be worth making this an instance of the `dist` class at some point.

**Author(s)**

R. Gentleman

**See Also**

[compCorrGraph](#)

**Examples**

```
example(compCorrGraph)
compGdist(corrG, nodes(corrG)[1:5])
```

---

GOHyperGResult-class    *Class "GOHyperGResult"*

---

**Description**

This class represents the results of a test for overrepresentation of GO categories among genes in a selected gene set based upon the Hypergeometric distribution.

For details on extracting information from this object, be sure to read the accessor documentation in the Category package: [HyperGResult-accessors](#).

**Objects from the Class**

Objects can be created by calls of the form `new("GOHyperGResult", ...)`.

**Slots**

**goDag:** Object of class "graph" representing the DAG of GO terms tested.

**pvalue.order:** Object of class "integer". The sort order of the computed p-values.

**annotation:** Object of class "character". The name of the annotation data package used in the analysis.

**geneIds:** Object of class "ANY". The intersection of the gene identifiers given as input and the computed gene universe.

**testName:** Object of class "character". Identifies the testing method used to produce this result instance.

**pvalueCutoff:** Object of class "numeric". The cutoff for significance used for some testing methods. Also used for pretty display in the show method.

**conditional:** A logical indicating whether the calculation should condition on the GO structure.

**testDirection:** A string which can be either "over" or "under". This determines whether the test performed detects over or under represented GO terms.

## Extends

Class "HyperGResultBase", directly.

## Methods

**goDag** signature(`r = "GOHyperGResult"`): return the graph instance representing the DAG of the GO terms that were tested.

**summary** signature(`r = "GOHyperGResult"`): Returns a data.frame summarizing the test result. Optional arguments `pvalue` and `categorySize` allow specification of maximum p-value and minimum categorySize, respectively. Optional argument `htmlLinks` is a logical value indicating whether to add HTML links (useful in conjunction with `xtables` print method with `type` set to "html").

**htmlReport** signature(`r = "GOHyperGResult"`): Write an HTML version of the table produced by the `summary` method. The path of a file to write the report to can be specified using the `file` argument. The default is `file=""` which will cause the report to be printed to the screen. If you wish to create a single report comprising multiple results you can set `append=TRUE`. The default is `FALSE` (overwrite preexisting report file). You can specify a string to use as an identifier for each table by providing a value for the `label` argument. Additional named arguments will be passed to the `summary` method.

**description** signature(`object = "GOHyperGResult"`): Return a string giving a one-line description of the result.

## Author(s)

Seth Falcon

## See Also

[HyperGResult-accessors](#)

---

GOstats-defunct

*Defunct Functions in GOstats Package*

---

## Description

The functions or variables listed here are no longer part of GOstats as they are not needed (any more).

## Usage

```
combGOGraph()
hyperGtable()
hyperG2Affy()
selectedGenes()
GOHyperG()
GOKEGGHyperG()
getGoGraph()
```

**Details**

combGOGraph was replaced by join. hyperGtable was replaced by summary. hyperG2Affy was replaced by probeSetSummary. GOLeaves was replaced by graph::leaves. selectedGenes was replaced by geneIdsByCategory. GOHyperG was replaced by hyperGTest. GOKEGGHyperG was replaced by hyperGTest. getGoGraph was replaced by GOGraph.

---

hyperGTest

*Hypergeometric Tests for GO term association*

---

**Description**

Given a GOHyperGParams instance containing a set of unique Entrez Gene Identifiers, a microarray annotation data package name, and the GO ontology of interest, this function will compute Hypergeometric p-values for over or under-representation of each GO term in the specified ontology among the GO annotations for the interesting genes. The computations can be done conditionally based on the structure of the GO graph.

**Arguments**

p                      A GOHyperGParams or OBOHyperGParams instance

**Details**

When conditional(p) == TRUE, the hyperGTest function uses the structure of the GO graph to estimate for each term whether or not there is evidence beyond that which is provided by the term's children to call the term in question statistically overrepresented.

The algorithm conditions on all child terms that are themselves significant at the specified p-value, odds ratio, minimum or maximum gene set size cutoff. Given a subgraph of one of the three GO ontologies, or the ontology given in the OBOHyperGParams instance, the terms with no child categories are tested first. Next the nodes whose children have already been tested are tested. If any of a given node's children tested significant, the appropriate conditioning is performed.

**Value**

A GOHyperGResult or OBOHyperGResult instance.

**Author(s)**

Seth Falcon

**References**

FIXME

**See Also**

[GOHyperGResult-class](#),  
[geneGoHyperGeoTest](#), [geneKeggHyperGeoTest](#)

idx2dimnames

*Index to Dimnames*

---

**Description**

A function to map from integer offsets in an array to the corresponding values of the row and column names. There is probably a better way but I didn't find it.

**Usage**

```
idx2dimnames(x, idx)
```

**Arguments**

x a matrix or data.frame.

idx An integer vector of offsets into the matrix (values between 1 and the length of the matrix).

**Value**

A list with two components. If it is a LIST, use

rowNames The row names corresponding to the integer index.

colNames The column names corresponding to the integer index.

**Author(s)**

R. Gentleman

**See Also**

[dimnames](#)

**Examples**

```
data(Ndists)
ltInf = is.finite(Ndists)
xx = idx2dimnames(Ndists, ltInf)
```

---

makeGOGraph*Construct a GO Graph*

---

**Description**

The directed acyclic graph (DAG) based on finding the most specific terms for the supplied Entrez Gene IDs is constructed and returned. The construction is per GO ontology (there are three, MF, BP and CC) and once the most specific terms have been identified then all less specific terms are found (these are the parents of the terms) and then their parents and so on, until the root is encountered.



**Usage**

```
makeGOGraph(x, Ontology = "MF", removeRoot = TRUE, mapfun = NULL,  
            chip = NULL)
```

**Arguments**

x	A vector of Entrez Gene IDs.
Ontology	Which GO ontology to use (CC, BP, or MF).
removeRoot	A logical value indicating whether the GO root node should be removed or not.
mapfun	A function taking a character vector of Entrez Gene IDs as its only argument and returning a list of "GO lists" matching the structure of the lists in the GO maps of annotation data packages. The function should behave similarly to <code>mget(x, eg2gomap, ifnotfound=NA)</code> , that is, NA should be returned if a specified Entrez ID has no GO mapping. See details for the interaction of <code>mapfun</code> and <code>chip</code> .
chip	The name of a DB-based annotation data package (the name will end in ".db"). This package will be used to generate an Entrez ID to GO ID mapping instead of <code>mapfun</code> .

**Details**

For each supplied Entrez Gene identifier all the GO annotations (in the specified ontology) are found. The mapping is achieved in one of three ways:

1. If `mapfun` is provided, it will be used to perform the needed lookups. In this case, `chip` will be ignored.
2. If `chip` is provided and `mapfun=NULL`, then the needed lookups will be done based on the Entrez to GO mappings encapsulated in the specified annotation data package. This is the recommended usage.
3. If `mapfun` and `chip` are NULL or missing, then the function will attempt to load the GO package (the environment-based package, distinct from GO.db). This package contains a legacy environment mapping Entrez IDs to GO IDs. If the GO package is not available, an error will be raised. Omitting both `mapfun` and `chip` is not recommended as it is not compatible with the DB-based annotation data packages.

The mappings are different for the different ontologies. Typically a GO identifier is used only in one specific ontology.

The resulting structure is stored in a graph using the graph package, again from Bioconductor.

**Value**

An object that inherits from the graph class. The particular implementation is not specified.

**Author(s)**

R. Gentleman

**References**

The Gene Ontology Consortium

**See Also**[oneGOGraph](#)**Examples**

```
library("hgu95av2.db")
set.seed(321)
gN <- unique(sample(keys(hgu95av2.db, 'ENTREZID'), 4))
gg1 <- makeGOGraph(gN, "BP", chip="hgu95av2.db")
```

---

**Ndists***Distance matrices for the BCR/ABL and NEG subgroups.*

---

**Description**

These are precomputed distance matrices between all transcription factors selected. In the future they will be computed on the fly but currently that takes about 3 hours and so precomputed versions are supplied.

**Usage**

```
data(Ndists)
data(Bdists)
```

**Format**

These are both distance matrices.

**Source**

They are based on the ALL data, [ALL](#).

**Examples**

```
data(Ndists)
data(Bdists)
```

---

**notConn***Find genes that are not connected to the others.*

---

**Description**

A function that takes as input a distance matrix and finds those entries that are not connected to any others (ie. those with distance Inf).

**Usage**

```
notConn(dists)
```

**Arguments**

dists            A distance matrix.

**Details**

It is a very naive implementation. It presumes that not connected entries are not connected to any other entries, and this might not be true. Using the `connComp` function from the `graph` package or the `RBGL` package might be a better approach.

**Value**

A vector of the names of the items that are not connected.

**Author(s)**

R. Gentleman

**See Also**

[connComp](#)

**Examples**

```
data(Ndists)
notConn(Ndists)
```

---

OBOHyperGResult-class    *Class "OBOHyperGResult"*

---

**Description**

This class represents the results of a test for overrepresentation of OBO categories among genes in a selected gene set based upon the Hypergeometric distribution.

For details on extracting information from this object, be sure to read the accessor documentation in the `Category` package: [HyperGResult-accessors](#).

**Objects from the Class**

Objects can be created by calls of the form `new("OBOHyperGResult", ...)`.

**Slots**

`goDag`: Object of class "graph" representing the DAG of OBO terms tested.

`pvalue.order`: Object of class "integer". The sort order of the computed p-values.

`annotation`: Object of class "character". The name of the annotation data package used in the analysis.

`geneIds`: Object of class "ANY". The intersection of the gene identifiers given as input and the computed gene universe.

`testName`: Object of class "character". Identifies the testing method used to produce this result instance.

**pvalueCutoff:** Object of class "numeric". The cutoff for significance used for some testing methods. Also used for pretty display in the show method.

**conditional:** A logical indicating whether the calculation should condition on the OBO structure.

**testDirection:** A string which can be either "over" or "under". This determines whether the test performed detects over or under represented OBO terms.

### Extends

Class "HyperGResultBase", directly.

### Methods

**goDag** signature(*r* = "OBOHyperGResult"): return the graph instance representing the DAG of the OBO terms that were tested.

**summary** signature(*r* = "OBOHyperGResult"): Returns a data.frame summarizing the test result. Optional arguments *pvalue* and *categorySize* allow specification of maximum p-value and minimum categorySize, respectively. Optional argument *htmlLinks* is a logical value indicating whether to add HTML links (useful in conjunction with *xtables* print method with *type* set to "html").

**htmlReport** signature(*r* = "OBOHyperGResult"): Write an HTML version of the table produced by the summary method. The path of a file to write the report to can be specified using the *file* argument. The default is *file=""* which will cause the report to be printed to the screen. If you wish to create a single report comprising multiple results you can set *append=TRUE*. The default is *FALSE* (overwrite preexisting report file). You can specify a string to use as an identifier for each table by providing a value for the *label* argument. Additional named arguments will be passed to the summary method.

**description** signature(*object* = "OBOHyperGResult"): Return a string giving a one-line description of the result.

### Author(s)

Robert Castelo

### See Also

[HyperGResult-accessors](#)

---

oneGOGraph

*Construct the GO graph given a set of leaves.*

---

### Description

Given one or more GO identifiers (which indicate the leaves in the graph) and a set of mappings to the less specific sets of nodes this function will construct the graph that includes that node and all children down to the root node for the ontology.

### Usage

```
oneGOGraph(x, dataenv)
GOGraph(x, dataenv)
```

**Arguments**

x                    A character vector of GO identifiers.  
dataenv             An environment for finding the parents of that term.

**Details**

For any set of GO identifiers (from a common ontology) we define the induced GO graph to be that graph, based on the DAG structure (child - parent) of the GO ontology of terms, which takes the most specific set of GO terms that apply (for that ontology) and then joins these to all less specific terms. These functions help construct such graphs.

**Value**

The induced GO graph (or NULL) for the given GO identifier.

**Author(s)**

R. Gentleman

**See Also**

[makeGOGraph](#)

**Examples**

```
library("GO.db")
g1 <- oneGOGraph("GO:0003680", GOMFPARENTS)
g2 <- oneGOGraph("GO:0003701", GOMFPARENTS)
g3 <- join(g1, g2)

g4 <- GOGraph(c("GO:0003680", "GO:0003701"), GOMFPARENTS)
if( require("Rgraphviz") && interactive() )
  plot(g3)
```

---

probeSetSummary

*Summarize Probe Sets Associated with a hyperGTest Result*

---

**Description**

Given the result of a hyperGTest run (an instance of GOHyperGResult), this function lists all Probe Set IDs associated with the selected Entrez IDs annotated at each significant GO term in the test result.

**Usage**

```
probeSetSummary(result, pvalue, categorySize, sigProbesets, ids = "ENTREZID")
```

**Arguments**

<code>result</code>	A <code>GOHyperGResult</code> instance. This is the output of the <code>hyperGTest</code> function when testing the GO category.
<code>pvalue</code>	Optional p-value cutoff. Only results for GO terms with a p-value less than the specified value will be returned. If omitted, <code>pvalueCutoff(result)</code> is used.
<code>categorySize</code>	Optional minimum size (number of annotations) for the GO terms. Only results for GO terms with <code>categorySize</code> or more annotations will be returned. If omitted, no category size criteria will be used.
<code>sigProbesets</code>	Optional vector of probeset IDs. See details for more information.
<code>ids</code>	Character. The type of IDs used in creating the <code>GOHyperGResult</code> object. Usually 'ENTREZID', but may be e.g., 'ACCNUM' if using <i>A. thaliana</i> chip.

**Details**

Usually the goal of doing a Fisher's exact test on a set of significant probesets is to find pathways or cellular activities that are being perturbed in an experiment. After doing the test, one usually gets a list of significant GO terms, and the next logical step might be to determine which probesets contributed to the significance of a certain term.

Because the input for the Fisher's exact test consists of a vector of unique Entrez Gene IDs, and there may be multiple probesets that interrogate a particular transcript, the output for this function lists all of the probesets that map to each Entrez Gene ID, along with an indicator that shows which of the probesets were used as input.

The rationale for this is that one might not be able to assume a given probeset actually interrogates the intended transcript, so it might be useful to be able to check to see what other similar probesets are doing.

Because one of the first steps before running `hyperGTest` is to subset the input vectors of `geneIds` and `universeGeneIds`, any information about probeset IDs that interrogate the same gene transcript is lost. In order to recover this information, one can pass a vector of probeset IDs that were considered significant. This vector will then be used to indicate which of the probesets that map to a given GO term were significant in the original analysis.

**Value**

A list of `data.frame`. Each element of the list corresponds to one of the GO terms (the term is provided as the name of the element). Each `data.frame` has three columns: the Entrez Gene ID (`EntrezID`), the probe set ID (`ProbeSetID`), and a 0/1 indicator of whether the probe set ID was provided as part of the initial input (`selected`)

Note that this 0/1 indicator will only be correct if the 'geneId' vector used to construct the `GOHyperGParams` object was a named vector (where the names are probeset IDs), or if a vector of 'sigProbesets' was passed to this function.

**Author(s)**

S. Falcon and J. MacDonald

**Examples**

```
## Fake up some data
library("hgu95av2.db")
library("annotate")
prbs <- ls(hgu95av2GO)[1:300]
```

```

## Only those with GO ids
hasGO <- sapply(mget(prbs, hgu95av2GO), function(ids)
if(!is.na(ids) && length(ids) > 1) TRUE else FALSE)
prbs <- prbs[hasGO]
prbs <- getEG(prbs, "hgu95av2")
## remove duplicates, but keep named vector
prbs <- prbs[!duplicated(prbs)]
## do the same for universe
univ <- ls(hgu95av2GO)[1:5000]
hasUnivGO <- sapply(mget(univ, hgu95av2GO), function(ids)
if(!is.na(ids) && length(ids) > 1) TRUE else FALSE)
univ <- univ[hasUnivGO]
univ <- unique(getEG(univ, "hgu95av2"))

p <- new("GOHyperGParams", geneIds=prbs, universeGeneIds=univ,
ontology="BP", annotation="hgu95av2", conditional=TRUE)
## this part takes time...
if(interactive()){
  hyp <- hyperGTest(p)
  ps <- probeSetSummary(hyp, 0.05, 10)
}

```

---

shortestPath

*Shortest Path Analysis*


---

## Description

The shortest path analysis was proposed by Zhou et. al. The basic computation is to find the shortest path in a supplied graph between two Entrez Gene IDs. Zhou et al claim that other genes annotated along that path are likely to have the same GO annotation as the two end points.

## Usage

```
shortestPath(g, GOnode, mapfun=NULL, chip=NULL)
```

## Arguments

<code>g</code>	An instance of the graph class.
<code>GOnode</code>	A length one character vector specifying the GO node of interest.
<code>mapfun</code>	A function taking a character vector of GO IDs as its only argument and returning a list of character vectors of Entrez Gene IDs annotated at each corresponding GO ID. The function should behave similarly to <code>mget(x, go2egmap, ifnotfound=NA)</code> , that is, NA should be returned if a specified GO ID has no Entrez ID mappings. See details for the interaction of <code>mapfun</code> and <code>chip</code> .
<code>chip</code>	The name of a DB-based annotation data package (the name will end in ".db"). This package will be used to generate an Entrez ID to GO ID mapping instead of <code>mapfun</code> .

## Details

The algorithm implemented here is quite simple. All Entrez Gene identifiers that are annotated at the GO node of interest are obtained. Those that are found as nodes in the graph are retained and used for the computation. For every pair of nodes at the GO term the shortest path between them is computed using `sp.between` from the RBGL package.

There is a presumption that the graph is undirected. This restriction could probably be lifted if there was some reason for it - a patch would be gratefully accepted.

The mapping of GO node to Entrez ID is achieved in one of three ways:

1. If `mapfun` is provided, it will be used to perform the needed lookups. In this case, `chip` will be ignored.
2. If `chip` is provided and `mapfun=NULL`, then the needed lookups will be done based on the GO to Entrez mappings encapsulated in the specified annotation data package. This is the recommended usage.
3. If `mapfun` and `chip` are `NULL` or missing, then the function will attempt to load the GO package (the environment-based package, distinct from `GO.db`). This package contains a legacy environment mapping GO IDs to Entrez IDs. If the GO package is not available, an error will be raised. Omitting both `mapfun` and `chip` is not recommended as it is not compatible with the DB-based annotation data packages.

## Value

The return values is a list with the following components:

<code>shortestpaths</code>	A list of the output from <code>sp.between</code> . The names are the names of the nodes used as the two endpoints
<code>nodesUsed</code>	A vector of the Entrez Gene IDs that were both found at the GO term of interest and were nodes in the supplied graph, <code>g</code> . These were used to compute the shortest paths.
<code>nodesNotUsed</code>	A vector of Entrez Gene IDs that were annotated at the GO term, but were not found in the graph <code>g</code> .

## Author(s)

R. Gentleman

## References

Transitive functional annotation by shortest-path analysis of gene expression data, by X. Zhou and M-C J. Kao and W. H. Wong, PNAS, 2002

## See Also

[sp.between](#)

## Examples

```
library("hgu95av2.db")
library("RBGL")

set.seed(321)
uniquen <- function(x) unique(unlist(x))
```



```

goid <- "GO:0005778"
egIds <- uniquemget(uniquemget(hgu95av2GO2PROBE[[goid]]),
                    hgu95av2ENTREZID))

v1 <- randomGraph(egIds, 1:10, .3, weights=FALSE)
## Since v1 is random, it might be disconnected and we need a
## connected graph to guarantee the existence of a path.
c1 <- connComp(v1)
largestComp <- c1[[which.max(sapply(c1, length))]]
v2 <- subGraph(largestComp, v1)

a1 <- shortestPath(v2, goid, chip="hgu95av2.db")

```

simLL

*Functions to compute similarities between GO graphs and also between Entrez Gene IDs based on their induced GO graphs.*

### Description

Both simUI and simLP compute a similarity measure between two GO graphs. For simLL, first the induced GO graph for each of its arguments is found and then these are passed to one of simUI or simLP.

### Usage

```

simLL(l11, l12, Ontology = "MF", measure = "LP", dropCodes = NULL,
      mapfun = NULL, chip = NULL)
simUI(g1, g2)
simLP(g1, g2)

```

### Arguments

l11	A Entrez Gene ID as a character vector.
l12	A Entrez Gene ID as a character vector.
Ontology	Which ontology to use ("MF", "BP", "CC").
measure	Which measure to use ("LP", "UI").
dropCodes	A set of evidence codes to be ignored in constructing the induced GO graphs.
mapfun	A function taking a character vector of Entrez Gene IDs as its only argument and returning a list of "GO lists" matching the structure of the lists in the GO maps of annotation data packages. The function should behave similarly to <code>mget(x, eg2gomap, ifnotfound=NA)</code> , that is, NA should be returned if a specified Entrez ID has no GO mapping. See details for the interaction of mapfun and chip.
chip	The name of a DB-based annotation data package (the name will end in ".db"). This package will be used to generate an Entrez ID to GO ID mapping instead of mapfun.
g1	An instance of the graph class.
g2	An instance of the graph class.

## Details

For each of 111 and 112 the set of most specific GO terms within the ontology specified (*Ontology*) that are not based on any excluded evidence code (*dropCodes*) are found. The mapping is achieved in one of three ways:

1. If *mapfun* is provided, it will be used to perform the needed lookups. In this case, *chip* will be ignored.
2. If *chip* is provided and *mapfun*=NULL, then the needed lookups will be done based on the Entrez to GO mappings encapsulated in the specified annotation data package. This is the recommended usage.
3. If *mapfun* and *chip* are NULL or missing, then the function will attempt to load the GO package (the environment-based package, distinct from *GO.db*). This package contains a legacy environment mapping Entrez IDs to GO IDs. If the GO package is not available, an error will be raised. Omitting both *mapfun* and *chip* is not recommended as it is not compatible with the DB-based annotation data packages.

Next, the induced GO graphs are computed.

Finally these graphs are passed to one of *simUI*, (union intersection), or *simLP* (longest path). For *simUI* the distance is the size of the intersection of the node sets divided by the size of the union of the node sets. Large values indicate more similarity. These similarities are between 0 and 1.

For *simLP* the length of the longest path in the intersection graph of the two supplied graph. Again, large values indicate more similarity. Similarities are between 0 and the maximum leaf depth of the graph for the specified ontology.

## Value

A list with:

<i>sim</i>	The numeric similarity measure.
<i>measure</i>	Which measure was used.
<i>g1</i>	The graph induced by 111.
<i>g2</i>	The graph induced by 112.

If one of the supplied Gene IDs does not have any GO terms associated with it, in the selected ontology and with the selected evidence codes then NA is returned.

## Author(s)

R. Gentleman

## See Also

[makeGOGraph](#)

## Examples

```
library("hgu95av2.db")
eg1 = c("9184", "3547")

bb = simLL(eg1[1], eg1[2], "BP", chip="hgu95av2.db")
```

termGraphs

*Extraction and Plotting of GO Terms from a GOHyperGResult Object***Description**

These functions extract and plot graph instances representing the relationships among GO terms tested using hyperGTest.

**Usage**

```
termGraphs(r, id = NULL, pvalue = NULL, use.terms = TRUE)
inducedTermGraph(r, id, children = TRUE, parents = TRUE)
plotGOTermGraph(g, r = NULL, add.counts = TRUE, max.nchar = 20,
  node.colors=c(sig="lightgray", not="white"),
  node.shape="plaintext", ...)
```

**Arguments**

r	A GOHyperGResult object as returned by hyperGTest when given a GOHyperGParams object as input.
id	A character vector of category IDs that specifies which terms should be included in the graph.
pvalue	Numeric p-value cutoff to use for selecting category terms to include. Will be ignored if id is present.
use.terms	Logical value indicating whether a "term" node attribute should be added to the returned graph providing the more descriptive, but possibly much longer, GO Terms.
children	A logical value indicating whether to include direct child terms of the terms specified by id.
parents	A logical value indicating whether to include direct parent terms of the terms specified by id.
g	A graph object as returned by inducedTermGraph or termGraphs.
add.counts	A logical value indicating whether category size counts should be added to the node labels when plotting.
max.nchar	The maximum character length for node labels in the plot.
node.colors	A named character vector of length two with components sig and not, giving color names for the significant and non-significant nodes, respectively.
node.shape	This argument controls the shape of the plotted nodes and must take on a value allowed by Rgraphviz.
...	For plotGOTermGraph, extra arguments are passed to the plot function.

**Details**

**termGraphs** returns a list of graph objects each representing one of the connected components of the subgraph of the GO ontology induced by selecting the specified GO IDs (if id is present) or by selecting the GO IDs that have a p-value less than pvalue. If use.terms is TRUE the GO IDs will be translated into GO Term names and attached to the nodes as node attributes (see nodeData). Edges in the graphs go from child (more specific) to parent (less specific).

**inducedTermGraph** returns a graph object representing the GO graph induced by the terms specified by `id`. The `children` and `parent` arguments control whether direct children and/or direct parents of the terms specified by `id` are added to the graph (at least one of the two must be TRUE).

**plotGOTermGraph** Create a plot using Rgraphviz of a graph object as returned by either `termGraphs` or `inducedTermGraph`. If a `GOHyperGResult` object is provided, then the nodes will be colored according to significance (based on the result object's `pvalueCutoff`) and counts will be added to show the size of the categories.

### Author(s)

Seth Falcon

---

triadCensus

*Triad Functions*

---

### Description

These functions provide some tools for finding triads in an undirected graph. A triad is a clique of size 3. The function `triadCensus` returns a list of all triads.

### Usage

```
triadCensus(graph)
isTriad(x, y, z, elz, ely)
reduce2Degreek(graph, k)
enumPairs(iVec)
```

### Arguments

<code>graph</code>	An instance of the graph class.
<code>k</code>	An integer indicating the minimum degree wanted.
<code>x</code>	A node
<code>y</code>	A node
<code>z</code>	A node
<code>elz</code>	The edgelist for z
<code>ely</code>	The edgelist for y
<code>iVec</code>	A vector of unique values

### Details

`enumPairs` takes a vector as input and returns a list of length `choose(length(iVec), 2)/2` containing all unordered pairs of elements.

`isTriad` takes three nodes as arguments. It is already known that `x` has edges to both `y` and `z` and we want to determine whether these are reciprocated. This is determined by examining `elz` for both `x` and `y` and then examining `ely` for both `x` and `z`.

`reduce2Degreek` is a function that takes an undirected graph as input and removes all nodes of degree less than `k`. This process is iterated until there are no nodes left (an error is thrown) or all

nodes remaining have degree at least  $k$ . The resultant subgraph is returned. It is used here because to be in a triad all nodes must have degree 2 or more.

`triadCensus` makes use of the helper functions described above and finds all triads in the graph.

**Value**

A list where each element is a triple indicating the members of the triad. Order is not important and all triads are reported in alphabetic order.

**Note**

See the graph package, `RBGL` and `Rgraphviz` for more details and alternatives.

**Author(s)**

R. Gentleman

**Examples**

```
##---- Should be DIRECTLY executable !! ----  
##-- ==> Define data, use random,
```

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