

# Package ‘APalyzer’

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**Type** Package

**Title** A toolkit for APA analysis using RNA-seq data

**Version** 1.21.0

**Description** Perform 3'UTR APA, Intronic APA and gene expression analysis using RNA-seq data.

**biocViews** Sequencing, RNASeq, DifferentialExpression, GeneExpression,  
GeneRegulation, Annotation, DataImport, Software

**Imports** GenomicRanges, GenomicFeatures, GenomicAlignments, DESeq2,  
ggrepel, SummarizedExperiment, Rsubread, stats, ggplot2,  
methods, rtracklayer, VariantAnnotation, dplyr, tidyr, repmis,  
Rsamtools, HybridMTest

**Suggests** knitr, rmarkdown, BiocStyle, org.Mm.eg.db, AnnotationDbi,  
TBX20BamSubset, testthat, pasillaBamSubset

**URL** <https://github.com/RJWANGbioinfo/APalyzer/>

**BugReports** <https://github.com/RJWANGbioinfo/APalyzer/issues>

**VignetteBuilder** knitr

**License** LGPL-3

**Encoding** UTF-8

**Depends** R (>= 3.5.0)

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|        |                                     |
|--------|-------------------------------------|
| APABox | <i>APABox, APA RED Box plotting</i> |
|--------|-------------------------------------|

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## Description

APA RED Box plotting

## Usage

```
APABox (df, xlab = "APAre", ylab = "RED",
        plot_title = NULL)
```

## Arguments

|            |                                     |
|------------|-------------------------------------|
| df         | a dataframe of APAdiff output       |
| xlab       | lable of x-axis, default is 'APAre' |
| ylab       | lable of y-axis, default is 'RED'   |
| plot_title | Main title of plot                  |

## Value

The function APABox return a Box plot.

## Author(s)

Ruijia Wang

**Examples**

```

library("TBX20BamSubset")
library("Rsamtools")
flsall = getBamFileList()
extpath = system.file("extdata",
"mm9_TBX20.APAout.RData", package="APALyzer")
load(extpath)
sampleTable1 = data.frame(samplename = c(names(flsall)),
condition = c(rep("NT",3),rep("KD",3)))
sampleTable2 = data.frame(samplename = c("SRR316184","SRR316187"),
condition = c("NT","KD"))
## 3'UTR APA plot
test_3UTRmuti=APAdiff(sampleTable1,DFUTRraw,
conKET='NT',trtKEY='KD',PAS='3UTR',CUTreads=0)
UTR_APA_PLOTBOX=APABox(test_3UTRmuti, plot_title='3UTR APA')

## IPA plot
test_IPAmuti=APAdiff(sampleTable1,IPA_OUTraw,
conKET='NT',trtKEY='KD',PAS='IPA',CUTreads=0)
IPA_PLOTBOX=APABox(test_IPAmuti, plot_title='IPA')

```

APAdiff

*APAdiff, calculate delta relative expression (RED) and statistics significance between two sample groups*

**Description**

Calculate delta relative expression (RED) and statistics significance between two sample groups.

**Usage**

```

APAdiff(sampleTable,mutiraw, conKET='NT',
trtKEY='KD', PAS='3UTR', CUTreads=0, p_adjust_methods="fdr", MultiTest='unpaired t-test')

```

**Arguments**

|             |   |
|-------------|---|
| sampleTable | a dataframe of sample table containing 8 columns for Intronic PASs: 'sample-name','condition' |
| mutiraw     | a dataframe output obtained using either PASEXP_3UTR or PASEXP_IPA                            |
| conKET      | the name of control in the sampletable, default is 'NT'                                       |
| trtKEY      | the name of control in the sampletable, default is 'KD'                                       |
| PAS         | type of PAS analyzed, either '3UTR' or 'IPA', default is '3UTR'                               |
| CUTreads    | reads cutoff used for the analysis, default is 0  |

`p_adjust_methods` p value correction method, the method can be "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none", default is "fdr"

`MultiTest` statistics testing method for muti-replicates designs, the method can be "unpaired t-test", "paired t-test", "ANOVA", default is "unpaired t-test"

### Value

The function `APAdiff` return a dataframe containing RED, pvalue and regulation pattern (UP, DN or NC) for either each gene (3'UTR APA) or each PAS (IPA).

### Author(s)

Ruijia Wang

### Examples

```
library("TBX20BamSubset")
library("Rsamtools")
flsall = getBamFileList()
extpath = system.file("extdata",
  "mm9_TBX20.APAout.RData", package="APALyzer")
load(extpath)
sampleTable1 = data.frame(samplename = c(names(flsall)),
  condition = c(rep("NT",3),rep("KD",3)))
sampleTable2 = data.frame(samplename = c("SRR316184","SRR316187"),
  condition = c("NT","KD"))
## Analysis 3'UTR APA between KD and NT group using muti-replicates
test_3UTRmuti=APAdiff(sampleTable1,DFUTRaw,
conKET='NT',trtKEY='KD',PAS='3UTR',CUTreads=0,p_adjust_methods="fdr",MultiTest='unpaired t-test')

## Analysis 3'UTR APA between KD and NT group without replicates
test_3UTRsing=APAdiff(sampleTable2,DFUTRaw,
conKET='NT',trtKEY='KD',PAS='3UTR',CUTreads=0,p_adjust_methods="fdr")

## Analysis IPA between KD and NT group
test_IPAmuti=APAdiff(sampleTable1,IPA_OUTraw,
conKET='NT',trtKEY='KD',PAS='IPA',CUTreads=0,p_adjust_methods="fdr",MultiTest='unpaired t-test')

## Analysis IPA between KD and NT group without replicates
test_IPAsing=APAdiff(sampleTable2,IPA_OUTraw,
conKET='NT',trtKEY='KD',PAS='IPA',CUTreads=0,p_adjust_methods="fdr")
```

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APAVolcano

*APAVolcano, APA Volcano plotting*

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### Description

APA Volcano plotting

**Usage**

```
APAVolcano (df, Pcol = "pvalue", PAS='3UTR',
            top = -1, markergenes = NULL,
            y_cutoff = 0.05, xlab = "RED", ylab = "-Log10(P-value)",
            PAScolor = c("gray80", "red", "blue"),
            alpha = 0.75, plot_title = NULL,
            width = 4, height = 2.5)
```

**Arguments**

|             |   |
|-------------|---|
| df          | a dataframe of APAdiff output   |
| Pcol        | p-value column used to for y-axis of volcano plot, default is 'pvalue'  |
| top         | number of genes/IPA to label in the plot, default is -1, which don't lable top genes, user can set it >0, e.g., top = 5 |
| markergenes | a set of genes to label in the plot   |
| PAS         | type of PAS analyzed, either '3UTR' or 'IPA', default is '3UTR'   |
| y_cutoff    | y cutoff line, default is 0.05  |
| xlab        | lable of x-axis, default is 'RED'   |
| ylab        | lable of y-axis, default is '-Log10(P-value)'   |
| PAScolor    | dot color for 'NC', 'UP' and 'DN' gene/IPAs, default is "gray80", "red", and "blue"                                     |
| alpha       | alpha of the dot, default is 0.75   |
| plot_title  | Main title of plot  |
| width       | width of the dot, default is 4  |
| height      | height of the dot, default is 2.5   |

**Value**

The function APAVolcano return a Volcano plot.

**Author(s)**

Ruijia Wang

**Examples**

```
library("TBX20BamSubset")
library("Rsamtools")
flsall = getBamFileList()
extpath = system.file("extdata",
                      "mm9_TBX20.APAout.RData", package="APALyzer")
load(extpath)
sampleTable1 = data.frame(samplename = c(names(flsall)),
                          condition = c(rep("NT",3),rep("KD",3)))
sampleTable2 = data.frame(samplename = c("SRR316184","SRR316187"),
                          condition = c("NT","KD"))
```

```

## 3'UTR APA plot
test_3UTRmuti=APAdiff(sampleTable1,DFUTRraw,
conKET='NT',trtKEY='KD',PAS='3UTR',CUTreads=0)
UTR_APA_PLOT=APAVolcano(test_3UTRmuti, PAS='3UTR', Pcol = "pvalue", top=5, plot_title='3UTR APA')

## IPA plot
test_IPAmuti=APAdiff(sampleTable1,IPA_OUTraw,
conKET='NT',trtKEY='KD',PAS='IPA',CUTreads=0)
IPA_PLOT=APAVolcano(test_IPAmuti, PAS='IPA', Pcol = "pvalue", top=5, plot_title='IPA')

```

---

|                  |   |
|------------------|---|
| download_testbam | <i>download_testbam, download bam files of mouse testis and heart</i> |
|------------------|---|

---

### Description

download bam files of mouse testis and heart

### Usage

```
download_testbam()
```

### Value

The function download\_testbam download test data bam files.

### Author(s)

Ruijia Wang

### Examples

```
download_testbam()
```

---

|            |  |
|------------|--|
| GENEXP_CDS | <i>GENEXP_CDS, count reads mapped to CDS regions and calculate TPM for coding gene</i> |
|------------|--|

---

### Description

Map reads to CDS regions and calculate TPM for each gene.

### Usage

```
GENEXP_CDS(CDSbygene, f1S, Strandtype="NONE")
```

**Arguments**

|            |  |
|------------|--|
| CDSbygene  | a genomic ranges of CDS regions for each coding gene   |
| fls        | bamfile lists containing the file and path of bam files  |
| Strandtype | strand type of the bam file; "forward" is forward sequencing, "invert" is reverse sequencing and "NONE" is non-strand specific, Default is "NONE". |

**Value**

The function GENEXP\_CDS() return a dataframe containing reads count, TPM for each gene

**Author(s)**

Ruijia Wang

**Examples**

```
## count reads mapped to CDS regions and calculate TPM for each gene
## using forward sequencing
library("TBX20BamSubset")
library("Rsamtools")
library("GenomicAlignments")
library("GenomicFeatures")
library("org.Mm.eg.db")
flsall = getBamFileList()
extpath = system.file("extdata", "mm9.chr19.refGene.R.DB", package="APalyzer")
txdb = loadDb(extpath, packageName='GenomicFeatures')
IDDB = org.Mm.eg.db
CDSdbraw = REFCDS(txdb, IDDB)
DFGENErw = GENEXP_CDS(CDSdbraw, flsall, Strandtype="forward")
```

---

PAS2GEF

*PAS2GEF, build reference regions for 3'UTR PASs*

---

**Description**

Build 3'UTR PAS and IPA (IPA and LE) Reference using GTF file.

**Usage**

```
PAS2GEF(GTFfile, AnnoMethod="V2")
```

**Arguments**

|            |   |
|------------|---|
| GTFfile    | GTF file of gene annotation   |
| AnnoMethod | annotation method used to build PAS reference, either 'legacy' or 'V2', default is 'V2' |

**Value**

The function PAS2GEF() returns 3 input tables of PAS references: PASREF\$refUTRraw is for 3'UTR PAS, PASREF\$dfIPA and PASREF\$dfLE are for IPA references.

**Author(s)**

Ruijia Wang

**Examples**

```
## build Reference ranges for 3'UTR PASs in mouse
download.file(url='ftp://ftp.ensembl.org/pub/release-99/gtf/mus_musculus/Mus_musculus.GRCm38.99.gtf.gz',
             destfile='Mus_musculus.GRCm38.99.gtf.gz')
GTFfile="Mus_musculus.GRCm38.99.gtf.gz"

PASREF=PAS2GEF(GTFfile, AnnoMethod="V2")
refUTRraw=PASREF$refUTRraw
dfIPA=PASREF$dfIPA
dfLE=PASREF$dfLE
```

---

PASEXP\_3UTR

*PASEXP\_3UTR, calculate relative expression of aUTR and cUTR regions*

---

**Description**

Map reads to 3'UTR APA regions and calculate relative expression of aUTR and cUTR regions.

**Usage**

```
PASEXP_3UTR(UTRdb, f1S, Strandtype="NONE")
```

**Arguments**

|            |  |
|------------|--|
| UTRdb      | a genomic ranges of aUTR(pPAS to dPAS) and cUTR(cdsend to pPAS) regions for each gene  |
| f1S        | bamfile lists containing the file and path of bam files  |
| Strandtype | strand type of the bam file; "forward" is forward sequencing, "invert" is reverse sequencing and "NONE" is non-strand specific, Default is "NONE". |

**Value**

The function PASEXP\_3UTR() return a dataframe containing reads count, RPKM and relative expression of aUTR and cUTR for each gene



**Author(s)**

Ruijia Wang

**Examples**

```
## count reads mapped to 3'UTR APA regions and
## calculate relative expression of aUTR and cUTR regions
## using forward sequencing
library("TBX20BamSubset")
library("Rsamtools")
library("GenomicAlignments")
library("repmis")
flsall = getBamFileList()
URL="https://github.com/RJWANGbioinfo/PAS_reference_RData/blob/master/"
file="mm9_REF.RData"
source_data(paste0(URL,file,"?raw=True"))
refUTRraw = refUTRraw[which(refUTRraw$Chrom=="chr19"),]
UTRdbraw = REF3UTR(refUTRraw)
DFUTRraw = PASEXP_3UTR(UTRdbraw, flsall, Strandtype="forward")
```

PASEXP\_IPA

*PASEXP\_IPA, calculate relative expression of IPA regions***Description**

Map reads to IPA regions and calculate relative expression of aUTR and cUTR regions.

**Usage**

```
PASEXP_IPA(dfIPArw, dfLErwh, fls, Strandtype="NONE", nts=1, minMQS=0, SeqType = "SingleEnd")
```

**Arguments**

|            |  |
|------------|--|
| dfIPArw    | a dataframe containing 8 columns for Intronic PASs: 'PASid', 'gene_symbol', 'Chrom', 'Strand', 'Pos', 'upstreamSS', 'downstreamSS'. 'upstreamSS' means closest 5'/3' splice site to IPA, 'downstreamSS' means closest 3' splice site |
| dfLErwh    | a dataframe containing 5 columns for 3' least exon: 'gene_symbol', 'Chrom', 'Strand', 'LEstart', 'TES'. 'LEstart' means the start position of last 3' exon.  |
| fls        | bamfile lists containing the file and path of bam files  |
| Strandtype | strand type of the bam file; "forward" is forward sequencing, "invert" is reverse sequencing and "NONE" is non-strand specific, Default is "NONE".   |
| nts        | number of threads used for computing, parameter used by <a href="#">featureCounts</a> , nthread option, Default is 1   |
| minMQS     | minimum mapping quality score of counted reads, parameter used by <a href="#">featureCounts</a> , minMQS option, Default is 0  |
| SeqType    | set the sequencing type of reads in bam files can be either 'SingleEnd' (default) or 'ThreeMostPairEnd'.   |

**Value**

The function PASEXP\_IPA() return a dataframe containning reads count, RPKM and relative expression of aUTR and cUTR for each gene

**Author(s)**

Ruijia Wang

**Examples**

```
## count reads mapped to IPA regions and
## calculte relative expression of aUTR and cUTR regions
## using forward sequencing
library("TBX20BamSubset")
library("Rsamtools")
library("GenomicAlignments")
library("repmis")
flsall = getBamFileList()
URL="https://github.com/RJWANGbioinfo/PAS_reference_RData/blob/master/"
file="mm9_REF.RData"
source_data(paste0(URL,file,"?raw=True"))
IPA_OUTraw=PASEXP_IPA(dfIPA, dfLE, flsall, Strandtype="forward", nts=1)
```

---

REF3UTR

*REF3UTR, build reference regions for 3'UTR PASs*

---

**Description**

Build 3'UTR PAS Reference for distal and proximal PAS.

**Usage**

```
REF3UTR(refUTR)
```

**Arguments**

refUTR            a dataframe containing 6 colmuns for 3'UTR PASs: 'gene\_symbol', 'Chrom', 'Strand', 'Proximal', 'Distal', 'cdsend'

**Value**

The function REF3UTR() returns a genomic ranges of aUTR(pPAS to dPAS) and cUTR(cdsend to pPAS) regions for each gene

**Author(s)**

Ruijia Wang

**Examples**

```
## build Reference ranges for 3'UTR PASs in mouse
library(repmis)
URL="https://github.com/RJWANGbioinfo/PAS_reference_RData/blob/master/"
file="mm9_REF.RData"
source_data(paste0(URL,file,"?raw=True"))
refUTRraw=refUTRraw[which(refUTRraw$Chrom=='chr19'),]
UTRdbraw=REF3UTR(refUTRraw)
```

---

|         |  |
|---------|--|
| REF4PAS | <i>REF4PAS, build reference regions for 3'UTR and Intronic PAS using dataframe formatted input</i> |
|---------|--|

---

**Description**

build reference regions for 3'UTR and Intronic PAS using dataframe formatted input

**Usage**

```
REF4PAS(refUTRraw, dfIPArw, dfLEraw)
```

**Arguments**

|           |  |
|-----------|--|
| refUTRraw | a dataframe containing 6 columns for 3'UTR PASs: 'gene_symbol', 'Chrom', 'Strand', 'Proximal', 'Distal', 'cdsend'  |
| dfIPArw   | a dataframe containing 8 columns for Intronic PASs: 'PASid', 'gene_symbol', 'Chrom', 'Strand', 'Pos', 'upstreamSS', 'downstreamSS'. 'upstreamSS' means closest 5'/3' splice site to IPA, 'downstreamSS' means closest 3' splice site |
| dfLEraw   | a dataframe containing 5 columns for 3' least exon: 'gene_symbol', 'Chrom', 'Strand', 'LEstart', 'TES'. 'LEstart' means the start position of last 3' exon.  |

**Value**

The function REF4PAS() returns list a genomic ranges of 3'UTR, Intronic PAS and last 3'exon regions for each gene

**Author(s)**

Ruijia Wang

**Examples**

```
## build Reference ranges for 3'UTR and Intronic PAS in mouse (mm9)
library(repmis)
URL="https://github.com/RJWANGbioinfo/PAS_reference_RData/blob/master/"
file="mm9_REF.RData"
source_data(paste0(URL,file,"?raw=True"))
refUTRraw=refUTRraw[which(refUTRraw$Chrom=='chr19'),]
```

```
dfIPAraw=dfIPA[which(dfIPA$Chrom=='chr19'),]
dfLEraw=dfLE[which(dfLE$Chrom=='chr19'),]
PASREF=REF4PAS(refUTRraw,dfIPAraw,dfLEraw)
UTRdbraw=PASREF$UTRdbraw
dfIPA=PASREF$dfIPA
dfLE=PASREF$dfLE
```

REFCDS

*REFCDS, build reference regions for CDS of protein coding genes***Description**

Build CDS reference for protein coding genes.

**Usage**

```
REFCDS(txdb, IDDB)
```

**Arguments**

|      |  |
|------|--|
| txdb | a TranscriptDb generate using GenomicFeatures                        |
| IDDB | Genome annotation of the corresponding species, e.g., "org.Hs.eg.db" |

**Value**

The function REFCDS() returns a genomic ranges of CDS regions for each coding gene

**Author(s)**

Ruijia Wang

**Examples**

```
## build Reference ranges for CDS in mouse coding genes
library("GenomicFeatures")
library("org.Mm.eg.db")
extpath = system.file("extdata", "mm9.chr19.refGene.R.DB", package="APalyzer")
txdb = loadDb(extpath, packageName='GenomicFeatures')
IDDB = org.Mm.eg.db
CDSdbraw = REFCDS(txdb, IDDB)
```

---

|                  |  |
|------------------|--|
| ThreeMostPairBam | <i>ThreeMostPairBam, extract 3 prime most alignment of a paired-end bam file</i> |
|------------------|--|

---

**Description**

extract 3 prime most alignment of a paired-end bam file and saved into a new bam file.

**Usage**

```
ThreeMostPairBam(BamfilePath, OutDirPath, StrandType="NONE")
```

**Arguments**

|             |  |
|-------------|--|
| BamfilePath | file path of a bam file  |
| OutDirPath  | output folder path   |
| StrandType  | strand type of the bam file; "forward-reverse": read 1 forward but read 2 is reverse sequencing, "reverse-forward": read 2 forward but read 1 is reverse sequencing, and "NONE" is non-strand specific, Default is "NONE". |

**Value**

The function `ThreeMostPairBam()` return a single-end bam file containing 3 prime most alignment of the input paired-end file

**Author(s)**

Ruijia Wang

**Examples**

```
## Extract 3 prime most alignment of a paired-end
## bam file and saved into a new bam file
library("pasillaBamSubset")

ThreeMostPairBam (BamfilePath=untreated3_chr4(),
                  OutDirPath=getwd(),
                  StrandType='forward-reverse')
```

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