

Exploring the MAQC data with Bioconductor

VJ Carey

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1 Introduction

See the Sept 2006 issue of Nature Biotechnology for several articles about the MAQC initiative. The *MAQCsubset* package includes excerpts from the data published at GEO GSE5350.

```
> library(MAQCsubset)
> data(afxsubRMAES)
> afxsubRMAES
```

```
ExpressionSet (storageMode: lockedEnvironment)
assayData: 54675 features, 24 samples
  element names: exprs
phenoData
  sampleNames: AFX_1_A1.CEL, AFX_1_A2.CEL, ..., AFX_3_D2.CEL (24 total)
  varLabels and varMetadata:
    site: from cel
    samp: rna src/mixture code
    repl: replicate
featureData
  featureNames: 1007_s_at, 1053_at, ..., AFFX-TrpnX-M_at (54675 total)
  varLabels and varMetadata: none
experimentData: use 'experimentData(object)'
```

```
Annotation [1] "hgu133plus2"

> pd = pData(afxsubRMAES)
> table(pd$site, pd$samp)
```

```
  A B C D
1 2 2 2 2
2 2 2 2 2
3 2 2 2 2
```

Samples labeled "A" have 100% stratagene universal human RNA, while samples labeled "B" have 100% Ambion human brain RNA. Samples labeled C have .75A+.25B, and samples labeled D have .75B+.25A.

2 The proboscis plot

For Figure 2 of Shippy et al., *Using RNA sample titrations* (Nat Biotech, 24(9):1123-1131, Sep 2006), genes differentially expressed between samples A and B using t tests at $p = 0.001$ are identified. If, for such genes, the A samples are up-regulated relative to the B samples, then a self-consistent monotone titration (SCMT) is declared if the C samples for such genes are up-regulated relative to the D samples. For genes upregulated on B samples relative to A samples, then SCMT occurs if the D samples are up-regulated relative to the C samples.

Figure 2 of Shippy et al. plots the proportion of genes exhibiting SCMT against the intensity ratios (A/B or B/A as appropriate). These plots, formed for each manufacturer/normalization combination and for each site, have the appearance of long pointy noses and are thus called proboscis plots. The following code computes the necessary quantities:

```
> proboscis = function(es, site = 1, ABp = 0.001, CDp = 0.01, mmrad = 100) {
+   require(genefilter)
+   mcall = match.call()
+   mm = function(x, rad) {
+     start = ceiling(rad/2)
+     stop = floor(length(x) - (rad/2))
+     sapply(start:stop, function(i) mean(x[(i - floor(rad/2)):(i +
+       floor(rad/2))]))
+   }
+   ess = es[, es$site == site]
+   essab = ess[, ess$samp %in% c("A", "B")]
+   essab$samp = factor(essab$samp)
+   esscd = ess[, ess$samp %in% c("C", "D")]
+   esscd$samp = factor(esscd$samp)
+   tt = rowttests(exprs(essab), essab$samp)
+   L = which(tt$p < ABp & tt$dm < 0)
+   R = which(tt$p < ABp & tt$dm > 0)
+   ttcd = rowttests(exprs(esscd), esscd$samp)
+   ABL = tt$dm[L]
+   CDL = ttcd$dm[L]
+   ABR = tt$dm[R]
+   CDR = ttcd$dm[R]
+   NN = list(ttab = tt, ttcd = ttcd, ABL = sort(ABL), cdokL = 1 *

```

```

+      (CDL < 0)[order(ABL)], ABR = sort(ABR), dcokR = 1 * (CDR >
+      0)[order(ABR)])
+    `A-B` = c(ONR <- mm(NN$ABL, mrad), mm(NN$ABR, mrad))
+    `P(SCMT|A-B)` = c(mm(NN$cdokL, mrad), mm(NN$dcokR, mrad))
+    new("proboStruct", call = mcall, list("A-B" = `A-B`, "P(SCMT|A-B)" = `P(SCMT|A-B)`,
+    leftinds = 1:length(ONR)))
+ }
> NN1 = proboscis(afxsubRMAES)
> NN2 = proboscis(afxsubRMAES, site = 2)
> NN3 = proboscis(afxsubRMAES, site = 3)

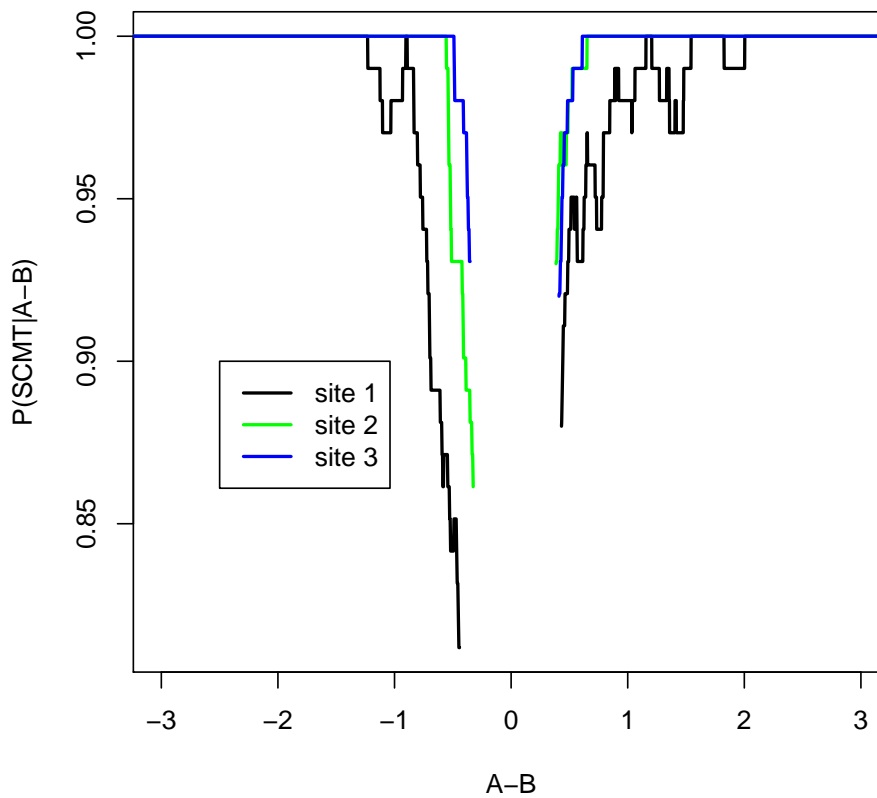
```

There are simple graphical methods:

```

> plot(NN1, lwd = 2)
> lines(NN2, col = "green", lwd = 2)
> lines(NN3, col = "blue", lwd = 2)
> legend(-2.5, 0.9, lty = 1, lwd = 2, legend = c("site 1", "site 2",
+ "site 3"), col = c("black", "green", "blue"))

```



These do not look

exactly like the plots in the Shippy paper, presumably because only two replicates per site are in use in this display.