

# BioC Introduction

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- 1 Getting Acquainted with Bioconductor
- 2 The ALL Dataset and ExpressionSet
- 3 BioC Introduction
- 4 Summary
- 5 Exercise

# Outline

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# Preparation

- Get acquainted with the Bioconductor website

- biocView packages

- <http://bioconductor.org/download>

- Getting help: mailing list

- <http://www.bioconductor.org/docs/mailList.html>

- Searchable mailing list

- <http://dir.gmane.org/gmane.science.biology.informatics.bioconductor>

- Easy approach to install packages

```
source("http://bioconductor.org/biocLite.R")
```

```
biocLite()
```

```
biocLite(pkgs)
```

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# The ALL ExpressionSet

## Code

```
> library(ALL)
> data(ALL)
> ALL
```

```
ExpressionSet (storageMode: lockedEnvironment)
assayData: 12625 features, 128 samples
  element names: exprs
phenoData
  sampleNames: 01005, 01010, ..., LAL4 (128 total)
  varLabels and varMetadata description:
    cod: Patient ID
    diagnosis: Date of diagnosis
    ...: ...
    date last seen: date patient was la
st seen
  (21 total)
featureData
  featureNames: 1000_at, 1001_at, ..., A
  FFX-YELO24w/RIP1_at (12625 total)
  fvarLabels and fvarMetadata description: none
experimentData: use 'experimentData(object)'
  pubMedIds: 14684422 16243790
Annotation: hgu95av2
```

# ExpressionSet

## Structure for genomic data

- `assayData`: Expression data from microarray experiments.
  - > `exprs(ALL)`
- `metadata`: `phenoData`, `featureData`, `annotation` – A description of the samples and features in experiment.
  - > `phenoData(ALL)`
  - > `sampleNames(ALL)`
  - > `featureData(ALL)`
  - > `head(featureNames(ALL))`
  - > `annotation(ALL)`
- `experimentData`: A flexible structure to describe experiment.
  - > `experimentData(ALL)`
  - > `abstract(ALL)`
- `protocoldata`: Equipment-generated variables describing sample phenotypes.

## Some operations on ExpressionSet

### Code

```
> class(ALL)

[1] "ExpressionSet"
attr("package")
[1] "Biobase"

> dim(ALL)

Features  Samples
  12625     128

> exprs(ALL)[1:3, 1:3]

           01005    01010    03002
1000_at    7.597323  7.479445  7.567593
1001_at    5.046194  4.932537  4.799294
1002_f_at  3.900466  4.208155  3.886169

> names(pData(ALL))
> varMetadata(ALL)[1:5, ,drop=FALSE]
> colnames(exprs(ALL))
> table(ALL$BT) # dollar-sign returns phenodata selection
```

# Some operations on ExpressionSet

## Exercise

- 1 Get familiar with the generic functions to access the phenotypical data and meta-data associated with ALL.
- 2 Use `pData`, `varLabels` and `VarMatedata` to extract details of phenotype information of ALL.
- 3 Try to find covariates carrying the information of the molecular biology and cell types (B- and T-cells) of the ALL samples.

## Data subsetting

Select samples originating from B-cell tumors (BT covariate) found to carry out BCR/ABL mutation and NEG with no cytogenetic abnormalities (mol.biol covariate).

### Code: subsetting

```
> bcell <- grep("^B", as.character(ALL$BT))
> types <- c("NEG", "BCR/ABL")
> moltyp <- which(as.character(ALL$mol.biol) %in% types)
> ALL_bcrneg <- ALL[, intersect(bcell, moltyp)]
```

### Code: reshaping

```
> ALL_bcrneg$BT <- factor(ALL_bcrneg$BT)
> ALL_bcrneg$mol.biol <- factor(ALL_bcrneg$mol.biol)
```

## Nonspecific filtering

`nsFilter` – filter out probe sets for a number of different criteria.

### code: `nsFilter`

```
> library("genefilter")
> library("hgu95av2.db")
> #openVignette("genefilter")
> filt_bcrneg <- nsFilter(ALL_bcrneg,
+                         require.entrez=TRUE,
+                         require.GOBP=TRUE,
+                         remove.dupEntrez=TRUE,
+                         feature.exclude="~AFFX",
+                         var.cutoff=0.5)
> ALLfilt_bcrneg <- filt_bcrneg$eset
> dim(ALLfilt_bcrneg)
```

Features	Samples
3842	79

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## Finding help in R

- `? foo` gets the manual page of function `foo`.
- `class ? foo` gets manual page of class `foo`.
- `help.start(foo)` gets html manual page of object `foo`.
- `openVignette()` provides interface for opening vignettes. Note that this function is in namespace of package *Biobase*.
- `apropos` finds objects in the search path that partially match the given character string.
- `sessionInfo()` prints version information of R and loaded packages.
- `search()` gives a list of attached packages in current working R session.

## Finding help in R

### Exercise:

- 1 There are many number of different plotting functions available. Can you find them?
- 2 Try to find function to use to perform a MannWhitney test.
- 3 Open the PDF version of the vignette “Bioconductor Overview” which is part of the *Biobase* package. Use either `biocLite()` or `install.packages()`.
- 4 What is the output of function `sessionInfo()`?
- 5 Try to install the *xtable* packages.

## Annotation mapping

*hgu95av2.db*: mappings between Affymetriex IDs and various forms of biological annotation.

```
> hgu95av2()  
> ls("package:hgu95av2.db")
```

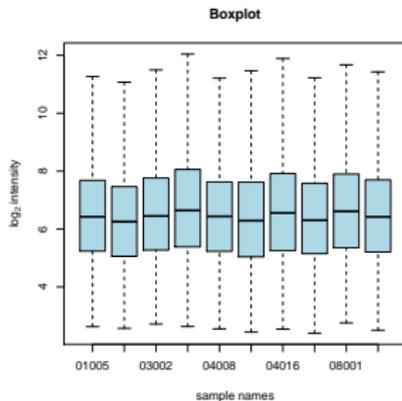
### Code: mapping

```
> hgu95av2SYMBOL$"1001_at"  
  
[1] "TIE1"  
  
> mget("1001_at", hgu95av2SYMBOL)  
  
$`1001_at`  
[1] "TIE1"  
  
> rmap <- revmap(hgu95av2SYMBOL) ## reverse mapping  
> get("TIE1", rmap)  
  
[1] "1001_at"
```

# Graphics

## Code: visualizing expression patterns

```
> apropos("plot")  
> x <- exprs(ALLfilt_bcrneg)[, 1]  
> y <- exprs(ALLfilt_bcrneg)[, 2]  
> plot(x=x, y=y)  
> smoothScatter(x=x, y=y)  
> boxplot(exprs(ALLfilt_bcrneg)[, 1:10])
```



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# Summary

- Logistics of access.
  - Install packages using `biocLite()`.
  - Load packages into the session using `library()`.
- ExpressionSet objects.
  - Fundamental facilities: `exprs()`, `$`.
  - Others: `phenoData()`, `featureData()`, `varLabels()`, `annotation()`.
- Annotation mapping and remapping.
  - Fundamental facilities: `get()`, `mget()`, and `revmap()`.
  - Annotation packages for certain platform `*platform*.db`.
- Visualization: `boxplot()`, `heatmap()`.
- Session information: `sessionInfo()`.

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## Exercise

- `hgu95avMAP` *environment* contains the mappings between affymetrix identifiers and chromosome band locations.
- `apply` family of functions: `apply()`, `sapply()`, `lapply()`, and `eapply()`.

### `eapply`

- 1 Find the chromosome band to which the probe `1001_at` maps.
- 2 Find all genes that map to the p arm of chromosome 17 (17p) using functions `grep` and `eapply`.