

# Package ‘DAPAR’

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**Type** Package

**Title** Tools for the Differential Analysis of Proteins Abundance with R

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**Description** This package contains a collection of functions for the visualisation and the statistical analysis of proteomic data.

**License** Artistic-2.0

**VignetteBuilder** knitr

**Depends** R (>= 3.5)

**Suggests** BiocGenerics, Biobase, testthat, BiocStyle, Prostar

**Imports** MSnbase, RColorBrewer,stats,preprocessCore,Cairo,png,  
lattice,reshape2,gplots,pcaMethods,ggplot2,  
limma,knitr,tmvtnorm,norm,impute, doParallel, stringr,  
parallel, foreach,grDevices, graphics, openxlsx, utils, cp4p  
(>= 0.3.5), scales, Matrix, vioplot, imp4p (>= 0.5),  
highcharter (>= 0.5.0), DAPARdata (>= 1.10.2), siggenes, graph,  
lme4, readxl, clusterProfiler, dplyr, tidyr,AnnotationDbi,  
tidyverse, imputeLCMD

**biocViews** Proteomics, Normalization, Preprocessing, MassSpectrometry,  
QualityControl, GO, DataImport

**NeedsCompilation** no

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---

|                  |  |
|------------------|--|
| addOriginOfValue | <i>Sets the OriginOfValues dataframe</i> |
|------------------|--|

---

**Description**

Sets the OriginOfValues dataframe in the fData table

**Usage**

```
addOriginOfValue(obj, index = NULL)
```

**Arguments**

|       |                            |
|-------|----------------------------|
| obj   | An object of class MSnSet  |
| index | A list of integer xxxxxxxx |

**Value**

An instance of class MSnSet.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
addOriginOfValue(Exp1_R25_pept)
```

---

|                    |  |
|--------------------|--|
| barplotEnrichGO_HC | <i>A barplot that shows the result of a GO enrichment, using the package highcharter</i> |
|--------------------|--|

---

**Description**

A barplot of GO enrichment analysis

**Usage**

```
barplotEnrichGO_HC(ego, maxRes = 5, title = NULL)
```

**Arguments**

|        |  |
|--------|--|
| ego    | The result of the GO enrichment, provides either by the function enrichGO in the package DAPAR or the function enrichGO of the package <a href="#">clusterProfiler</a> |
| maxRes | The maximum number of categories to display in the plot  |
| title  | The title of the plot  |

**Value**

A barplot

**Author(s)**

Samuel Wieczorek

---

|                   |   |
|-------------------|---|
| barplotGroupGO_HC | <i>A barplot which shows the result of a GO classification, using the package highcharter</i> |
|-------------------|---|

---

**Description**

A barplot of GO classification analysis

**Usage**

```
barplotGroupGO_HC(ggo, maxRes = 5, title = "")
```

**Arguments**

|        |   |
|--------|---|
| ggo    | The result of the GO classification, provides either by the function group_GO in the package DAPAR or the function groupGO in the package <a href="#">clusterProfiler</a> |
| maxRes | An integer which is the maximum number of classes to display in the plot  |
| title  | The title of the plot   |

**Value**

A barplot

**Author(s)**

Samuel Wieczorek

---

|          |  |
|----------|--|
| boxPlotD | <i>Builds a boxplot from a dataframe</i> |
|----------|--|

---

**Description**

Boxplot for quantitative proteomics data

**Usage**

```
boxPlotD(qData, dataForXAxis = NULL, labels = NULL,
group2Color = "Condition")
```

**Arguments**

|              |   |
|--------------|---|
| qData        | A dataframe that contains quantitative data.  |
| dataForXAxis | A vector containing the types of replicates to use as X-axis. Available values are: Label, Bio.Rep, Bio.Rep and Tech.Rep. Default is "Label".                                   |
| labels       | A vector of the conditions (labels) (one label per sample).   |
| group2Color  | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A boxplot

**Author(s)**

Florence Combes, Samuel Wiczorek

**See Also**

[densityPlotD](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
types <- c("Label", "Bio.Rep")
dataForXAxis <- Biobase::pData(Exp1_R25_pept)[, types]
labels <- Biobase::pData(Exp1_R25_pept)[, "Label"]
boxPlotD(qData, dataForXAxis, labels)
```

---

boxPlotD\_HC

*Builds a boxplot from a dataframe using the library highcharter*

---

**Description**

Boxplot for quantitative proteomics data using the library highcharter

**Usage**

```
boxPlotD_HC(qData, dataForXAxis = "Label", labels = NULL,
            group2Color = "Condition")
```

**Arguments**

|              |   |
|--------------|---|
| qData        | A dataframe that contains quantitative data.  |
| dataForXAxis | A vector containing the types of replicates to use as X-axis. Available values are: Label, Bio.Rep, Bio.Rep and Tech.Rep. Default is "Label".                                   |
| labels       | A vector of the conditions (labels) (one label per sample).   |
| group2Color  | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A boxplot

**Author(s)**

Samuel Wieczorek

**See Also**

[densityPlotD\\_HC](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
types <- c("Label", "Bio.Rep")
dataForXAxis <- Biobase::pData(Exp1_R25_pept)[, types]
labels <- Biobase::pData(Exp1_R25_pept)[, "Label"]
boxPlotD_HC(qData, dataForXAxis, labels)
```

---

BuildAdjacencyMatrix *Function matrix of appartenance group*

---

**Description**

Method to create a binary matrix with proteins in columns and peptides in lines on a MSnSet object (peptides)

**Usage**

```
BuildAdjacencyMatrix(obj.pep, protID, unique = TRUE)
```

**Arguments**

|         |   |
|---------|---|
| obj.pep | An object (peptides) of class MSnSet.   |
| protID  | The name of proteins ID column  |
| unique  | A boolean to indicate whether only the unique peptides must be considered (TRUE) or if the shared peptides have to be integrated (FALSE). |

**Value**

A binary matrix

**Author(s)**

Florence Combes, Samuel Wieczorek, Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], "Protein.group.IDs", TRUE)
```



---

BuildColumnToProteinDataset

*creates a column for the protein dataset after agregation by using the previous peptide dataset.*

---

## Description

This function creates a column for the protein dataset after agregation by using the previous peptide dataset.

## Usage

```
BuildColumnToProteinDataset(peptideData, matAdj, columnName, proteinNames)
```

## Arguments

|              |   |
|--------------|---|
| peptideData  | A data.frame of meta data of peptides. It is the fData of the MSnset object.                                |
| matAdj       | The adjacency matrix used to agregate the peptides data.  |
| columnName   | The name of the column in fData(peptides_MSnset) that the user wants to keep in the new protein data.frame. |
| proteinNames | The names of the protein in the new dataset (i.e. rownames)   |

## Value

A vector

## Author(s)

Samuel Wiczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, FALSE)
data <- Biobase::fData(Exp1_R25_pept[1:1000])
protData <- pepAgregate(Exp1_R25_pept[1:1000], 'Protein_group.IDs', 'sum overall', M)
name <- "Protein.group.IDs"
proteinNames <- rownames(Biobase::fData(protData))
BuildColumnToProteinDataset(data, M, name,proteinNames )
```

---

BuildColumnToProteinDataset\_par

*creates a column for the protein dataset after agregation by using the previous peptide dataset.*

---

## Description

This function creates a column for the protein dataset after agregation by using the previous peptide dataset. It is a parallel version of the function BuildColumnToProteinDataset

## Usage

```
BuildColumnToProteinDataset_par(peptideData, matAdj, columnName, proteinNames)
```

## Arguments

|              |   |
|--------------|---|
| peptideData  | A data.frame of meta data of peptides. It is the fData of the MSnset object.                                |
| matAdj       | The adjacency matrix used to agregate the peptides data.  |
| columnName   | The name of the column in fData(peptides_MSnset) that the user wants to keep in the new protein data.frame. |
| proteinNames | The names of the protein in the new dataset (i.e. rownames)   |

## Value

A vector

## Author(s)

Samuel Wiczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, FALSE)
data <- Biobase::fData(Exp1_R25_pept[1:1000])
protData <- pepAgregate(Exp1_R25_pept[1:1000], ProtID, 'sum overall', M)
name <- "Protein.group.IDs"
proteinNames <- rownames(Biobase::fData(protData))
BuildColumnToProteinDataset_par(data, M, name,proteinNames )
```

---

|              |   |
|--------------|---|
| buildLogText | <i>Build the text information to be saved</i> |
|--------------|---|

---

**Description**

Build the text information to be saved after a process on an object of class MSnSet

**Usage**

```
buildLogText(name, l.params, ...)
```

**Arguments**

|          |  |
|----------|--|
| name     | The name of the process in Prostar                         |
| l.params | A list of parameters related to the process of the dataset |
| ...      | Parameter for the function getTextForImputation            |

**Value**

A string

**Author(s)**

Samuel Wieczorek

**Examples**

```
buildLogText("Original", list(filename="foo.MSnset"))
```

---

|                  |                                     |
|------------------|-------------------------------------|
| check.conditions | <i>Check if the design is valid</i> |
|------------------|-------------------------------------|

---

**Description**

This function check the validity of the conditions

**Usage**

```
## S3 method for class 'conditions'  
check(conds)
```

**Arguments**

|       |          |
|-------|----------|
| conds | A vector |
|-------|----------|

**Value**

A list

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
check.conditions(Biobase::pData(Exp1_R25_pept)$Label)
```

---

check.design

*Check if the design is valid*

---

**Description**

This function check the validity of the experimental design

**Usage**

```
## S3 method for class 'design'
check(sTab)
```

**Arguments**

sTab            The data.frame which correspond to the pData function of MSnbase

**Value**

A boolean

**Author(s)**

Thomas Burger, Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
check.design(Biobase::pData(Exp1_R25_pept)[,1:3])
```

---

compareNormalizationD *Builds a plot from a dataframe*

---

### Description

Plot to compare the quantitative proteomics data before and after normalization

### Usage

```
compareNormalizationD(qDataBefore, qDataAfter, labelsForLegend = NULL,  
  indData2Show = NULL, group2Color = "Condition")
```

### Arguments

|                 |   |
|-----------------|---|
| qDataBefore     | A dataframe that contains quantitative data before normalization.   |
| qDataAfter      | A dataframe that contains quantitative data after normalization.  |
| labelsForLegend | A vector of the conditions (labels) (one label per sample).   |
| indData2Show    | A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data.frame qDataBefore.                                      |
| group2Color     | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

### Value

A plot

### Author(s)

Samuel Wiczorek

### Examples

```
require(DAPARdata)  
data(Exp1_R25_pept)  
qDataBefore <- Biobase::exprs(Exp1_R25_pept)  
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]  
qDataAfter <- normalizeD(qDataBefore, labels, "Quantile Centering",  
  "within conditions")  
compareNormalizationD(qDataBefore, qDataAfter, labels)
```

---

compareNormalizationD\_HC

*Builds a plot from a dataframe. Same as compareNormalizationD but uses the library highcharter*

---

## Description

Plot to compare the quantitative proteomics data before and after normalization using the library highcharter

## Usage

```
compareNormalizationD_HC(qDataBefore, qDataAfter, labelsForLegend = NULL,
  indData2Show = NULL, group2Color = "Condition")
```

## Arguments

|                 |   |
|-----------------|---|
| qDataBefore     | A dataframe that contains quantitative data before normalization.   |
| qDataAfter      | A dataframe that contains quantitative data after normalization.  |
| labelsForLegend | A vector of the conditions (labels) (one label per sample).   |
| indData2Show    | A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data.frame qDataBefore.                                      |
| group2Color     | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

## Value

A plot

## Author(s)

Samuel Wieczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
qDataBefore <- Biobase::exprs(obj)
labels <- Biobase::pData(obj)[,"Label"]
qDataAfter <- normalizeD(qDataBefore,labels,"Quantile Centering",
  "within conditions")
compareNormalizationD_HC(qDataBefore, qDataAfter, labels)
```

---

```
compute.t.tests      xxxxxx
```

---

### Description

This function is xxxxxx

### Usage

```
## S3 method for class 't.tests'
compute(qData, Conditions, Contrast = "OnevsOne",
        type = "Student")
```

### Arguments

|            |   |
|------------|---|
| qData      | A matrix of quantitative data, without any missing values.  |
| Conditions | A vector of factor which indicates the name of the biological condition for each replicate.   |
| Contrast   | Indicates if the test consists of the comparison of each biological condition versus each of the other ones (Contrast=1; for example H0:"C1=C2" vs H1:"C1!=C2", etc.) or each condition versus all others (Contrast=2; e.g. H0:"C1=(C2+C3)/2" vs H1:"C1!=(C2+C3)/2", etc. if there are three conditions). |
| type       | xxxxx   |

### Value

A list of two items : FC and P\_Value; both are dataframe. The first one contains the logFC values of all the comparisons (one column for one comparison), the second one contains the pvalue of all the comparisons (one column for one comparison). The names of the columns for those two dataframes are identical and correspond to the description of the comparison.

### Author(s)

Florence Combes, Samuel Wiczorek

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
lapala <- findMECBlock(obj)
obj <- wrapper.impute.detQuant(obj)
obj <- reIntroduceMEC(obj, lapala)
obj <- wrapper.impute.detQuant(obj)
ttest <- compute.t.tests(Biobase::exprs(obj), Biobase::pData(obj)[,"Label"],1)
```

---

|             |   |
|-------------|---|
| corrMatrixD | <i>Displays a correlation matrix of the quantitative data of the exprs() table.</i> |
|-------------|---|

---

**Description**

Correlation matrix based on a MSnSet object

**Usage**

```
corrMatrixD(qData, samplesData, gradientRate = 5)
```

**Arguments**

|              |   |
|--------------|---|
| qData        | A dataframe of quantitative data.   |
| samplesData  | A dataframe where lines correspond to samples and columns to the meta-data for those samples. |
| gradientRate | The rate parameter to control the exponential law for the gradient of colors                  |

**Value**

A colored correlation matrix

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
samplesData <- Biobase::pData(Exp1_R25_pept)
corrMatrixD(qData, samplesData)
```

---

|                |   |
|----------------|---|
| corrMatrixD_HC | <i>Displays a correlation matrix of the quantitative data of the exprs() table.</i> |
|----------------|---|

---

**Description**

Correlation matrix based on a MSnSet object. Same as the function [corrMatrixD](#) but uses the package highcharter

**Usage**

```
corrMatrixD_HC(object, samplesData = NULL, rate = 0.5)
```



**Arguments**

|             |  |
|-------------|--|
| object      | The result of the cor function.  |
| samplesData | A dataframe in which lines correspond to samples and columns to the meta-data for those samples. |
| rate        | The rate parameter to control the exponential law for the gradient of colors                     |

**Value**

A colored correlation matrix

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
samplesData <- Biobase::pData(Exp1_R25_pept)
res <- cor(qData,use = 'pairwise.complete.obs')
corrMatrixD_HC(res, samplesData)
```

---

CountPep

*Compute the number of peptides used to aggregate proteins*

---

**Description**

This function computes the number of peptides used to aggregate proteins.

**Usage**

```
CountPep(M)
```

**Arguments**

|   |  |
|---|--|
| M | A "valued" adjacency matrix in which lines and columns correspond respectively to peptides and proteins. |
|---|--|

**Value**

A vector of boolean which is the adjacency matrix but with NA values if they exist in the intensity matrix.

**Author(s)**

Alexia Dorffer

**Examples**

```
library(DAPARdata)
data(Exp1_R25_pept)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, FALSE)
CountPep(M)
```

---

|              |   |
|--------------|---|
| createMSnset | <i>Creates an object of class MSnSet from text file</i> |
|--------------|---|

---

**Description**

Builds an object of class MSnSet from a single tabulated-like file for quantitative and meta-data and a dataframe for the samples description. It differs from the original MSnSet builder which requires three separated files tabulated-like quantitative proteomic data into a MSnSet object, including meta-data.

**Usage**

```
createMSnset(file, metadata = NULL, indExpData, indFData, indiceID = NULL,
  indexForOriginOfValue = NULL, logData = FALSE, replaceZeros = FALSE,
  pep_prot_data = NULL, versions = NULL)
```

**Arguments**

|                       |  |
|-----------------------|--|
| file                  | The name of a tab-separated file that contains the data.   |
| metadata              | A dataframe describing the samples (in lines).   |
| indExpData            | A vector of string where each element is the name of a column in designTable that have to be integrated in the fData() table of the MSnSet object. |
| indFData              | The name of column in file that will be the name of rows for the exprs() and fData() tables  |
| indiceID              | The indice of the column containing the ID of entities (peptides or proteins)  |
| indexForOriginOfValue | xxxxxxxxxxxx   |
| logData               | A boolean value to indicate if the data have to be log-transformed (Default is FALSE)  |
| replaceZeros          | A boolean value to indicate if the 0 and NaN values of intensity have to be replaced by NA (Default is FALSE)                                      |
| pep_prot_data         | A string that indicates whether the dataset is about   |
| versions              | A list of the following items: Prostar_Version, DAPAR_Version peptides or proteins.  |

**Value**

An instance of class MSnSet.

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```

require(DAPARdata)
require(Matrix)
exprsFile <- system.file("extdata", "Exp1_R25_pept.txt", package="DAPARdata")
metadataFile <- system.file("extdata", "samples_Exp1_R25.txt", package="DAPARdata")
metadata = read.table(metadataFile, header=TRUE, sep="\t", as.is=TRUE)
indExpData <- c(56:61)
indFData <- c(1:55,62:71)
indiceID <- 64
createMSnset(exprsFile, metadata,indExpData, indFData, indiceID, indexForOriginOfValue = NULL, pep_prot_dat

```

CVDistD

*Distribution of CV of entities***Description**

Builds a densityplot of the CV of entities in the `exprs()` table of a object. The CV is calculated for each condition (Label) present in the dataset (see the slot 'Label' in the `pData()` table)

**Usage**

```
CVDistD(qData, labels = NULL)
```

**Arguments**

|                     |   |
|---------------------|---|
| <code>qData</code>  | A dataframe that contains quantitative data.                |
| <code>labels</code> | A vector of the conditions (labels) (one label per sample). |

**Value**

A density plot

**Author(s)**

Florence Combes, Samuel Wiczorek

**See Also**

[densityPlotD](#).

**Examples**

```

require(DAPARdata)
data(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
CVDistD(Biobase::exprs(Exp1_R25_pept), labels)

```

---

`CVDistD_HC`*Distribution of CV of entities*

---

**Description**

Builds a densityplot of the CV of entities in the `exprs()` table of a object. The CV is calculated for each condition (Label) present in the dataset (see the slot 'Label' in the `pData()` table) Same as the function `CVDistD` but uses the package `highcharter`

**Usage**

```
CVDistD_HC(qData, labels = NULL)
```

**Arguments**

`qData` A dataframe that contains quantitative data.  
`labels` A vector of the conditions (labels) (one label per sample).

**Value**

A density plot

**Author(s)**

Samuel Wiczorek

**See Also**

[densityPlotD](#).

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
CVDistD_HC(Biobase::exprs(Exp1_R25_pept), labels)
```

---

`deleteLinesFromIndices`*Delete the lines in the matrix of intensities and the metadata table given their indice.*

---

**Description**

Delete the lines of `exprs()` table identified by their indice.

**Usage**

```
deleteLinesFromIndices(obj, deleteThat = NULL, processText = "")
```

**Arguments**

|             |  |
|-------------|--|
| obj         | An object of class MSnSet containing quantitative data.        |
| deleteThat  | A vector of integers which are the indices of lines to delete. |
| processText | A string to be included in the MSnSet object for log.          |

**Value**

An instance of class MSnSet that have been filtered.

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
deleteLinesFromIndices(Exp1_R25_pept, c(1:10))
```

---

|              |  |
|--------------|--|
| densityPlotD | <i>Builds a densityplot from a dataframe</i> |
|--------------|--|

---

**Description**

Densityplot of quantitative proteomics data over samples.

**Usage**

```
densityPlotD(qData, labelsForLegend = NULL, indData2Show = NULL,
             group2Color = "Condition")
```

**Arguments**

|                 |   |
|-----------------|---|
| qData           | A dataframe that contains quantitative data.  |
| labelsForLegend | A vector of the conditions (labels) (one label per sample).   |
| indData2Show    | A vector of indices to show in densityplot. If NULL, then all labels are displayed.   |
| group2Color     | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A density plot

**Author(s)**

Florence Combes, Samuel Wiczorek

**See Also**

[boxPlotD](#), [CVDistD](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
labels <- lab2Show <- Biobase::pData(Exp1_R25_pept)[,"Label"]
densityPlotD(qData, labels)
```

---

|                 |  |
|-----------------|--|
| densityPlotD_HC | <i>Builds a densityplot from a dataframe</i> |
|-----------------|--|

---

**Description**

Densityplot of quantitative proteomics data over samples. Same as the function [densityPlotD](#) but uses the package [highcharter](#)

**Usage**

```
densityPlotD_HC(qData, labelsForLegend = NULL, indData2Show = NULL,
  group2Color = "Condition")
```

**Arguments**

|                 |   |
|-----------------|---|
| qData           | A dataframe that contains quantitative data.  |
| labelsForLegend | A vector of the conditions (labels) (one label per sample).   |
| indData2Show    | A vector of indices to show in densityplot. If NULL, then all labels are displayed.   |
| group2Color     | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A density plot

**Author(s)**

Samuel Wieczorek

**See Also**

[boxPlotD](#), [CVDistD](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
labels <- lab2Show <- Biobase::pData(Exp1_R25_pept)[,"Label"]
densityPlotD_HC(qData, labels)
```

---

|                   |  |
|-------------------|--|
| diffAnaComputeFDR | <i>Computes the FDR corresponding to the p-values of the differential analysis using</i> |
|-------------------|--|

---

**Description**

This function is a wrapper to the function `adjust.p` from the `cp4p` package. It returns the FDR corresponding to the p-values of the differential analysis. The FDR is computed with the function `p.adjust{stats}`.

**Usage**

```
diffAnaComputeFDR(FC, pval, threshold_PVal = 0, threshold_LogFC = 0,
  pi0Method = 1)
```

**Arguments**

|                 |  |
|-----------------|--|
| FC              | The result (FC values) of the differential analysis processed by <code>limmaCompleteTest</code>            |
| pval            | The result (p-values) of the differential analysis processed by <code>limmaCompleteTest</code>             |
| threshold_PVal  | The threshold on p-value to distinguish between differential and non-differential data                     |
| threshold_LogFC | The threshold on log(Fold Change) to distinguish between differential and non-differential data            |
| pi0Method       | The parameter <code>pi0.method</code> of the method <code>adjust.p</code> in the package <code>cp4p</code> |

**Value**

The computed FDR value (floating number)

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
limma <- limmaCompleteTest(qData,sTab)
diffAnaComputeFDR(limma$FC[,1],limma$P_Value[,1])
```

---

`diffAnaGetSignificant` *Returns a MSnSet object with only proteins significant after differential analysis.*

---

### Description

Returns a MSnSet object with only proteins significant after differential analysis.

### Usage

```
diffAnaGetSignificant(obj)
```

### Arguments

`obj` An object of class MSnSet.

### Value

A MSnSet

### Author(s)

Alexia Dorffer

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
limma <- limmaCompleteTest(qData, sTab)
fc <- limma$FC[1]
pval <- limma$P_Value[1]
params <- list(design="OnevsOne", method="limma", th_logFC=0)
obj <- diffAnaSave(obj, limma, list(FC=fc, P_Value = pval), params)
signif <- diffAnaGetSignificant(obj)
```

---

`diffAnaSave` *Returns a MSnSet object with the results of the differential analysis performed with [limma](#) package.*

---

### Description

This method returns a class MSnSet object with the results of differential analysis.

### Usage

```
diffAnaSave(obj, allComp, data = NULL, l.params)
```



**Arguments**

|          |  |
|----------|--|
| obj      | An object of class MSnSet.   |
| allComp  | A list of two items which is the result of the function wrapper.limmaCompleteTest or xxxx  |
| data     | The result of the differential analysis processed by <code>limmaCompleteTest</code>  |
| l.params | A list of parameters: comp The name of the comparison th_pVal A float that indicates the threshold on p-value chosen to discriminate differential proteins. fdr The FDR based on the values of threshold_pVal and threshold_logFC cal-ibMethod The calibration method used to compute the calibration plot design xxxxxx th_logFC xxxx |

**Value**

A MSnSet

**Author(s)**

Alexia Dorffer, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
allComp <- limmaCompleteTest(qData,sTab)
data <- list(FC=allComp$FC[1], P_Value = allComp$P_Value[1])
params <- list(design="OnevsOne", method="limma", th_logFC=0)
diffAnaSave(obj, allComp, data, params)
```

---

diffAnaVolcanoplot      *Volcanoplot of the differential analysis*

---

**Description**

Plots a volcano plot after the differential analysis. Typically, the log of Fold Change is represented on the X-axis and the log<sub>10</sub> of the p-value is drawn on the Y-axis. When the threshold\_pVal and the threshold\_logFC are set, two lines are drawn respectively on the y-axis and the X-axis to visually distinguish between differential and non differential data.

**Usage**

```
diffAnaVolcanoplot(FC = NULL, pVal = NULL, threshold_pVal = 1e-60,
  threshold_logFC = 0, conditions = NULL)
```

**Arguments**

|                 |  |
|-----------------|--|
| FC              | A vector of the log(fold change) values of the differential analysis.  |
| pVal            | A vector of the p-value values returned by the differential analysis.  |
| threshold_pVal  | A floating number which represents the p-value that separates differential and non-differential data.                |
| threshold_logFC | A floating number which represents the log of the Fold Change that separates differential and non-differential data. |
| conditions      | A list of the names of condition 1 and 2 used for the differential analysis.   |

**Value**

A volcanoplot

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
limma <- limmaCompleteTest(qData,sTab)
diffAnaVolcanoplot(limma$FC[,1], limma$P_Value[,1])
```

---

diffAnaVolcanoplot\_rCharts

*Volcanoplot of the differential analysis*

---

**Description**

Plots an interactive volcanoplot after the differential analysis. Typically, the log of Fold Change is represented on the X-axis and the log10 of the p-value is drawn on the Y-axis. When the threshold\_pVal and the threshold\_logFC are set, two lines are drawn respectively on the y-axis and the X-axis to visually distinguish between differential and non differential data. With the use of the package Highcharter, a customizable tooltip appears when the user put the mouse's pointer over a point of the scatter plot.

**Usage**

```
diffAnaVolcanoplot_rCharts(df, threshold_pVal = 1e-60, threshold_logFC = 0,
  conditions = NULL, clickFunction = NULL)
```

**Arguments**

|                 |   |
|-----------------|---|
| df              | A dataframe which contains the following slots : x : a vector of the log(fold change) values of the differential analysis, y : a vector of the p-value values returned by the differential analysis. index : a vector of the rownames of the data. This dataframe must has been built with the option stringsAsFactors set to FALSE. There may be additional slots which will be used to show informations in the tooltip. The name of these slots must begin with the prefix "tooltip_". It will be automatically removed in the plot. |
| threshold_pVal  | A floating number which represents the p-value that separates differential and non-differential data.   |
| threshold_logFC | A floating number which represents the log of the Fold Change that separates differential and non-differential data.  |
| conditions      | A list of the names of condition 1 and 2 used for the differential analysis.  |
| clickFunction   | A string that contains a JavaScript function used to show info from slots in df. The variable this.index refers to the slot named index and allows to retrieve the right row to show in the tooltip   |

**Value**

An interactive volcanoplot

**Author(s)**

Samuel Wieczorek

**Examples**

```
library(highcharter)
library(tidyverse)
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
data <- limmaCompleteTest(qData,sTab)
df <- data.frame(x=data$FC, y = -log10(data$P_Value), index = as.character(rownames(obj)))
colnames(df) <- c("x", "y", "index")
tooltipSlot <- c("Sequence", "Score")
df <- cbind(df,Biobase::fData(obj)[tooltipSlot])
colnames(df) <- gsub(".", "_", colnames(df), fixed=TRUE)
if (ncol(df) > 3){
  colnames(df)[4:ncol(df)] <-
  paste("tooltip_", colnames(df)[4:ncol(df)], sep="")}
hc_clickFunction <- JS("function(event) {
Shiny.onInputChange('eventPointClicked', [this.index]);}")
cond <- c("25fmol", "10fmol")
diffAnaVolcanoplot_rCharts(df, 2.5, 1, cond,hc_clickFunction)
```

---

|           |   |
|-----------|---|
| enrich_GO | <i>Calculates GO enrichment classes for a given list of proteins/genes ID. It results an enrichResult instance.</i> |
|-----------|---|

---

### Description

This function is a wrapper to the function `enrichGO` from the package `clusterProfiler`. Given a vector of genes/proteins, it returns an `enrichResult` instance.

### Usage

```
enrich_GO(data, idFrom, orgdb, ont, readable = FALSE, pval, universe)
```

### Arguments

|                       |  |
|-----------------------|--|
| <code>data</code>     | A vector of ID (among ENSEMBL, ENTREZID, GENENAME, REFSEQ, UNIGENE, UNIPROT -can be different according to organisms)    |
| <code>idFrom</code>   | character indicating the input ID format (among ENSEMBL, ENTREZID, GENENAME, REFSEQ, UNIGENE, UNIPROT)                   |
| <code>orgdb</code>    | annotation Bioconductor package to use (character format)  |
| <code>ont</code>      | One of "MF", "BP", and "CC" subontologies  |
| <code>readable</code> | TRUE or FALSE (default FALSE)  |
| <code>pval</code>     | The qvalue cutoff (same parameter as in the function <code>enrichGO</code> of the package <code>clusterProfiler</code> ) |
| <code>universe</code> | a list of ID to be considered as the background for enrichment calculation   |

### Value

A `groupGOResult` instance.

### Author(s)

Florence Combes

### Examples

```
require(DAPARdata)
data(Exp1_R25_prot)
univ<-univ_AnnotDbPkg("org.Sc.sgd.db") #univ is the background
ego<-enrich_GO(data=fData(Exp1_R25_prot)$Protein.IDs, idFrom="UNIPROT",
orgdb="org.Sc.sgd.db",ont="MF", pval=0.05, universe = univ)
```

---

|              |  |
|--------------|--|
| findMECBlock | <i>Finds the LAPALA into a MSnSet object</i> |
|--------------|--|

---

**Description**

This method finds the LAPALA in a dataset.

**Usage**

```
findMECBlock(obj)
```

**Arguments**

obj                    An object of class MSnSet.

**Value**

A data.frame that contains the indexes of LAPALA

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
lapala <- findMECBlock(obj)
```

---

|                   |             |
|-------------------|-------------|
| formatLimmaResult | <i>xxxx</i> |
|-------------------|-------------|

---

**Description**

This function is xxxx

**Usage**

```
formatLimmaResult(fit, conds, contrast)
```

**Arguments**

fit                    xxxx  
conds                  xxxx  
contrast                xxxx

**Value**

A list of two dataframes : FC and P\_Value. The first one contains the logFC values of all the comparisons (one column for one comparison), the second one contains the pvalue of all the comparisons (one column for one comparison). The names of the columns for those two dataframes are identical and correspond to the description of the comparison.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
limma <- limmaCompleteTest(qData,sTab)
```

---

fudge2LRT

*Heuristic to choose the value of the hyperparameter (fudge factor) used to regularize the variance estimator in the likelihood ratio statistic*

---

**Description**

fudge2LRT: heuristic to choose the value of the hyperparameter (fudge factor) used to regularize the variance estimator in the likelihood ratio statistic (as implemented in samLRT). We follow the heuristic described in [1] and adapt the code of the fudge2 function in the siggene R package. [1] Tusher, Tibshirani and Chu, Significance analysis of microarrays applied to the ionizing radiation response, PNAS 2001 98: 5116-5121, (Apr 24).

**Usage**

```
fudge2LRT(lmm.res.h0, lmm.res.h1, cc, n, p, s, alpha = seq(0, 1, 0.05),
  include.zero = TRUE)
```

**Arguments**

|            |  |
|------------|--|
| lmm.res.h0 | a vector of object containing the estimates (used to compute the statistic) under H0 for each connected component. If the fast version of the estimator was used (as implemented in this package), lmm.res.h0 is a vector containing averages of squared residuals. If a fixed effect model was used, it is a vector of lm objects and if a mixed effect model was used it is a vector or lmer object. |
| lmm.res.h1 | similar to lmm.res.h0, a vector of object containing the estimates (used to compute the statistic) under H1 for each protein.  |
| cc         | a list containing the indices of peptides and proteins belonging to each connected component.  |
| n          | the number of samples used in the test   |

|              |  |
|--------------|--|
| p            | the number of proteins in the experiment   |
| s            | a vector containing the maximum likelihood estimate of the variance for the chosen model. When using the fast version of the estimator implemented in this package, this is the same thing as the input lmm.res.h1. For other models (e.g. mixed models) it can be obtained from samLRT. |
| alpha        | A vector of proportions used to build candidate values for the regularizer. We use quantiles of s with these proportions. Default to seq(0, 1, 0.05)   |
| include.zero | logical value indicating if 0 should be included in the list of candidates. Default to TRUE.   |

**Value**

(same as the fudge2 function of siggene): s.zero: the value of the fudge factor s0. alpha.hat: the optimal quantile of the 's' values. If s0=0, 'alpha.hat' will not be returned. vec.cv: the vector of the coefficients of variations. Following Tusher et al. (2001), the optimal 'alpha' quantile is given by the quantile that leads to the smallest CV of the modified test statistics. msg: a character string summarizing the most important information about the fudge factor.

**Author(s)**

Thomas Burger, Laurent Jacob

---

getIndicesConditions *Gets the conditions indices.*

---

**Description**

Returns a list for the two conditions where each slot is a vector of indices for the samples.

**Usage**

```
getIndicesConditions(labels, cond1, cond2)
```

**Arguments**

|        |   |
|--------|---|
| labels | A vector of strings containing the column "Label" of the pData().     |
| cond1  | A vector of Labels (a slot in the pData() table) for the condition 1. |
| cond2  | A vector of Labels (a slot in the pData() table) for the condition 2. |

**Value**

A list with two slots iCond1 and iCond2 containing respectively the indices of samples in the pData() table of the dataset.

**Author(s)**

Florence Combes, Samuel Wiczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
getIndicesConditions(labels, "25fmol", "10fmol")
```

---

getIndicesOfLinesToRemove

*Get the indices of the lines to delete, based on a prefix string*

---

## Description

This function returns the indice of the lines to delete, based on a prefix string

## Usage

```
getIndicesOfLinesToRemove(obj, idLine2Delete = NULL, prefix = NULL)
```

## Arguments

|               |  |
|---------------|--|
| obj           | An object of class MSnSet.                                   |
| idLine2Delete | The name of the column that correspond to the data to filter |
| prefix        | A character string that is the prefix to find in the data    |

## Value

A vector of integers.

## Author(s)

Samuel Wieczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
getIndicesOfLinesToRemove(Exp1_R25_pept, "Potential.contaminant", prefix="+")
```



---

`getListNbValuesInLines`*Returns the possible number of values in lines in the data*

---

**Description**

Returns the possible number of values in lines in a matrix.

**Usage**

```
getListNbValuesInLines(obj, type = "wholeMatrix")
```

**Arguments**

|                   |                           |
|-------------------|---------------------------|
| <code>obj</code>  | An object of class MSnSet |
| <code>type</code> | xxxxxxx                   |

**Value**

An integer

**Author(s)**

Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
getListNbValuesInLines(Exp1_R25_pept)
```

---

`getNumberOf`*Number of lines with prefix*

---

**Description**

Returns the number of lines, in a given column, where content matches the prefix.

**Usage**

```
getNumberOf(obj, name = NULL, prefix = NULL)
```

**Arguments**

|                     |                            |
|---------------------|----------------------------|
| <code>obj</code>    | An object of class MSnSet. |
| <code>name</code>   | The name of a column.      |
| <code>prefix</code> | A string                   |

**Value**

An integer

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
getNumberOf(Exp1_R25_pept, "Potential.contaminant", "+")
```

---

getNumberOfEmptyLines *Returns the number of empty lines in the data*

---

**Description**

Returns the number of empty lines in a matrix.

**Usage**

```
getNumberOfEmptyLines(qData)
```

**Arguments**

qData            A matrix corresponding to the quantitative data.

**Value**

An integer

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
getNumberOfEmptyLines(qData)
```

---

getPaletteForLabels     *Palette for plots in DAPAR*

---

**Description**

Selects colors for the plots in DAPAR based on the different conditions in the dataset. The palette is derived from the brewer palette "Dark2" (see [RColorBrewer](#)).

**Usage**

```
getPaletteForLabels(labels)
```

**Arguments**

labels                    A vector of labels (strings).

**Value**

A palette designed for the data manipulated in DAPAR

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
getPaletteForLabels(labels)
```

---

getPaletteForLabels\_HC     *Palette for highcharter plots used in DAPAR*

---

**Description**

Selects colors for the highcharter plots in DAPAR based on the different conditions in the dataset. The palette is derived from the brewer palette "Dark2" (see [RColorBrewer](#)).

**Usage**

```
getPaletteForLabels_HC(labels)
```

**Arguments**

labels                    A vector of labels (strings).

**Value**

A palette designed for the data manipulated in DAPAR

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
getPaletteForLabels_HC(labels)
```

---

getPaletteForReplicates

*Palette for plot the replicates in DAPAR*

---

**Description**

Selects colors for the plots in DAPAR based on the replicates in the dataset. The palette is derived from the brewer palette "Dark2" (see [RColorBrewer](#)).

**Usage**

```
getPaletteForReplicates(nColors)
```

**Arguments**

nColors            The desired number of colors

**Value**

A palette designed for the data manipulated in DAPAR

**Author(s)**

Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
n <- nrow(Biobase::pData(Exp1_R25_pept))
getPaletteForReplicates(n)
```

---

`getPaletteForReplicates_HC`*Palette for highcharter plot the replicates in DAPAR*

---

**Description**

Selects colors for the highcharter plots in DAPAR based on the replicates in the dataset. The palette is derived from the brewer palette "Dark2" (see [RColorBrewer](#)).

**Usage**

```
getPaletteForReplicates_HC(nColors)
```

**Arguments**

`nColors`            The desired number of colors

**Value**

A palette designed for the data manipulated in DAPAR

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
n <- nrow(Biobase::pData(Exp1_R25_pept))
getPaletteForReplicates_HC(n)
```

---

`getPourcentageOfMV`    *Percentage of missing values*

---

**Description**

Returns the percentage of missing values in the quantitative data (`exprs()` table of the dataset).

**Usage**

```
getPourcentageOfMV(obj)
```

**Arguments**

`obj`                An object of class MSnSet.

**Value**

A floating number

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
getPourcentageOfMV(Exp1_R25_pept)
```

---

getProcessingInfo      *Returns the contents of the slot processing of an object of class MSnSet*

---

**Description**

Returns the contents of the slot processing of an object of class MSnSet

**Usage**

```
getProcessingInfo(obj)
```

**Arguments**

obj                    An object (peptides) of class MSnSet.

**Value**

The slot processing of obj@processingData

**Author(s)**

Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
getProcessingInfo(Exp1_R25_pept)
```

---

|                  |   |
|------------------|---|
| getProteinsStats | <i>computes the number of proteins that are only defined by specific peptides, shared peptides or a mixture of two.</i> |
|------------------|---|

---

**Description**

This function computes the number of proteins that are only defined by specific peptides, shared peptides or a mixture of two.

**Usage**

```
getProteinsStats(matUnique, matShared)
```

**Arguments**

|           |  |
|-----------|--|
| matUnique | The adjacency matrix with only specific peptides.            |
| matShared | The adjacency matrix with both specific and shared peptides. |

**Value**

A list

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
protID <- "Protein.group.IDs"
MShared <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, FALSE)
MUnique <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, TRUE)
getProteinsStats(MUnique, MShared)
```

---

|                 |   |
|-----------------|---|
| getQuantile4Imp | <i>Quantile imputation value definition</i> |
|-----------------|---|

---

**Description**

This method returns the q-th quantile of each column of an expression set, up to a scaling factor

**Usage**

```
getQuantile4Imp(qData, qval = 0.025, factor = 1)
```

**Arguments**

|        |  |
|--------|--|
| qData  | An expression set containing quantitative values of various replicates |
| qval   | The quantile used to define the imputation value                       |
| factor | A scaling factor to multiply the imputation value with                 |

**Value**

A list of two vectors, respectively containing the imputation values and the rescaled imputation values

**Author(s)**

Thomas Burger

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
getQuantile4Imp(qData)
```

---

getTextForAggregation *Build the text information for the Aggregation process*

---

**Description**

Builds the text information for the Aggregation process

**Usage**

```
getTextForAggregation(l.params)
```

**Arguments**

l.params            A list of parameters related to the process of the dataset

**Value**

A string

**Author(s)**

Samuel Wieczorek

**Examples**

```
getTextForAggregation(list(POV_algorithm="slsa",MEC_algorithm="fixedValue", MEC_fixedValue = 0))
```



---

`getTextForAnaDiff`      *Build the text information for the Aggregation process*

---

**Description**

Build the text information for the differential Analysis process

**Usage**

```
getTextForAnaDiff(l.params)
```

**Arguments**

`l.params`      A list of parameters related to the process of the dataset

**Value**

A string

**Author(s)**

Samuel Wieczorek

**Examples**

```
getTextForAnaDiff(list(design="OnevsOne",method="Limma"))
```

---

`getTextForFiltering`      *Build the text information for the filtering process*

---

**Description**

Build the text information for the filtering process

**Usage**

```
getTextForFiltering(l.params)
```

**Arguments**

`l.params`      A list of parameters related to the process of the dataset

**Value**

A string

**Author(s)**

Samuel Wieczorek

**Examples**

```
getTextForFiltering(list(mvFilterType="wholeMatrix",mvThNA=3))
```

---

getTextForGOAnalysis *Build the text information for the Aggregation process*

---

**Description**

Build the text information for the Aggregation process

**Usage**

```
getTextForGOAnalysis(l.params)
```

**Arguments**

l.params            A list of parameters related to the process of the dataset

**Value**

A string

**Author(s)**

Samuel Wieczorek

**Examples**

```
getTextForGOAnalysis(list())
```

---

getTextForImputation *Build the text information for the Imputation process*

---

**Description**

Build the text information for the Imputation process

**Usage**

```
getTextForImputation(l.params, level)
```

**Arguments**

l.params            A list of parameters related to the process of the dataset  
level                The type of data (peptide or protein)

**Value**

A string

**Author(s)**

Samuel Wieczorek

**Examples**

```
getTextForImputation(list(POV_algorithm="slsa",MEC_algorithm="fixedValue", MEC_fixedValue = 0), level="prot
```

---

```
getTextForNewDataset Build the text information for a new dataset
```

---

**Description**

Build the text information for a new dataset

**Usage**

```
getTextForNewDataset(l.params)
```

**Arguments**

l.params            A list of parameters related to the process of the dataset

**Value**

A string

**Author(s)**

Samuel Wiczorek

**Examples**

```
getTextForNewDataset(list(filename="foo.MSnset"))
```

---

```
getTextForNormalization  
Build the text information for the Normalization process
```

---

**Description**

Build the text information for the Normalization process

**Usage**

```
getTextForNormalization(l.params)
```

**Arguments**

l.params            A list of parameters related to the process of the dataset

**Value**

A string

**Author(s)**

Samuel Wiczorek

**Examples**

```
getTextForNormalization(list(method="Sum by columns"))
```

---

|                |   |
|----------------|---|
| GOAnalysisSave | <i>Returns an MSnSet object with the results of the GO analysis performed with the functions <code>enrichGO</code> and/or <code>groupGO</code> of the <code>clusterProfiler</code> package.</i> |
|----------------|---|

---

**Description**

This method returns an MSnSet object with the results of the Gene Ontology analysis.

**Usage**

```
GOAnalysisSave(obj, ggo_res = NULL, ego_res = NULL, organism, ontology,
  levels, pvalueCutoff, typeUniverse)
```

**Arguments**

|                           |   |
|---------------------------|---|
| <code>obj</code>          | An object of the class MSnSet   |
| <code>ggo_res</code>      | The object returned by the function <code>group_GO</code> of the package DAPAR or the function <code>groupGO</code> of the package <code>clusterProfiler</code>   |
| <code>ego_res</code>      | The object returned by the function <code>enrich_GO</code> of the package DAPAR or the function <code>enrichGO</code> of the package <code>clusterProfiler</code> |
| <code>organism</code>     | The parameter <code>OrgDb</code> of the functions <code>bitr</code> , <code>groupGO</code> and <code>enrichGO</code>  |
| <code>ontology</code>     | One of "MF", "BP", and "CC" subontologies   |
| <code>levels</code>       | A vector of the different GO grouping levels to save  |
| <code>pvalueCutoff</code> | The qvalue cutoff (same parameter as in the function <code>enrichGO</code> of the package <code>clusterProfiler</code> )  |
| <code>typeUniverse</code> | The type of background to be used. Values are 'Entire Organism', 'Entire dataset' or 'Custom'. In the latter case, a file should be uploaded by the user          |

**Value**

An object of the class MSnSet

**Author(s)**

Samuel Wiczorek

---

|              |  |
|--------------|--|
| GraphPepProt | <i>Function to create a histogram that shows the repartition of peptides w.r.t. the proteins</i> |
|--------------|--|

---

**Description**

Method to create a plot with proteins and peptides on a MSnSet object (peptides)

**Usage**

```
GraphPepProt(mat)
```

**Arguments**

mat                    An adjacency matrix.

**Value**

A histogram

**Author(s)**

Alexia Dorffer, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
mat <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], "Protein.group.IDs")
GraphPepProt(mat)
```

---

|          |  |
|----------|--|
| group_GO | <i>Calculates the GO profile of a vector of genes/proteins at a given level of the Gene Ontology</i> |
|----------|--|

---

**Description**

This function is a wrapper to the function `groupGO` from the package `clusterProfiler`. Given a vector of genes/proteins, it returns the GO profile at a specific level. It returns a `groupGOResult` instance.

**Usage**

```
group_GO(data, idFrom, orgdb, ont, level, readable = FALSE)
```

**Arguments**

|          |   |
|----------|---|
| data     | A vector of ID (among ENSEMBL, ENTREZID, GENENAME, REFSEQ, UNIGENE, UNIPROT -can be different according to organisms) |
| idFrom   | character indicating the input ID format (among ENSEMBL, ENTREZID, GENENAME, REFSEQ, UNIGENE, UNIPROT)                |
| orgdb    | annotation Bioconductor package to use (character format)   |
| ont      | on which ontology to perform the analysis (MF, BP or CC)  |
| level    | level of the ontolofy to perform the analysis   |
| readable | TRUE or FALSE (default FALSE)   |

**Value**

GO profile at a specific level

**Author(s)**

Florence Combes

**Examples**

```
require(DAPARdata)
data(Exp1_R25_prot)
ggo<-group_GO(data=fData(Exp1_R25_prot)$Protein.IDs, idFrom="UNIPROT",
orgdb="org.Sc.sgd.db", ont="MF", level=2)
```

---

hc\_FC\_DensityPlot      *Density plots of FC values*

---

**Description**

This function show the density plots of Fold Change (the same as calculated by limma) for a list of the comparisons of conditions in a differnetial analysis.

**Usage**

```
hc_FC_DensityPlot(df_FC, threshold_LogFC = 0)
```

**Arguments**

|                 |   |
|-----------------|---|
| df_FC           | A dataframe that contains the FC values   |
| threshold_LogFC | The threshold on log(Fold Change) to distinguish between differential and non-differential data |

**Value**

A highcharts density plot

**Author(s)**

Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
limma <- limmaCompleteTest(qData,sTab)
hc_FC_DensityPlot(limma$FC)
```

---

`hc_mvTypePlot2`*Distribution of Observed values with respect to intensity values*

---

**Description**

This method shows density plots which represents the repartition of Partial Observed Values for each replicate in the dataset. The colors correspond to the different conditions (slot Label in in the dataset of class MSnSet). The x-axis represent the mean of intensity for one condition and one entity in the dataset (i. e. a protein) whereas the y-axis count the number of observed values for this entity and the considered condition.

**Usage**

```
hc_mvTypePlot2(qData, labels)
```

**Arguments**

|                     |   |
|---------------------|---|
| <code>qData</code>  | A dataframe that contains quantitative data.                |
| <code>labels</code> | A vector of the conditions (labels) (one label per sample). |

**Value**

Density plots

**Author(s)**

Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
hc_mvTypePlot2(qData, labels)
```

---

|               |  |
|---------------|--|
| heatmap.DAPAR | <i>This function is inspired from the function <a href="#">heatmap.2</a> that displays quantitative data in the <code>exprs()</code> table of an object of class <code>MSet</code>. For more information, please refer to the help of the <code>heatmap.2</code> function.</i> |
|---------------|--|

---

## Description

Heatmap inspired by the `heatmap.2` function.

## Usage

```
heatmap.DAPAR(x, col = heat.colors(100), srtCol = NULL, labCol = NULL,  
  labRow = NULL, key = TRUE, key.title = NULL, main = NULL,  
  ylab = NULL)
```

## Arguments

|                        |  |
|------------------------|--|
| <code>x</code>         | A dataframe that contains quantitative data.                                     |
| <code>col</code>       | colors used for the image. Defaults to heat colors ( <code>heat.colors</code> ). |
| <code>srtCol</code>    | angle of column labels, in degrees from horizontal                               |
| <code>labCol</code>    | character vectors with column labels to use.                                     |
| <code>labRow</code>    | character vectors with row labels to use.  |
| <code>key</code>       | logical indicating whether a color-key should be shown.                          |
| <code>key.title</code> | main title of the color key. If set to <code>NA</code> no title will be plotted. |
| <code>main</code>      | main title; default to none.   |
| <code>ylab</code>      | y-axis title; default to none.   |

## Value

A heatmap

## Author(s)

Samuel Wieczorek

## Examples

```
require(DAPARdata)  
data(Exp1_R25_pept)  
obj <- mvFilter(Exp1_R25_pept, "wholeMatrix", 6)  
qData <- Biobase::exprs(obj)  
heatmap.DAPAR(qData)
```



---

|          |  |
|----------|--|
| heatmapD | <i>This function is a wrapper to <a href="#">heatmap.2</a> that displays quantitative data in the <code>exprs()</code> table of an object of class <code>MSnSet</code></i> |
|----------|--|

---

**Description**

Heatmap of the quantitative proteomic data of a `MSnSet` object

**Usage**

```
heatmapD(qData, distance = "euclidean", cluster = "complete",  
         dendro = FALSE)
```

**Arguments**

|                       |   |
|-----------------------|---|
| <code>qData</code>    | A dataframe that contains quantitative data.  |
| <code>distance</code> | The distance used by the clustering algorithm to compute the dendrogram. See <code>help(heatmap.2)</code> |
| <code>cluster</code>  | the clustering algorithm used to build the dendrogram. See <code>help(heatmap.2)</code>                   |
| <code>dendro</code>   | A boolean to indicate if the dendrogram has to be displayed   |

**Value**

A heatmap

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)  
data(Exp1_R25_pept)  
obj <- mvFilter(Exp1_R25_pept[1:1000], "wholeMatrix", 6)  
qData <- Biobase::exprs(obj)  
heatmapD(qData)
```

---

|                              |                                 |
|------------------------------|---------------------------------|
| <code>impute.detQuant</code> | <i>Deterministic imputation</i> |
|------------------------------|---------------------------------|

---

**Description**

This method replaces each missing value by a given value

**Usage**

```
impute.detQuant(qData, values)
```

**Arguments**

|        |  |
|--------|--|
| qData  | An expression set containing quantitative or missing values      |
| values | A vector with as many elements as the number of columns of qData |

**Value**

An imputed dataset

**Author(s)**

Thomas Burger

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
values <- getQuantile4Imp(qData)$shiftedImpVal
impute.detQuant(qData, values)
```

---

impute.pa2

*Missing values imputation from a MSnSet object*

---

**Description**

This method is a variation to the function `impute.pa` from the package `imp4p`.

**Usage**

```
impute.pa2(tab, conditions, q.min = 0, q.norm = 3, eps = 0,
  distribution = "unif")
```

**Arguments**

|              |   |
|--------------|---|
| tab          | An object of class <code>MSnSet</code> .  |
| conditions   | A vector of conditions in the dataset   |
| q.min        | A quantile value of the observed values allowing defining the maximal value which can be generated. This maximal value is defined by the quantile <code>q.min</code> of the observed values distribution minus <code>eps</code> . Default is 0 (the maximal value is the minimum of observed values minus <code>eps</code> ). |
| q.norm       | A quantile value of a normal distribution allowing defining the minimal value which can be generated. Default is 3 (the minimal value is the maximal value minus $qn * \text{median}(\text{sd}(\text{observed values}))$ where <code>sd</code> is the standard deviation of a row in a condition).                            |
| eps          | A value allowing defining the maximal value which can be generated. This maximal value is defined by the quantile <code>q.min</code> of the observed values distribution minus <code>eps</code> . Default is 0.   |
| distribution | The type of distribution used. Values are <code>unif</code> or <code>beta</code> .  |

**Value**

The object obj which has been imputed

**Author(s)**

Thomas Burger, Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.impute.pa2(Exp1_R25_pept[1:1000], distribution="beta")
```

---

is.MV

*Similar to the function is.na but focused on the equality with the missing values in the dataset (type 'POV' and 'MEC')*

---

**Description**

Similar to the function is.na but focused on the equality with the missing values in the dataset (type 'POV' and 'MEC')

**Usage**

```
is.MV(data)
```

**Arguments**

data            A data.frame

**Value**

A boolean dataframe

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept
data <- Biobase::fData(obj)[,obj@experimentData@other$OriginOfValues]
is.MV(data)
```

---

|           |  |
|-----------|--|
| is.OfType | <i>Similar to the function is.na but focused on the equality with the paramter 'type'.</i> |
|-----------|--|

---

**Description**

Similar to the function `is.na` but focused on the equality with the paramter 'type'.

**Usage**

```
is.OfType(data, type)
```

**Arguments**

|                   |                                      |
|-------------------|--------------------------------------|
| <code>data</code> | A <code>data.frame</code>            |
| <code>type</code> | The value to search in the dataframe |

**Value**

A boolean dataframe

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept
data <- Biobase::fData(obj)[,obj@experimentData@other$OriginOfValues]
is.OfType(data, "MEC")
```

---

LH0

xxxxxx

---

**Description**

This function is xxxxxxxx

**Usage**

```
LH0(X, y1, y2)
```

**Arguments**

|                 |   |
|-----------------|---|
| <code>X</code>  | an <code>n.pep*n.prot</code> indicator matrix.                      |
| <code>y1</code> | <code>n.pep*n.samples</code> matrice giving the observed counts for |
| <code>y2</code> | <code>n.pep*n.samples</code> matrice giving the observed counts for |

**Value**

xxxxxxxxxx..

**Author(s)**

Thomas Burger, Laurent Jacob

---

LH0.lmxxxxxx

---

**Description**

This function is xxxxxxxx

**Usage**

LH0.lm(X, y1, y2)

**Arguments**

|    |   |
|----|---|
| X  | an n.pep*n.prot indicator matrix.   |
| y1 | n.pep*n.samples matrice giving the observed counts for each peptide in each sample from the condition 1 |
| y2 | n.pep*n.samples matrice giving the observed counts for each peptide in each sample from the condition 2 |

**Value**

xxxxxxxxxx..

**Author(s)**

Thomas Burger, Laurent Jacob

---

LH1xxxxxx

---

**Description**

This function is xxxxxxxx

**Usage**

LH1(X, y1, y2, j)

**Arguments**

|    |   |
|----|---|
| X  | an n.pep*n.prot indicator matrix.                             |
| y1 | n.pep*n.samples matrice giving the observed counts for        |
| y2 | n.pep*n.samples matrice giving the observed counts for        |
| j  | the index of the protein being tested, ie which has different |

**Value**

XXXXXXXXXX..

**Author(s)**

Thomas Burger, Laurent Jacob

LH1.lm

xxxxxx

**Description**

This function is xxxxxxxx

**Usage**

LH1.lm(X, y1, y2, j)

**Arguments**

|    |   |
|----|---|
| X  | an n.pep*n.prot indicator matrix.                             |
| y1 | n.pep*n.samples matrix giving the observed counts for         |
| y2 | n.pep*n.samples matrix giving the observed counts for         |
| j  | the index of the protein being tested, ie which has different |

**Value**

XXXXXXXXXX..

**Author(s)**

Thomas Burger, Laurent Jacob

limmaCompleteTest

*Computes a hierarchical differential analysis*

**Description**

This function is a limmaCompleteTest

**Usage**

limmaCompleteTest(qData, sTab, comp.type = "OnevsOne")

**Arguments**

|           |  |
|-----------|--|
| qData     | A matrix of quantitative data, without any missing values.   |
| sTab      | A dataframe of experimental design (pData()).  |
| comp.type | A string that corresponds to the type of comparison. Values are: 'OnevsOne' and 'OnevsAll'; default is 'OnevsOne'. |

**Value**

A list of two dataframes : FC and P\_Value. The first one contains the logFC values of all the comparisons (one column for one comparison), the second one contains the pvalue of all the comparisons (one column for one comparison). The names of the columns for those two dataframes are identical and correspond to the description of the comparison.

**Author(s)**

Thomas Burger, Quentin Giai-Gianetto, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
limma <- limmaCompleteTest(qData,sTab)
```

---

listSheets

*This function returns the list of the sheets names in a Excel file.*

---

**Description**

This function lists all the sheets of an Excel file.

**Usage**

```
listSheets(file)
```

**Arguments**

file            The name of the Excel file.

**Value**

A vector

**Author(s)**

Samuel Wiczorek

---

make.contrast                      *Builds the contrast matrix*

---

### Description

This function builds the contrast matrix

### Usage

```
make.contrast(design, condition, contrast = 1)
```

### Arguments

|           |   |
|-----------|---|
| design    | The data.frame which correspond to the pData function of MSnbase  |
| condition | xxxxx   |
| contrast  | An integer that Indicates if the test consists of the comparison of each biological condition versus each of the other ones (Contrast=1; for example H0:"C1=C2" vs H1:"C1!=C2", etc.) or each condition versus all others (Contrast=2; e.g. H0:"C1=(C2+C3)/2" vs H1:"C1!=(C2+C3)/2", etc. if there are three conditions). |

### Value

A constrat matrix

### Author(s)

Thomas Burger, Quentin Giai-Gianetto, Samuel Wiczorek

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
design <- make.design(Biobase::pData(Exp1_R25_pept))
conds <- Biobase::pData(Exp1_R25_pept)$Label
make.contrast(design, conds)
```

---

make.design                      *Builds the design matrix*

---

### Description

This function builds the design matrix

### Usage

```
make.design(sTab)
```

### Arguments

|      |  |
|------|--|
| sTab | The data.frame which correspond to the pData function of MSnbase |
|------|--|



**Value**

A design matrix

**Author(s)**

Thomas Burger, Quentin Giai-Gianetto, Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
make.design(Biobase::pData(Exp1_R25_pept))
```

---

|                            |  |
|----------------------------|--|
| <code>make.design.1</code> | <i>Builds the design matrix for designs of level 1</i> |
|----------------------------|--|

---

**Description**

This function builds the design matrix for design of level 1

**Usage**

```
make.design.1(sTab)
```

**Arguments**

sTab            The data.frame which correspond to the pData function of MSnbase

**Value**

A design matrix

**Author(s)**

Thomas Burger, Quentin Giai-Gianetto, Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
make.design.1(Biobase::pData(Exp1_R25_pept))
```

---

|               |  |
|---------------|--|
| make.design.2 | <i>Builds the design matrix for designs of level 2</i> |
|---------------|--|

---

**Description**

This function builds the design matrix for design of level 2

**Usage**

```
make.design.2(sTab)
```

**Arguments**

sTab            The data.frame which correspond to the pData function of MSnbase

**Value**

A design matrix

**Author(s)**

Thomas Burger, Quentin Giai-Gianetto, Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
make.design.2(Biobase::pData(Exp1_R25_pept))
```

---

|               |  |
|---------------|--|
| make.design.3 | <i>Builds the design matrix for designs of level 3</i> |
|---------------|--|

---

**Description**

This function builds the design matrix for design of level 3

**Usage**

```
make.design.3(sTab)
```

**Arguments**

sTab            The data.frame which correspond to the pData function of MSnbase

**Value**

A design matrix

**Author(s)**

Thomas Burger, Quentin Giai-Gianetto, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
make.design.3(Biobase::pData(Exp1_R25_pept))
```

---

|              |  |
|--------------|--|
| MeanPeptides | <i>Compute the intensity of proteins as the mean of the intensities of their peptides.</i> |
|--------------|--|

---

**Description**

This function computes the intensity of proteins as the mean of the intensities of their peptides.

**Usage**

```
MeanPeptides(matAdj, expr)
```

**Arguments**

|        |  |
|--------|--|
| matAdj | An adjacency matrix in which lines and columns correspond respectively to peptides and proteins. |
| expr   | A matrix of intensities of peptides  |

**Value**

A matrix of intensities of proteins

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
protID <- "Protein.group.IDs"
matAdj <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, FALSE)
MeanPeptides(matAdj, Biobase::exprs(Exp1_R25_pept[1:1000]))
```

---

`mvFilter`*Filter lines in the matrix of intensities w.r.t. some criteria*

---

### Description

Filters the lines of `exprs()` table with conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

### Usage

```
mvFilter(obj, type, th, processText = NULL)
```

### Arguments

|                          |  |
|--------------------------|--|
| <code>obj</code>         | An object of class <code>MSnSet</code> containing quantitative data.                                       |
| <code>type</code>        | Method used to choose the lines to delete. Values are : "None", "wholeMatrix", "allCond", "atLeastOneCond" |
| <code>th</code>          | An integer value of the threshold  |
| <code>processText</code> | A string to be included in the <code>MSnSet</code> object for log.   |

### Details

The different methods are : "wholeMatrix": given a threshold `th`, only the lines that contain at least `th` values are kept. "allCond": given a threshold `th`, only the lines which contain at least `th` values for each of the conditions are kept. "atLeastOneCond": given a threshold `th`, only the lines that contain at least `th` values, and for at least one condition, are kept.

### Value

An instance of class `MSnSet` that have been filtered.

### Author(s)

Florence Combes, Samuel Wiczorek

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
mvFilter(Exp1_R25_pept, "wholeMatrix", 2)
```

---

mvFilterFromIndices     *Filter lines in the matrix of intensities w.r.t. some criteria*

---

## Description

Filters the lines of `exprs()` table with conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

## Usage

```
mvFilterFromIndices(obj, keepThat = NULL, processText = "")
```

## Arguments

|                          |  |
|--------------------------|--|
| <code>obj</code>         | An object of class <code>MSnSet</code> containing quantitative data. |
| <code>keepThat</code>    | A vector of integers which are the indices of lines to keep.         |
| <code>processText</code> | A string to be included in the <code>MSnSet</code> object for log.   |

## Details

The different methods are : "wholeMatrix": given a threshold `th`, only the lines that contain at least `th` values are kept. "allCond": given a threshold `th`, only the lines which contain at least `th` values for each of the conditions are kept. "atLeastOneCond": given a threshold `th`, only the lines that contain at least `th` values, and for at least one condition, are kept.

## Value

An instance of class `MSnSet` that have been filtered.

## Author(s)

Florence Combes, Samuel Wiczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
mvFilterFromIndices(Exp1_R25_pept, c(1:10))
```

---

mvFilterGetIndices      *Filter lines in the matrix of intensities w.r.t. some criteria*

---

### Description

Returns the indices of the lines of `exprs()` table to delete w.r.t. the conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

### Usage

```
mvFilterGetIndices(obj, type, th)
```

### Arguments

|                   |  |
|-------------------|--|
| <code>obj</code>  | An object of class <code>MSnSet</code> containing quantitative data.   |
| <code>type</code> | Method used to choose the lines to delete. Values are : "None", "EmptyLines", "wholeMatrix", "allCond", "atLeastOneCond" |
| <code>th</code>   | An integer value of the threshold  |

### Details

The different methods are : "wholeMatrix": given a threshold `th`, only the lines that contain at least `th` values are kept. "allCond": given a threshold `th`, only the lines which contain at least `th` values for each of the conditions are kept. "atLeastOneCond": given a threshold `th`, only the lines that contain at least `th` values, and for at least one condition, are kept.

### Value

An vector of indices that correspond to the lines to keep.

### Author(s)

Florence Combes, Samuel Wiczorek

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
mvFilterGetIndices(Exp1_R25_pept, "wholeMatrix", 2)
```

---

|         |                                    |
|---------|------------------------------------|
| mvHisto | <i>Histogram of missing values</i> |
|---------|------------------------------------|

---

**Description**

This method plots a histogram of missing values.

**Usage**

```
mvHisto(qData, samplesData, labels, indLegend = "auto", showValues = FALSE)
```

**Arguments**

|             |   |
|-------------|---|
| qData       | A dataframe that contains quantitative data.  |
| samplesData | A dataframe where lines correspond to samples and columns to the meta-data for those samples. |
| labels      | A vector of the conditions (labels) (one label per sample).                                   |
| indLegend   | The indices of the column name's in pData() tab   |
| showValues  | A logical that indicates whether numeric values should be drawn above the bars.               |

**Value**

A histogram

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
samplesData <- Biobase::pData(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
mvHisto(qData, samplesData, labels, indLegend="auto", showValues=TRUE)
```

---

|            |                                    |
|------------|------------------------------------|
| mvHisto_HC | <i>Histogram of missing values</i> |
|------------|------------------------------------|

---

**Description**

This method plots a histogram of missing values. Same as the function mvHisto but uses the package highcharter

**Usage**

```
mvHisto_HC(qData, samplesData, labels, indLegend = "auto",
  showValues = FALSE)
```

**Arguments**

|             |   |
|-------------|---|
| qData       | A dataframe that contains quantitative data.  |
| samplesData | A dataframe where lines correspond to samples and columns to the meta-data for those samples. |
| labels      | A vector of the conditions (labels) (one label per sample).                                   |
| indLegend   | The indices of the column name's in pData() tab   |
| showValues  | A logical that indicates wether numeric values should be drawn above the bars.                |

**Value**

A histogram

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
samplesData <- Biobase::pData(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
mvHisto_HC(qData, samplesData, labels, indLegend="auto", showValues=TRUE)
```

---

mvImage

*Heatmap of missing values*


---

**Description**

Plots a heatmap of the quantitative data. Each column represent one of the conditions in the object of class MSnSet and the color is proportional to the mean of intensity for each line of the dataset. The lines have been sorted in order to vizualize easily the different number of missing values. A white square is plotted for missing values.

**Usage**

```
mvImage(qData, labels)
```

**Arguments**

|        |   |
|--------|---|
| qData  | A dataframe that contains quantitative data.                |
| labels | A vector of the conditions (labels) (one label per sample). |

**Value**

A heatmap

**Author(s)**

Samuel Wiczorek, Thomas Burger



**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
mvImage(qData, labels)
```

---

mvImputation

*Missing values imputation from a matrix*


---

**Description**

This method is a wrapper to the `imputeLCMD` package adapted to a matrix.

**Usage**

```
mvImputation(qData, method)
```

**Arguments**

`qData`                    A dataframe that contains quantitative data.  
`method`                    The imputation method to be used. Choices are QRILC, KNN, BPCA and MLE.

**Value**

The matrix imputed

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)[1:1000]
mvImputation(qData, "MLE")
```

---

mvPerLinesHisto

*Bar plot of missing values per lines*


---

**Description**

This method plots a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins).

**Usage**

```
mvPerLinesHisto(qData, samplesData, indLegend = "auto", showValues = FALSE)
```

**Arguments**

|             |  |
|-------------|--|
| qData       | A dataframe that contains the data to plot.                                    |
| samplesData | A dataframe which contains informations about the replicates.                  |
| indLegend   | The indice of the column name's in pData() tab                                 |
| showValues  | A logical that indicates wether numeric values should be drawn above the bars. |

**Value**

A bar plot

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
samplesData <- Biobase::pData(Exp1_R25_pept)
mvPerLinesHisto(qData, samplesData)
```

---

mvPerLinesHistoPerCondition

*Bar plot of missing values per lines and per condition*

---

**Description**

This method plots a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins) and per conditions.

**Usage**

```
mvPerLinesHistoPerCondition(qData, samplesData, indLegend = "auto",
  showValues = FALSE)
```

**Arguments**

|             |   |
|-------------|---|
| qData       | A dataframe that contains quantitative data.  |
| samplesData | A dataframe where lines correspond to samples and columns to the meta-data for those samples. |
| indLegend   | The indice of the column name's in pData() tab  |
| showValues  | A logical that indicates wether numeric values should be drawn above the bars.                |

**Value**

A bar plot

**Author(s)**

Samuel Wiczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
samplesData <- Biobase::pData(Exp1_R25_pept)
mvPerLinesHistoPerCondition(qData, samplesData)
```

---

mvPerLinesHistoPerCondition\_HC

*Bar plot of missing values per lines and per condition*

---

## Description

This method plots a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins) and per conditions. Same as the function [mvPerLinesHistoPerCondition](#) but uses the package `highcharter`.

## Usage

```
mvPerLinesHistoPerCondition_HC(qData, samplesData, indLegend = "auto",
  showValues = FALSE)
```

## Arguments

|             |   |
|-------------|---|
| qData       | A dataframe that contains quantitative data.  |
| samplesData | A dataframe where lines correspond to samples and columns to the meta-data for those samples. |
| indLegend   | The indice of the column name's in <code>pData()</code> tab                                   |
| showValues  | A logical that indicates wether numeric values should be drawn above the bars.                |

## Value

A bar plot

## Author(s)

Samuel Wieczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
samplesData <- Biobase::pData(Exp1_R25_pept)
mvPerLinesHistoPerCondition_HC(qData, samplesData)
```

---

mvPerLinesHisto\_HC      *Bar plot of missing values per lines using highcharter*

---

### Description

This method plots a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins).

### Usage

```
mvPerLinesHisto_HC(qData, samplesData, indLegend = "auto",
  showValues = FALSE)
```

### Arguments

|             |  |
|-------------|--|
| qData       | A dataframe that contains the data to plot.                                    |
| samplesData | A dataframe which contains informations about the replicates.                  |
| indLegend   | The indice of the column name's in pData() tab                                 |
| showValues  | A logical that indicates wether numeric values should be drawn above the bars. |

### Value

A bar plot

### Author(s)

Florence Combes, Samuel Wiczorek

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
samplesData <- Biobase::pData(Exp1_R25_pept)
mvPerLinesHisto_HC(qData, samplesData)
```

---

mvTypePlot      *Distribution of missing values with respect to intensity values*

---

### Description

This method plots a scatter plot which represents the distribution of missing values. The colors correspond to the different conditions (slot Label in in the dataset of class MSnSet). The x-axis represent the mean of intensity for one condition and one entity in the dataset (i. e. a protein) whereas the y-axis count the number of missing values for this entity and the considered condition. The data have been jittered for an easier vizualisation.

### Usage

```
mvTypePlot(qData, labels, threshold = 0, type = "MV")
```

**Arguments**

|           |   |
|-----------|---|
| qData     | A dataframe that contains quantitative data.  |
| labels    | A vector of the conditions (labels) (one label per sample).                             |
| threshold | An integer for the intensity that delimits MNAR and MCAR missing values.                |
| type      | A string to indicate whether to show nb of missing values (MV) or nb of values (values) |

**Value**

A scatter plot

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
mvTypePlot(qData, labels, threshold=0)
```

---

my\_hc\_chart

*Customised resetZoomButton of highcharts plots*


---

**Description**

Customise the resetZoomButton of highcharts plots.

**Usage**

```
my_hc_chart(hc, chartType, zoomType = "None")
```

**Arguments**

|           |  |
|-----------|--|
| hc        | A highcharter object                                 |
| chartType | The type of the plot                                 |
| zoomType  | The type of the zoom (one of "x", "y", "xy", "None") |

**Value**

A highchart plot

**Author(s)**

Samuel Wiczorek

**Examples**

```
library("highcharter")
hc <- highchart()
hc_chart(hc,type = "line")
hc_add_series(hc,data = c(29, 71, 40))
my_hc_ExportMenu(hc,filename='foo')
```

---

my\_hc\_ExportMenu

*Customised contextual menu of highcharts plots*

---

**Description**

Customise the contextual menu of highcharts plots.

**Usage**

```
my_hc_ExportMenu(hc, filename)
```

**Arguments**

|          |   |
|----------|---|
| hc       | A highcharter object                              |
| filename | The filename under which the plot has to be saved |

**Value**

A contextual menu for highcharts plots

**Author(s)**

Samuel Wiczorek

**Examples**

```
library("highcharter")
hc <- highchart()
hc_chart(hc,type = "line")
hc_add_series(hc,data = c(29, 71, 40))
my_hc_ExportMenu(hc,filename='foo')
```

---

|         |   |
|---------|---|
| nonzero | <i>Retrieve the indices of non-zero elements in sparse matrices</i> |
|---------|---|

---

**Description**

This function retrieves the indices of non-zero elements in sparse matrices of class dgCMatrix from package Matrix. Thi

**Usage**

```
nonzero(x)
```

**Arguments**

x                    A sparse matrix of class dgCMatrix

**Value**

A two-column matrix

**Author(s)**

Samuel Wieczorek

**Examples**

```
library(Matrix)
mat <- Matrix(c(0,0,0,0,0,1,0,0,1,1,0,0,0,0,1),nrow=5, byrow=TRUE, sparse=TRUE)
res <- nonzero(mat)
```

---

|            |                      |
|------------|----------------------|
| normalized | <i>Normalisation</i> |
|------------|----------------------|

---

**Description**

Provides several methods to normalize data from a matrix. They are organized in four main families : Strong Rescaling, Median Centering, Mean Centering, Mean CenteringScaling. For the first family, two sub-categories are available : the sum by columns and the quantiles method. For the three other families, two categories are available : "Overall" which means that the value for each protein (ie line in the expression data tab) is computed over all the samples ; "within conditions" which means that the value for each protein (ie line in the matrix) is computed condition by condition.

**Usage**

```
normalized(qData, labels, method, type = NULL, scaling = FALSE,
           quantile = 0.15)
```

**Arguments**

|          |   |
|----------|---|
| qData    | A dataframe that contains quantitative data.  |
| labels   | A vector of strings containing the column "Label" of the pData().   |
| method   | One of the following : Global Alignment, Quantile Centering, Mean Centering.  |
| type     | For the method "Global Alignment", the parameters are: "sum by columns": operates on the original scale (not the log2 one) and propose to normalize each abundance by the total abundance of the sample (so as to focus on the analyte proportions among each sample). "Alignment on all quantiles": proposes to align the quantiles of all the replicates; practically it amounts to replace abundances by order statistics. For the two other methods, the parameters are "overall" (shift all the sample distributions at once) or "within conditions" (shift the sample distributions within each condition at a time). |
| scaling  | A boolean that indicates if the variance of the data have to be forced to unit (variance reduction) or not.   |
| quantile | A float that corresponds to the quantile used to align the data.  |

**Value**

A matrix normalized

**Author(s)**

Samuel Wieczorek, Thomas Burger

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
qData <- Biobase::exprs(Exp1_R25_pept[1:1000])
labels <- Biobase::pData(Exp1_R25_pept[1:1000])["Label"]
normalizedD(qData, labels, "Quantile Centering", "within conditions", quantile = 0.15)
```

---

pepa.test

*PEptide based Protein differential Abundance test*

---

**Description**

This function is PEptide based Protein differential Abundance test

**Usage**

```
pepa.test(X, y, n1, n2, global = FALSE, use.lm = FALSE)
```

**Arguments**

|    |   |
|----|---|
| X  | Binary q x p design matrix for q peptides and p proteins. X_(ij)=1 if peptide i belongs to protein j, 0 otherwise.              |
| y  | q x n matrix representing the log intensities of q peptides among n MS samples.   |
| n1 | number of samples under condition 1. It is assumed that the first n1 columns of y correspond to observations under condition 1. |



|        |  |
|--------|--|
| n2     | number of samples under condition 2.   |
| global | if TRUE, the test statistic for each protein uses all residues, including the ones for peptides in different connected components. Can be much faster as it does not require to compute connected components. However the p-values are not well calibrated in this case, as it amounts to adding a ridge to the test statistic. Calibrating the p-value would require knowing the amplitude of the ridge, which in turns would require computing the connected components. |
| use.lm | if TRUE (and if global=FALSE), use lm() rather than the result in Proposition 1 to compute the test statistic  |

**Value**

A list of the following elements: llr: log likelihood ratio statistic (maximum likelihood version). llr.map: log likelihood ratio statistic (maximum a posteriori version). llr.pv: p-value for llr. llr.map.pv: p-value for llr.map. mse.h0: Mean squared error under H0 mse.h1: Mean squared error under H1 s: selected regularization hyperparameter for llr.map. wchi2: weight used to make llr.map chi2-distributed under H0.

**Author(s)**

Thomas Burger, Laurent Jacob

---

|             |   |
|-------------|---|
| pepAgregate | <i>Function agregate peptides to proteins</i> |
|-------------|---|

---

**Description**

Method to agregate with a method peptides to proteins on a MSnSet object (peptides)

**Usage**

```
pepAgregate(obj.pep, protID, method = "sum overall", matAdj = NULL,
            n = NULL)
```

**Arguments**

|         |  |
|---------|--|
| obj.pep | An object (peptides) of class MSnSet.  |
| protID  | The name of proteins ID column   |
| method  | The method used to aggregate the peptides into proteins. Values are "sum", "mean" or "sum on top n" : do the sum / mean of intensity on all peptides belonging to proteins. Default is "sum" |
| matAdj  | An adjacency matrix  |
| n       | The number of peptides considered for the aggregation.   |

**Value**

An object of class MSnSet with proteins

**Author(s)**

Alexia Dorffer, Samuel Wiczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
protID <- "Protein.group.IDs"
mat <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, TRUE)
pepAgregate(Exp1_R25_pept[1:1000], protID, "sum overall", mat)
```

---

proportionConRev\_HC *Barplot of proportion of contaminants and reverse*

---

## Description

Plots a barplot of proportion of contaminants and reverse. Same as the function `proportionConRev` but uses the package `highcharter`

## Usage

```
proportionConRev_HC(nBoth = 0, nCont = 0, nRev = 0, lDataset = 0)
```

## Arguments

|                       |  |
|-----------------------|--|
| <code>nBoth</code>    | The number of both contaminants and reverse identified in the dataset. |
| <code>nCont</code>    | The number of contaminants identified in the dataset.                  |
| <code>nRev</code>     | The number of reverse entities identified in the dataset.              |
| <code>lDataset</code> | The total length (number of rows) of the dataset                       |

## Value

A barplot

## Author(s)

Samuel Wiczorek

## Examples

```
proportionConRev_HC(10, 20, 100)
```

---

|             |  |
|-------------|--|
| rbindMSnset | <i>Similar to the function rbind but applies on two subsets of the same MSnSet object.</i> |
|-------------|--|

---

**Description**

Similar to the function rbind but applies on two subsets of the same MSnSet object.

**Usage**

```
rbindMSnset(df1 = NULL, df2)
```

**Arguments**

|     |   |
|-----|---|
| df1 | An object (or subset of) of class MSnSet. May be NULL |
| df2 | A subset of the same object as df1                    |

**Value**

An instance of class MSnSet.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R2_pept)
df1 <- Exp1_R2_pept[1:100]
df2 <- Exp1_R2_pept[200:250]
rbindMSnset(df1, df2)
```

---

|           |   |
|-----------|---|
| readExcel | <i>This function reads a sheet of an Excel file and put the data into a data.frame.</i> |
|-----------|---|

---

**Description**

This function reads a sheet of an Excel file and put the data into a data.frame.

**Usage**

```
readExcel(file, extension, sheet)
```

**Arguments**

|           |                             |
|-----------|-----------------------------|
| file      | The name of the Excel file. |
| extension | The extension of the file   |
| sheet     | The name of the sheet       |

**Value**

A data.frame

**Author(s)**

Samuel Wieczorek

---

|                |   |
|----------------|---|
| reIntroduceMEC | <i>Put back LAPALA into a MSnSet object</i> |
|----------------|---|

---

**Description**

This method is used to put back the LAPALA that have been identified previously

**Usage**

```
reIntroduceMEC(obj, MECIndex)
```

**Arguments**

obj                   An object of class MSnSet.  
MECIndex              A data.frame that contains index of MEC (see findMECBlock).

**Value**

The object obj where LAPALA have been reintroduced

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
lapala <- findMECBlock(obj)
obj <- wrapper.impute.detQuant(obj)
obj <- reIntroduceMEC(obj, lapala)
```

---

|             |   |
|-------------|---|
| removeLines | <i>Removes lines in the dataset based on a prefix string.</i> |
|-------------|---|

---

**Description**

This function removes lines in the dataset based on a prefix string.

**Usage**

```
removeLines(obj, idLine2Delete = NULL, prefix = NULL)
```

**Arguments**

|               |  |
|---------------|--|
| obj           | An object of class MSnSet.                                   |
| idLine2Delete | The name of the column that correspond to the data to filter |
| prefix        | A character string that is the prefix to find in the data    |

**Value**

An object of class MSnSet.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
removeLines(Exp1_R25_pept, "Potential.contaminant")
removeLines(Exp1_R25_pept, "Reverse")
```

---

|        |        |
|--------|--------|
| samLRT | xxxxxx |
|--------|--------|

---

**Description**

This function computes a regularized version of the likelihood ratio statistic. The regularization adds a user-input fudge factor  $s1$  to the variance estimator. This is straightforward when using a fixed effect model (cases 'numeric' and 'lm') but requires some more care when using a mixed model.

**Usage**

```
samLRT(lmm.res.h0, lmm.res.h1, cc, n, p, s1)
```

**Arguments**

|            |  |
|------------|--|
| lmm.res.h0 | a vector of object containing the estimates (used to compute the statistic) under H0 for each connected component. If the fast version of the estimator was used (as implemented in this package), lmm.res.h0 is a vector containing averages of squared residuals. If a fixed effect model was used, it is a vector of lm objects and if a mixed effect model was used it is a vector or lmer object. |
| lmm.res.h1 | similar to lmm.res.h0, a vector of object containing the estimates (used to compute the statistic) under H1 for each protein.  |
| cc         | a list containing the indices of peptides and proteins belonging to each connected component.  |
| n          | the number of samples used in the test   |
| p          | the number of proteins in the experiment   |
| s1         | the fudge factor to be added to the variance estimate  |

**Value**

llr.sam: a vector of numeric containing the regularized log likelihood ratio statistic for each protein.  
s: a vector containing the maximum likelihood estimate of the variance for the chosen model. When using the fast version of the estimator implemented in this package, this is the same thing as the input lmm.res.h1.  
lh1.sam: a vector of numeric containing the regularized log likelihood under H1 for each protein.  
lh0.sam: a vector of numeric containing the regularized log likelihood under H0 for each connected component.  
sample.sizes: a vector of numeric containing the sample size (number of biological samples times number of peptides) for each protein. This number is the same for all proteins within each connected component.

**Author(s)**

Thomas Burger, Laurent Jacob

---

saveParameters

*Saves the parameters of a tool in the pipeline of Prostar*

---

**Description**

Saves the parameters of a tool in the pipeline of Prostar

**Usage**

```
saveParameters(obj, name = NULL, l.params = NULL)
```

**Arguments**

|          |  |
|----------|--|
| obj      | An object of class MSnSet  |
| name     | The name of the tool. Available values are: "Norm, Imputation, anaDiff, GO-Analysis,Aggregation" |
| l.params | A list that contains the parameters  |

**Value**

An instance of class MSnSet.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
l.params=list(method="Global quantile alignment", type="overall")
saveParameters(Exp1_R25_pept, "Imputation",l.params)
```

---

scatterplotEnrichGO\_HC

*A dotplot that shows the result of a GO enrichment, using the package highcharter*

---

**Description**

A scatter plot of GO enrichment analysis

**Usage**

```
scatterplotEnrichGO_HC(ego, maxRes = 10, title = NULL)
```

**Arguments**

|        |  |
|--------|--|
| ego    | The result of the GO enrichment, provides either by the function <code>enrichGO</code> in DAPAR or the function <code>enrichGO</code> of the package <a href="#">clusterProfiler</a> |
| maxRes | The maximum number of categories to display in the plot  |
| title  | The title of the plot  |

**Value**

A dotplot

**Author(s)**

Samuel Wieczorek

---

|        |   |
|--------|---|
| setMEC | <i>Sets the MEC tag in the OriginOfValues</i> |
|--------|---|

---

**Description**

Sets the MEC tag in the OriginOfValues

**Usage**

```
setMEC(obj)
```

**Arguments**

|     |                           |
|-----|---------------------------|
| obj | An object of class MSnSet |
|-----|---------------------------|

**Value**

An instance of class MSnSet.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
setMEC(Exp1_R25_pept)
```

---

|                      |  |
|----------------------|--|
| StringBasedFiltering | <i>Removes lines in the dataset based on a prefix strings (contaminants, reverse or both).</i> |
|----------------------|--|

---

**Description**

This function removes lines in the dataset based on prefix strings (contaminants, reverse or both).

**Usage**

```
StringBasedFiltering(obj, idCont2Delete = NULL, prefix_Cont = NULL,
  idRev2Delete = NULL, prefix_Rev = NULL)
```

**Arguments**

|               |  |
|---------------|--|
| obj           | An object of class MSnSet.   |
| idCont2Delete | The name of the column that correspond to the contaminants to filter           |
| prefix_Cont   | A character string that is the prefix for the contaminants to find in the data |
| idRev2Delete  | The name of the column that correspond to the reverse data to filter           |
| prefix_Rev    | A character string that is the prefix for the reverse to find in the data      |



**Value**

An list of 4 items : obj : an object of class MSnSet in which the lines have been deleted deleted.both : an object of class MSnSet which contains the deleted lines corresponding to both contaminants and reverse, deleted.contaminants : n object of class MSnSet which contains the deleted lines corresponding to contaminants, deleted.reverse : an object of class MSnSet which contains the deleted lines corresponding to reverse,

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
StringBasedFiltering(Exp1_R25_pept, 'Potential.contaminant', '+', 'Reverse', '+')
```

---

StringBasedFiltering2 *Removes lines in the dataset based on a prefix strings.*

---

**Description**

This function removes lines in the dataset based on prefix strings.

**Usage**

```
StringBasedFiltering2(obj, cname = NULL, tag = NULL)
```

**Arguments**

|       |  |
|-------|--|
| obj   | An object of class MSnSet.   |
| cname | The name of the column that correspond to the line to filter                   |
| tag   | A character string that is the prefix for the contaminants to find in the data |

**Value**

An list of 4 items : obj : an object of class MSnSet in which the lines have been deleted deleted : an object of class MSnSet which contains the deleted lines

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
StringBasedFiltering2(Exp1_R25_pept, 'Potential.contaminant', '+')
```

---

|             |   |
|-------------|---|
| SumPeptides | <i>Compute the intensity of proteins with the sum of the intensities of their peptides.</i> |
|-------------|---|

---

**Description**

This function computes the intensity of proteins based on the sum of the intensities of their peptides.

**Usage**

```
SumPeptides(matAdj, expr)
```

**Arguments**

|        |  |
|--------|--|
| matAdj | An adjacency matrix in which lines and columns correspond respectively to peptides and proteins. |
| expr   | A matrix of intensities of peptides  |

**Value**

A matrix of intensities of proteins

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, FALSE)
SumPeptides(M, Biobase::exprs(Exp1_R25_pept[1:1000]))
```

---

|             |                        |
|-------------|------------------------|
| test.design | <i>Check if xxxxxx</i> |
|-------------|------------------------|

---

**Description**

This function check xxxxx

**Usage**

```
test.design(tab)
```

**Arguments**

|     |   |
|-----|---|
| tab | A data.frame which correspond to xxxxxx |
|-----|---|

**Value**

A list of two items

**Author(s)**

Thomas Burger, Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
test.design(Biobase::pData(Exp1_R25_pept)[,1:3])
```

---

|              |  |
|--------------|--|
| TopnPeptides | <i>Compute the intensity of proteins as the sum of the intensities of their n best peptides.</i> |
|--------------|--|

---

**Description**

This function computes the intensity of proteins as the sum of the intensities of their n best peptides.

**Usage**

```
TopnPeptides(matAdj, expr, n)
```

**Arguments**

|        |  |
|--------|--|
| matAdj | An adjacency matrix in which lines and columns correspond respectively to peptides and proteins. |
| expr   | A matrix of intensities of peptides  |
| n      | The maximum number of peptides used to aggregate a protein.                                      |

**Value**

A matrix of intensities of proteins

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
protID <- "Protein.group.IDs"
matAdj <- BuildAdjacencyMatrix(Exp1_R25_pept[1:1000], protID, FALSE)
TopnPeptides(matAdj, Biobase::exprs(Exp1_R25_pept[1:1000]), 3)
```

---

translatedRandomBeta *Generator of simulated values*

---

### Description

Generator of simulated values

### Usage

```
translatedRandomBeta(n, min, max, param1 = 3, param2 = 1)
```

### Arguments

|        |   |
|--------|---|
| n      | An integer which is the number of simulation (same as in rbeta) |
| min    | An integer that corresponds to the lower bound of the interval  |
| max    | An integer that corresponds to the upper bound of the interval  |
| param1 | An integer that is the first parameter of rbeta function.       |
| param2 | An integer that is second parameter of rbeta function.          |

### Value

A vector of n simulated values

### Author(s)

Thomas Burger

### Examples

```
translatedRandomBeta(1000, 5, 10, 1, 1)
```

---

univ\_AnnotDbPkg *Returns the totality of ENTREZ ID (gene id) of an OrgDb annotation package. Careful : org.Pf.plasmo.db : no ENTREZID but ORF*

---

### Description

Function to compute the "universe" argument for the enrich\_GO function, in case this latter should be the entire organism. Returns all the ID of the OrgDb annotation package for the corresponding organism.

### Usage

```
univ_AnnotDbPkg(orgdb)
```

### Arguments

|       |   |
|-------|---|
| orgdb | a Bioconductor OrgDb annotation package |
|-------|---|

**Value**

A vector of ENTREZ ID

**Author(s)**

Florence Combes

---

|             |   |
|-------------|---|
| violinPlotD | <i>Builds a violinplot from a dataframe</i> |
|-------------|---|

---

**Description**

ViolinPlot for quantitative proteomics data

**Usage**

```
violinPlotD(qData, dataForXAxis = NULL, labels = NULL,
            group2Color = "Condition")
```

**Arguments**

|              |   |
|--------------|---|
| qData        | A dataframe that contains quantitative data.  |
| dataForXAxis | A vector containing the types of replicates to use as X-axis. Available values are: Label, Bio.Rep, Bio.Rep and Tech.Rep. Default is "Label".                                   |
| labels       | A vector of the conditions (labels) (one label per sample).   |
| group2Color  | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A violinplot

**Author(s)**

Florence Combes, Samuel Wiczorek

**See Also**

[densityPlotD](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
library(vioplot)
qData <- Biobase::exprs(Exp1_R25_pept)
types <- c("Label", "Bio.Rep")
dataForXAxis <- Biobase::pData(Exp1_R25_pept)[, types]
labels <- Biobase::pData(Exp1_R25_pept)[, "Label"]
violinPlotD(qData, dataForXAxis, labels)
```

---

wrapper.boxPlotD      *Wrapper to the boxplotD function on an object MSnSet*

---

### Description

This function is a wrapper for using the boxPlotD function with objects of class MSnSet

### Usage

```
wrapper.boxPlotD(obj, dataForXAxis = "Label", group2Color = "Condition")
```

### Arguments

|              |   |
|--------------|---|
| obj          | An object of class MSnSet.  |
| dataForXAxis | A vector of strings containing the names of columns in pData() to print labels on X-axis (Default is "Label").  |
| group2Color  | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

### Value

A boxplot

### Author(s)

Florence Combes, Samuel Wiczorek

### See Also

[wrapper.densityPlotD](#)

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
types <- c("Label", "Bio.Rep")
wrapper.boxPlotD(Exp1_R25_pept, types)
```

---

wrapper.boxPlotD\_HC      *Wrapper to the boxplotD\_HC function on an object MSnSet*

---

### Description

This function is a wrapper for using the boxPlotD\_HC function with objects of class MSnSet

### Usage

```
wrapper.boxPlotD_HC(obj, dataForXAxis = "Label", group2Color = "Condition")
```

**Arguments**

|              |   |
|--------------|---|
| obj          | An object of class MSnSet.  |
| dataForXAxis | A vector of strings containing the names of columns in pData() to print labels on X-axis (Default is "Label").  |
| group2Color  | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A boxplot

**Author(s)**

Samuel Wieczorek

**See Also**

[wrapper.densityPlotD](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
types <- c("Label", "Bio.Rep")
wrapper.boxPlotD_HC(Exp1_R25_pept, types)
```

---

```
wrapper.compareNormalizationD
```

*Builds a plot from a dataframe*

---

**Description**

Wrapper to the function that plot to compare the quantitative proteomics data before and after normalization

**Usage**

```
wrapper.compareNormalizationD(objBefore, objAfter, labelsForLegend = NULL,
  indData2Show = NULL, group2Color = "Condition")
```

**Arguments**

|                 |   |
|-----------------|---|
| objBefore       | A dataframe that contains quantitative data before normalization.   |
| objAfter        | A dataframe that contains quantitative data after normalization.  |
| labelsForLegend | A vector of the conditions (labels) (one label per sample).   |
| indData2Show    | A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the dataframe qDataBefore.                                       |
| group2Color     | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A plot

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
objAfter <- wrapper.normalized(Exp1_R25_pept, "Quantile Centering",
"within conditions")
wrapper.compareNormalizationD(Exp1_R25_pept, objAfter, labels)
```

---

```
wrapper.compareNormalizationD_HC
```

*Builds a plot from a dataframe*

---

**Description**

Wrapper to the function that plot to compare the quantitative proteomics data before and after normalization. Same as the function [wrapper.compareNormalizationD](#) but uses the package `highcharter`

**Usage**

```
wrapper.compareNormalizationD_HC(objBefore, objAfter, labelsForLegend = NULL,
indData2Show = NULL, group2Color = "Condition")
```

**Arguments**

|                              |   |
|------------------------------|---|
| <code>objBefore</code>       | A dataframe that contains quantitative data before normalization.   |
| <code>objAfter</code>        | A dataframe that contains quantitative data after normalization.  |
| <code>labelsForLegend</code> | A vector of the conditions (labels) (one label per sample).   |
| <code>indData2Show</code>    | A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data.frame <code>qDataBefore</code> .                        |
| <code>group2Color</code>     | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A plot

**Author(s)**

Samuel Wieczorek



## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
objAfter <- wrapper.normalized(Exp1_R25_pept, "Quantile Centering",
"within conditions")
wrapper.compareNormalizationD_HC(Exp1_R25_pept, objAfter, labels)
```

---

wrapper.corrMatrixD *Displays a correlation matrix of the quantitative data of the exprs() table*

---

## Description

Builds a correlation matrix based on a MSnSet object.

## Usage

```
wrapper.corrMatrixD(obj, rate = 5)
```

## Arguments

obj            An object of class MSnSet.  
rate           A float that defines the gradient of colors.

## Value

A colored correlation matrix

## Author(s)

Alexia Dorffer

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.corrMatrixD(Exp1_R25_pept)
```

wrapper.corrMatrixD\_HC

*Displays a correlation matrix of the quantitative data of the exprs() table*

---

### Description

Builds a correlation matrix based on a MSnSet object. Same as the function [wrapper.corrMatrixD](#) but uses the package highcharter

### Usage

```
wrapper.corrMatrixD_HC(obj, rate = 0.5)
```

### Arguments

obj                    An object of class MSnSet.  
rate                   A float that defines the gradient of colors.

### Value

A colored correlation matrix

### Author(s)

Samuel Wieczorek

### Examples

```
require(DAPARdata)  
data(Exp1_R25_pept)  
wrapper.corrMatrixD_HC(Exp1_R25_pept)
```

---

wrapper.CVDistD

*Distribution of CV of entities*

---

### Description

Builds a densityplot of the CV of entities in the exprs() table of an object MSnSet. The variance is calculated for each condition (Label) present in the dataset (see the slot 'Label' in the pData() table).

### Usage

```
wrapper.CVDistD(obj)
```

### Arguments

obj                    An object of class MSnSet.

**Value**

A density plot

**Author(s)**

Alexia Dorffer

**See Also**

[wrapper.densityPlotD](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.CVDistD(Exp1_R25_pept)
```

---

wrapper.CVDistD\_HC      *Distribution of CV of entities*

---

**Description**

Builds a densityplot of the CV of entities in the `exprs()` table. of an object `MSnSet`. The variance is calculated for each condition (Label) present in the dataset (see the slot 'Label' in the `pData()` table). Same as the function `wrapper.CVDistD` but uses the package `highcharter`

**Usage**

```
wrapper.CVDistD_HC(obj)
```

**Arguments**

`obj`                      An object of class `MSnSet`.

**Value**

A density plot

**Author(s)**

Samuel Wieczorek

**See Also**

[wrapper.densityPlotD](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.CVDistD_HC(Exp1_R25_pept)
```

---

 wrapper.dapar.impute.mi

*Missing values imputation using the LSImpute algorithm.*


---

### Description

This method is a wrapper to the function `impute.mi` of the package `imp4p` adapted to an object of class `MSnSet`.

### Usage

```
wrapper.dapar.impute.mi(obj, nb.iter = 3, nknn = 15, selec = 600,
  siz = 500, weight = 1, ind.comp = 1, progress.bar = TRUE,
  x.step.mod = 300, x.step.pi = 300, nb.rei = 100, method = 4,
  gridsize = 300, q = 0.95, q.min = 0, q.norm = 3, eps = 0,
  methodi = "slsa", lapala = TRUE, distribution = "unif")
```

### Arguments

|                           |  |
|---------------------------|--|
| <code>obj</code>          | An object of class <code>MSnSet</code> .   |
| <code>nb.iter</code>      | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>nknn</code>         | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>selec</code>        | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>siz</code>          | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>weight</code>       | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>ind.comp</code>     | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>progress.bar</code> | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>x.step.mod</code>   | Same as the function <code>estim.mix</code> in the package <code>imp4p</code>                |
| <code>x.step.pi</code>    | Same as the function <code>estim.mix</code> in the package <code>imp4p</code>                |
| <code>nb.rei</code>       | Same as the function <code>estim.mix</code> in the package <code>imp4p</code>                |
| <code>method</code>       | Same as the function <code>estim.mix</code> in the package <code>imp4p</code>                |
| <code>gridsize</code>     | Same as the function <code>estim.mix</code> in the package <code>imp4p</code>                |
| <code>q</code>            | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>q.min</code>        | Same as the function <code>impute.pa</code> in the package <code>imp4p</code>                |
| <code>q.norm</code>       | Same as the function <code>impute.pa</code> in the package <code>imp4p</code>                |
| <code>eps</code>          | Same as the function <code>impute.pa</code> in the package <code>imp4p</code>                |
| <code>methodi</code>      | Same as the function <code>mi.mix</code> in the package <code>imp4p</code>                   |
| <code>lapala</code>       | xxxxxxxxxxx  |
| <code>distribution</code> | The type of distribution used. Values are <code>unif</code> (default) or <code>beta</code> . |

### Value

The `exprs(obj)` matrix with imputed values instead of missing values.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
dat <- mvFilter(Exp1_R25_pept[1:1000], type="allCond", th = 1)
dat <- wrapper.dapar.impute.mi(dat, nb.iter=1)
```

---

wrapper.densityPlotD *Builds a densityplot from an object of class MSnSet*

---

**Description**

This function is a wrapper for using the densityPlotD function with objects of class MSnSet

**Usage**

```
wrapper.densityPlotD(obj, labelsForLegend = NULL, indData2Show = NULL,
  group2Color = "Condition")
```

**Arguments**

|                 |   |
|-----------------|---|
| obj             | An object of class MSnSet.  |
| labelsForLegend | A vector of labels to show in densityplot.  |
| indData2Show    | A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data frame qDataBefore in the density plot.                  |
| group2Color     | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A density plot

**Author(s)**

Alexia Dorffer

**See Also**

[wrapper.boxPlotD](#), [wrapper.CVDistD](#)

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]
wrapper.densityPlotD(Exp1_R25_pept, labels)
```

---

`wrapper.densityPlotD_HC`*Builds a densityplot from an object of class MSnSet*

---

### Description

This function is a wrapper for using the `densityPlotD` function with objects of class `MSnSet`. Same as the function `wrapper.densityPlotD` but uses the package `highcharter`

### Usage

```
wrapper.densityPlotD_HC(obj, labelsForLegend = NULL, indData2Show = NULL,  
  group2Color = "Condition")
```

### Arguments

|                              |   |
|------------------------------|---|
| <code>obj</code>             | An object of class <code>MSnSet</code> .  |
| <code>labelsForLegend</code> | A vector of labels to show in densityplot.  |
| <code>indData2Show</code>    | A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data frame <code>qDataBefore</code> in the density plot.     |
| <code>group2Color</code>     | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

### Value

A density plot

### Author(s)

Samuel Wieczorek

### See Also

[wrapper.boxPlotD](#), [wrapper.CVDistD](#)

### Examples

```
require(DAPARdata)  
data(Exp1_R25_pept)  
labels <- Biobase::pData(Exp1_R25_pept)[,"Label"]  
wrapper.densityPlotD_HC(Exp1_R25_pept, labels)
```

---

`wrapper.hc_mvTypePlot2`

*Distribution of observed values with respect to intensity values from a MSnSet object*

---

### Description

This method is a wrapper for the function [hc\\_mvTypePlot2](#) adapted to objects of class MSnSet).

### Usage

```
wrapper.hc_mvTypePlot2(obj)
```

### Arguments

`obj` An object of class MSnSet.  
... See [hc\\_mvTypePlot2](#)

### Value

A scatter plot

### Author(s)

Florence Combes, Samuel Wiczorek

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.hc_mvTypePlot2(Exp1_R25_pept)
```

---

`wrapper.heatmapD`

*This function is a wrapper to [heatmap.2](#) that displays quantitative data in the `exprs()` table of an object of class MSnSet*

---

### Description

Builds a heatmap of the quantitative proteomic data of a MSnSet object.

### Usage

```
wrapper.heatmapD(obj, distance = "euclidean", cluster = "complete",
  dendro = FALSE)
```

**Arguments**

|          |   |
|----------|---|
| obj      | An object of class MSnSet.  |
| distance | The distance used by the clustering algorithm to compute the dendrogram. See <code>help(heatmap.2)</code> . |
| cluster  | the clustering algorithm used to build the dendrogram. See <code>help(heatmap.2)</code>                     |
| dendro   | A boolean to indicate if the dendrogram has to be displayed   |

**Value**

A heatmap

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- mvFilter(Exp1_R25_pept[1:1000], "wholeMatrix", 6)
wrapper.heatmapD(obj)
```

---

```
wrapper.impute.detQuant
```

*Wrapper of the function `impute.detQuant` for objects of class MSnSet*

---

**Description**

This method is a wrapper of the function `impute.detQuant` for objects of class MSnSet

**Usage**

```
wrapper.impute.detQuant(obj, qval = 0.025, factor = 1)
```

**Arguments**

|        |  |
|--------|--|
| obj    | An instance of class MSnSet  |
| qval   | An expression set containing quantitative values of various replicates |
| factor | A scaling factor to multiply the imputation value with                 |

**Value**

An imputed instance of class MSnSet

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.impute.detQuant(Exp1_R25_pept)
```



---

```
wrapper.impute.fixedValue
```

*Missing values imputation from a MSnSet object*

---

### Description

This method is a wrapper to objects of class MSnSet and imputes missing values with a fixed value.

### Usage

```
wrapper.impute.fixedValue(obj, fixVal)
```

### Arguments

|        |                            |
|--------|----------------------------|
| obj    | An object of class MSnSet. |
| fixVal | A float .                  |

### Value

The object obj which has been imputed

### Author(s)

Samuel Wieczorek

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.impute.fixedValue(Exp1_R25_pept[1:1000], 0.001)
```

---

```
wrapper.impute.KNN
```

*KNN missing values imputation from a MSnSet object*

---

### Description

This method is a wrapper for objects of class MSnSet and imputes missing values with a fixed value. This function imputes the missing values condition by condition.

### Usage

```
wrapper.impute.KNN(obj, K)
```

### Arguments

|     |                            |
|-----|----------------------------|
| obj | An object of class MSnSet. |
| K   | the number of neighbors.   |

### Value

The object obj which has been imputed

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.impute.KNN(Exp1_R25_pept[1:1000], 3)
```

---

wrapper.impute.pa      *Imputation of peptides having no values in a biological condition.*

---

**Description**

This method is a wrapper to the function `impute.pa` of the package `imp4p` adapted to an object of class `MSnSet`.

**Usage**

```
wrapper.impute.pa(obj, q.min = 0.025)
```

**Arguments**

`obj`                    An object of class `MSnSet`.  
`q.min`                 Same as the function `impute.pa` in the package `imp4p`

**Value**

The `exprs(obj)` matrix with imputed values instead of missing values.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
dat <- mvFilter(Exp1_R25_pept[1:1000], type="allCond", th = 1)
dat <- wrapper.impute.pa(dat)
```

---

wrapper.impute.pa2      *Missing values imputation from a MSnSet object*

---

### Description

This method is a wrapper to the function `impute.pa` from the package `imp4p` adapted to objects of class `MSnSet`.

### Usage

```
wrapper.impute.pa2(obj, q.min = 0, q.norm = 3, eps = 0,  
  distribution = "unif")
```

### Arguments

|                           |   |
|---------------------------|---|
| <code>obj</code>          | An object of class <code>MSnSet</code> .  |
| <code>q.min</code>        | A quantile value of the observed values allowing defining the maximal value which can be generated. This maximal value is defined by the quantile <code>q.min</code> of the observed values distribution minus <code>eps</code> . Default is 0 (the maximal value is the minimum of observed values minus <code>eps</code> ). |
| <code>q.norm</code>       | A quantile value of a normal distribution allowing defining the minimal value which can be generated. Default is 3 (the minimal value is the maximal value minus <code>qn*median(sd(observed values))</code> where <code>sd</code> is the standard deviation of a row in a condition).  |
| <code>eps</code>          | A value allowing defining the maximal value which can be generated. This maximal value is defined by the quantile <code>q.min</code> of the observed values distribution minus <code>eps</code> . Default is 0.   |
| <code>distribution</code> | The type of distribution used. Values are <code>unif</code> (default) or <code>beta</code> .  |

### Value

The object `obj` which has been imputed

### Author(s)

Thomas Burger, Samuel Wiczorek

### Examples

```
require(DAPARdata)  
data(Exp1_R25_pept)  
wrapper.impute.pa2(Exp1_R25_pept[1:1000], distribution="beta")
```

---

wrapper.impute.slsa     *Imputation of peptides having no values in a biological condition.*

---

**Description**

This method is a wrapper to the function `impute.slsa` of the package `imp4p` adapted to an object of class `MSnSet`.

**Usage**

```
wrapper.impute.slsa(obj)
```

**Arguments**

`obj`                    An object of class `MSnSet`.

**Value**

The `exprs(obj)` matrix with imputed values instead of missing values.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
dat <- mvFilter(Exp1_R25_pept[1:1000], type="allCond", th = 1)
dat <- wrapper.impute.slsa(dat)
```

---

wrapper.mvHisto             *Histogram of missing values from a MSnSet object*

---

**Description**

This method plots from a `MSnSet` object a histogram of missing values.

**Usage**

```
wrapper.mvHisto(obj, indLegend = "auto", showValues = FALSE)
```

**Arguments**

`obj`                    An object of class `MSnSet`.  
`indLegend`            The indices of the column name's in `pData()` tab.  
`showValues`           A logical that indicates wether numeric values should be drawn above the bars.

**Value**

A histogram

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.mvHisto(Exp1_R25_pept, showValues=TRUE)
```

---

wrapper.mvHisto\_HC      *Histogram of missing values from a MSnSet object*

---

**Description**

This method plots from a MSnSet object a histogram of missing values.

**Usage**

```
wrapper.mvHisto_HC(obj, indLegend = "auto", showValues = FALSE)
```

**Arguments**

|            |  |
|------------|--|
| obj        | An object of class MSnSet.   |
| indLegend  | The indices of the column name's in pData() tab.                               |
| showValues | A logical that indicates wether numeric values should be drawn above the bars. |

**Value**

A histogram

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.mvHisto(Exp1_R25_pept, showValues=TRUE)
```

---

|                 |   |
|-----------------|---|
| wrapper.mvImage | <i>Heatmap of missing values from a MSnSet object</i> |
|-----------------|---|

---

**Description**

Plots a heatmap of the quantitative data. Each column represent one of the conditions in the object of class MSnSet and the color is proportional to the mean of intensity for each line of the dataset. The lines have been sorted in order to visualize easily the different number of missing values. A white square is plotted for missing values.

**Usage**

```
wrapper.mvImage(obj)
```

**Arguments**

obj                    An object of class MSnSet.

**Value**

A heatmap

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.mvImage(Exp1_R25_pept)
```

---

|                      |   |
|----------------------|---|
| wrapper.mvImputation | <i>Missing values imputation from a MSnSet object</i> |
|----------------------|---|

---

**Description**

This method is a wrapper to the `imputeLCMD` package adapted to objects of class MSnSet.

**Usage**

```
wrapper.mvImputation(obj, method)
```

**Arguments**

obj                    An object of class MSnSet.  
method                The imputation method to be used. Choices are QRILC, KNN, BPCA and MLE.

**Value**

The object obj which has been imputed

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.mvImputation(Exp1_R25_pept[1:1000], "MLE")
```

---

`wrapper.mvPerLinesHisto`*Histogram of missing values per lines from an object MSnSet*

---

**Description**

This method is a wrapper to plots from a MSnSet object a histogram which represents the distribution of the number of missing values (NA) per lines (ie proteins).

**Usage**

```
wrapper.mvPerLinesHisto(obj, indLegend = "auto", showValues = FALSE)
```

**Arguments**

|                         |  |
|-------------------------|--|
| <code>obj</code>        | An object of class MSnSet.   |
| <code>indLegend</code>  | The indice of the column name's in <code>pData()</code> tab .                  |
| <code>showValues</code> | A logical that indicates wether numeric values should be drawn above the bars. |

**Value**

A histogram

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.mvPerLinesHisto(Exp1_R25_pept)
```

---

```
wrapper.mvPerLinesHistoPerCondition
```

*Bar plot of missing values per lines and per conditions from an object MSnSet*

---

### Description

This method is a wrapper to plots from a MSnSet object a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins) and per conditions.

### Usage

```
wrapper.mvPerLinesHistoPerCondition(obj, indLegend = "auto",  
  showValues = FALSE)
```

### Arguments

|            |  |
|------------|--|
| obj        | An object of class MSnSet.   |
| indLegend  | The indice of the column name's in pData() tab .                               |
| showValues | A logical that indicates wether numeric values should be drawn above the bars. |

### Value

A bar plot

### Author(s)

Samuel Wieczorek

### Examples

```
require(DAPARdata)  
data(Exp1_R25_pept)  
wrapper.mvPerLinesHistoPerCondition(Exp1_R25_pept)
```

---

```
wrapper.mvPerLinesHistoPerCondition_HC
```

*Bar plot of missing values per lines and per conditions from an object MSnSet*

---

### Description

This method is a wrapper to plots (using highcharts) from a MSnSet object a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins) and per conditions.

### Usage

```
wrapper.mvPerLinesHistoPerCondition_HC(obj, indLegend = "auto",  
  showValues = FALSE)
```



**Arguments**

|            |  |
|------------|--|
| obj        | An object of class MSnSet.   |
| indLegend  | The indice of the column name's in pData() tab .                               |
| showValues | A logical that indicates wether numeric values should be drawn above the bars. |

**Value**

A bar plot

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.mvPerLinesHistoPerCondition(Exp1_R25_pept)
```

---

```
wrapper.mvPerLinesHisto_HC
```

*Histogram of missing values per lines from an object using highcharacter MSnSet*

---

**Description**

This method is a wrapper to plots from a MSnSet object a histogram which represents the distribution of the number of missing values (NA) per lines (ie proteins).

**Usage**

```
wrapper.mvPerLinesHisto_HC(obj, indLegend = "auto", showValues = FALSE)
```

**Arguments**

|            |  |
|------------|--|
| obj        | An object of class MSnSet.   |
| indLegend  | The indice of the column name's in pData() tab .                               |
| showValues | A logical that indicates wether numeric values should be drawn above the bars. |

**Value**

A histogram

**Author(s)**

Alexia Dorffer

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.mvPerLinesHisto(Exp1_R25_pept)
```

---

|                    |   |
|--------------------|---|
| wrapper.mvTypePlot | <i>Distribution of missing values with respect to intensity values from a MSnSet object</i> |
|--------------------|---|

---

### Description

This method plots a scatter plot which represents the distribution of missing values. The colors correspond to the different conditions (slot Label in in the dataset of class MSnSet). The x-axis represent the mean of intensity for one condition and one entity in the dataset (i. e. a protein) whereas the y-axis count the number of missing values for this entity and the considered condition. The data have been jittered for an easier vizualisation.

### Usage

```
wrapper.mvTypePlot(obj, ...)
```

### Arguments

|     |                                |
|-----|--------------------------------|
| obj | An object of class MSnSet.     |
| ... | See <a href="#">mvTypePlot</a> |

### Value

A scatter plot

### Author(s)

Florence Combes, Samuel Wieczorek

### Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.mvTypePlot(Exp1_R25_pept)
```

---

|                    |                      |
|--------------------|----------------------|
| wrapper.normalized | <i>Normalisation</i> |
|--------------------|----------------------|

---

### Description

Provides several methods to normalize quantitative data from a MSnSet object. They are organized in four main families : Strong Rescaling, Median Centering, Mean Centering, Mean CenteringScaling. For the first family, two sub-categories are available : the sum by columns and the quantiles method. For the three other families, two categories are available : "Overall" which means that the value for each protein (ie line in the expression data tab) is computed over all the samples ; "within conditions" which means that the value for each protein (ie line in the exprs() data tab) is computed condition by condition.

**Usage**

```
wrapper.normalized(obj, method, type = NULL, scaling = FALSE,
  quantile = 0.15)
```

**Arguments**

|          |  |
|----------|--|
| obj      | An object of class MSnSet.   |
| method   | One of the following : Global Alignment (for normalizations of important magnitude), Quantile Centering, Mean Centering.   |
| type     | For the method "Global Alignment", the parameters are: "sum by columns": operates on the original scale (not the log2 one) and propose to normalize each abundance by the total abundance of the sample (so as to focus on the analyte proportions among each sample). "Alignment on all quantiles": proposes to align the quantiles of all the replicates; practically it amounts to replace abundances by order statistics. For the two other methods, the parameters are "over-all" (shift all the sample distributions at once) or "within conditions" (shift the sample distributions within each condition at a time). |
| scaling  | A boolean that indicates if the variance of the data have to be forced to unit (variance reduction) or not.  |
| quantile | A float that corresponds to the quantile used to align the data.   |

**Value**

An instance of class MSnSet where the quantitative data in the exprs() tab has been normalized.

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
wrapper.normalized(Exp1_R25_pept[1:1000], "Quantile Centering", "within conditions")
```

---

```
wrapper.t_test_Complete
      xxxxx
```

---

**Description**

This function is a wrapper xxxxx

**Usage**

```
wrapper.t_test_Complete(obj, ...)
```

**Arguments**

|     |  |
|-----|--|
| obj | An object of class MSnSet with no missing values |
| ... | See compute.t.tests                              |

**Value**

xxxxxxx

**Author(s)**

Samuel Wiczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
lapala <- findMECBlock(obj)
obj <- wrapper.impute.detQuant(obj)
obj <- reIntroduceMEC(obj, lapala)
obj <- wrapper.impute.detQuant(obj)
ttest <- wrapper.t_test_Complete(obj, 1)
```

---

wrapper.violinPlotD    *Wrapper to the violinPlotD function on an object MSnSet*

---

**Description**

This function is a wrapper for using the violinPlotD function with objects of class MSnSet

**Usage**

```
wrapper.violinPlotD(obj, dataForXAxis = "Label", group2Color = "Condition")
```

**Arguments**

|              |   |
|--------------|---|
| obj          | An object of class MSnSet.  |
| dataForXAxis | A vector of strings containing the names of columns in pData() to print labels on X-axis (Default is "Label").  |
| group2Color  | A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition. |

**Value**

A violin plot

**Author(s)**

Samuel Wiczorek

**See Also**

[wrapper.densityPlotD](#), [wrapper.boxPlotD](#)

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
library(vioplot)
types <- c("Label", "Bio.Rep")
wrapper.violinPlotD(Exp1_R25_pept, types)
```

---

wrapperCalibrationPlot

*Performs a calibration plot on an MSnSet object, calling the cp4p package functions.*

---

## Description

This function is a wrapper to the calibration.plot method of the cp4p package for use with MSnSet objects.

## Usage

```
wrapperCalibrationPlot(vPVal, pi0Method = "pounds")
```

## Arguments

|           |   |
|-----------|---|
| vPVal     | A dataframe that contains quantitative data.                |
| pi0Method | A vector of the conditions (labels) (one label per sample). |

## Value

A plot

## Author(s)

Samuel Wieczorek

## Examples

```
require(DAPARdata)
data(Exp1_R25_pept)
obj <- Exp1_R25_pept[1:1000]
keepThat <- mvFilterGetIndices(obj, 'wholeMatrix', ncol(obj))
obj <- mvFilterFromIndices(obj, keepThat)
qData <- Biobase::exprs(obj)
sTab <- Biobase::pData(obj)
limma <- limmaCompleteTest(qData, sTab)
wrapperCalibrationPlot(limma$P_Value[,1])
```

---

|                  |   |
|------------------|---|
| writeMSnsetToCSV | <i>Exports a MSnset dataset into a zip archive containing three zipped CSV files.</i> |
|------------------|---|

---

**Description**

This function exports a MSnset dataset into three csv files compressed in a zip file

**Usage**

```
writeMSnsetToCSV(obj, fname)
```

**Arguments**

|       |                               |
|-------|-------------------------------|
| obj   | An object of class MSnSet.    |
| fname | The name of the archive file. |

**Value**

A compressed file

**Author(s)**

Samuel Wieczorek

**Examples**

```
require(DAPARdata)
data(Exp1_R2_pept)
obj <- Exp1_R2_pept[1:1000]
writeMSnsetToCSV(obj, "foo")
```

---

|                    |   |
|--------------------|---|
| writeMSnsetToExcel | <i>This function exports a MSnSet object to a Excel file.</i> |
|--------------------|---|

---

**Description**

This function exports a MSnSet data object to a Excel file. Each of the three data.frames in the MSnSet object (ie experimental data, phenoData and metaData are respectively integrated into separate sheets in the Excel file). The colored cells in the experimental data correspond to the original missing values which have been imputed.

**Usage**

```
writeMSnsetToExcel(obj, filename)
```

**Arguments**

`obj` An object of class MSnSet.  
`filename` A character string for the name of the Excel file.

**Value**

A Excel file (.xlsx)

**Author(s)**

Samuel Wieczorek

**Examples**

```
Sys.setenv("R_ZIPCMD"= Sys.which("zip"))
require(DAPARdata)
data(Exp1_R2_pept)
obj <- Exp1_R2_pept[1:1000]
writeMSnsetToExcel(obj, "foo")
```

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