

Introduction to RBM package

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1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code. Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

| | Length | Class | Mode |
|----------------------------|--------|--------|---------|
| <code>ordfit_t</code> | 1000 | -none- | numeric |
| <code>ordfit_pvalue</code> | 1000 | -none- | numeric |
| <code>ordfit_beta0</code> | 1000 | -none- | numeric |
| <code>ordfit_beta1</code> | 1000 | -none- | numeric |
| <code>permutation_p</code> | 1000 | -none- | numeric |
| <code>bootstrap_p</code> | 1000 | -none- | numeric |

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 18

> which(myresult$permutation_p<=0.05)

[1] 23 40 117 146 212 221 296 410 453 467 484 512 521 633 788 874 915 970

> sum(myresult$bootstrap_p<=0.05)

[1] 3

> which(myresult$bootstrap_p<=0.05)

[1] 265 360 510

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 0

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 43

> which(myresult2$bootstrap_p<=0.05)

[1] 35 42 44 58 133 154 192 203 223 229 238 249 323 332 404 420 436 443 447
[20] 457 465 481 514 542 582 649 684 719 721 738 741 757 802 823 862 874 879 883
[39] 891 900 913 943 962

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000  -none-  numeric
ordfit_pvalue 3000  -none-  numeric
ordfit_beta1  3000  -none-  numeric
permutation_p 3000  -none-  numeric
bootstrap_p   3000  -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 77

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 72

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 68

> which(myresult_F$permutation_p[, 1]<=0.05)

[1]  4 28 43 85 87 93 139 149 150 152 159 182 188 206 217 222 231 235 255
[20] 271 274 283 290 296 299 333 349 383 393 398 403 429 493 507 514 527 528 532
[39] 557 564 588 604 629 630 633 656 657 669 696 698 726 728 733 740 744 750 762
[58] 802 808 811 826 835 845 853 860 868 878 881 889 891 896 901 925 944 966 978
[77] 990

> which(myresult_F$permutation_p[, 2]<=0.05)

[1]  6 28 43 57 85 87 93 139 150 152 159 174 182 206 213 217 231 235 255
[20] 271 274 283 290 299 324 333 349 383 386 393 398 403 429 434 493 514 527 528
[39] 532 557 604 629 630 633 656 657 696 698 717 728 733 740 750 756 808 811 835
[58] 845 853 860 867 868 878 881 889 891 896 904 913 924 950 978

> which(myresult_F$permutation_p[, 3]<=0.05)

[1]  6 66 93 117 139 142 149 150 159 174 188 206 217 231 233 235 255 271 274
[20] 290 299 324 333 349 386 393 398 403 429 434 493 514 528 557 564 585 604 629
[39] 630 633 656 657 669 679 696 698 728 740 762 802 808 811 835 845 853 860 868
[58] 881 889 896 901 906 913 925 950 954 966 978

```

```

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 16

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 14

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 14

> which(con2_adjp<=0.05/3)

[1] 159 235 274 290 514 557 629 633 728 808 853 860 889 978

> which(con3_adjp<=0.05/3)

[1] 159 235 255 274 290 333 514 629 657 728 740 808 860 896

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1  3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p   3000   -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 50

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 41

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 47

```

```

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 63 83 84 85 126 154 175 181 184 215 224 239 252 282 284 285 367 372 383
[20] 388 412 419 447 457 461 472 515 553 571 598 600 604 636 642 650 653 686 697
[39] 703 766 773 784 879 884 893 909 918 968 971 988

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 63 83 84 126 181 184 224 226 252 255 282 367 372 383 419 447 461 472 483
[20] 515 553 571 600 636 650 653 663 686 697 703 766 784 811 821 879 884 893 900
[39] 909 924 962

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 21 63 83 84 126 175 184 224 226 252 282 367 372 378 383 388 412 419 447
[20] 472 501 515 553 571 598 600 636 642 650 653 663 686 703 766 773 784 794 821
[39] 827 879 884 893 909 918 921 924 988

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 4

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 1

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 5

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "/tmp/RtmpfnMHPG/Rinst17b2dc2c71ea67/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

      IlmnID      Beta      exmdata2[, 2]      exmdata3[, 2]
cg00000292: 1  Min.   :0.01058  Min.   :0.01187  Min.   :0.009103
cg00002426: 1  1st Qu.:0.04111  1st Qu.:0.04407  1st Qu.:0.041543
cg00003994: 1  Median :0.08284  Median :0.09531  Median :0.087042
cg00005847: 1  Mean   :0.27397  Mean   :0.28872  Mean   :0.283729
cg00006414: 1  3rd Qu.:0.52135  3rd Qu.:0.59032  3rd Qu.:0.558575
cg00007981: 1  Max.   :0.97069  Max.   :0.96937  Max.   :0.970155
(Other)    :994      NA's    :4
exmdata4[, 2]  exmdata5[, 2]  exmdata6[, 2]  exmdata7[, 2]
Min.   :0.01019  Min.   :0.01108  Min.   :0.01937  Min.   :0.01278
1st Qu.:0.04092  1st Qu.:0.04059  1st Qu.:0.05060  1st Qu.:0.04260
Median :0.09042  Median :0.08527  Median :0.09502  Median :0.09362
Mean   :0.28508  Mean   :0.28482  Mean   :0.27348  Mean   :0.27563
3rd Qu.:0.57502  3rd Qu.:0.57300  3rd Qu.:0.52099  3rd Qu.:0.52240
Max.   :0.96658  Max.   :0.97516  Max.   :0.96681  Max.   :0.95974
      NA's    :1
exmdata8[, 2]
Min.   :0.01357
1st Qu.:0.04387
Median :0.09282
Mean   :0.28679
3rd Qu.:0.57217
Max.   :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

      Length Class  Mode
ordfit_t      1000  -none- numeric
ordfit_pvalue 1000  -none- numeric
ordfit_beta0  1000  -none- numeric
ordfit_beta1  1000  -none- numeric
permutation_p 1000  -none- numeric
bootstrap_p   1000  -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)

[1] 47

```

```

> sum(diff_results$permutation_p<=0.05)

[1] 54

> sum(diff_results$bootstrap_p<=0.05)

[1] NA

> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)

[1] 0

> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)

[1] 3

> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)

[1] NA

> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_perm, ])
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
103 cg00094319 0.7378428 0.73532960 0.7557490 0.73830220
259 cg00234961 0.0419217 0.04321576 0.0570714 0.05327565
851 cg00830029 0.5836250 0.59397870 0.6473961 0.67269640
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
103 0.67349260 0.73510200 0.75715920 0.78981220
259 0.04030003 0.03996053 0.05086962 0.05445672
851 0.50820240 0.34657470 0.66276570 0.64634510
      diff_results$ordfit_t[diff_list_perm]
103 -2.343784
259 -2.833203
851 -2.986319
      diff_results$permutation_p[diff_list_perm]
103 0
259 0
851 0

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_boot)

```


| | IlmnID | Beta | exmdata2[, 2] | exmdata3[, 2] | exmdata4[, 2] |
|---|---------------|---------------|---------------|---------------|---------------|
| 146 | cg00134539 | 0.61101320 | 0.53321780 | 0.45999340 | 0.46787420 |
| 189 | cg00176210 | 0.28756520 | 0.39161870 | 0.44272520 | 0.44725330 |
| 280 | cg00260778 | 0.64319890 | 0.60488960 | 0.56735060 | 0.53150910 |
| 632 | cg00615377 | 0.11265030 | 0.16140570 | 0.19404450 | 0.17468600 |
| 743 | cg00717862 | 0.07999436 | 0.07873347 | 0.06089359 | 0.06171374 |
| 887 | cg00862290 | 0.43640520 | 0.54047160 | 0.60786800 | 0.56325950 |
| 911 | cg00888479 | 0.07388961 | 0.07361080 | 0.10149800 | 0.09985076 |
| 979 | cg00945507 | 0.13432250 | 0.23854600 | 0.34749760 | 0.28903340 |
| | exmdata5[, 2] | exmdata6[, 2] | exmdata7[, 2] | exmdata8[, 2] | |
| 146 | 0.67191510 | 0.63137380 | 0.47929610 | 0.45428300 | |
| 189 | 0.34106080 | 0.33765930 | 0.41252110 | 0.37024890 | |
| 280 | 0.61920530 | 0.61925200 | 0.46753250 | 0.55632410 | |
| 632 | 0.12573100 | 0.14483660 | 0.16338240 | 0.20130510 | |
| 743 | 0.07594936 | 0.09062161 | 0.06475791 | 0.07271878 | |
| 887 | 0.50259740 | 0.40111730 | 0.56646700 | 0.54552980 | |
| 911 | 0.08633986 | 0.06765189 | 0.09070268 | 0.12417730 | |
| 979 | 0.11848510 | 0.16653850 | 0.30718420 | 0.26624740 | |
| diff_results\$ordfit_t[diff_list_boot] | | | | | |
| 146 | | | 5.636263 | | |
| 189 | | | -3.232921 | | |
| 280 | | | 4.337628 | | |
| 632 | | | -3.722206 | | |
| 743 | | | 2.918806 | | |
| 887 | | | -3.368752 | | |
| 911 | | | -3.490240 | | |
| 979 | | | -4.968792 | | |
| diff_results\$bootstrap_p[diff_list_boot] | | | | | |
| 146 | | | 0 | | |
| 189 | | | 0 | | |
| 280 | | | 0 | | |
| 632 | | | 0 | | |
| 743 | | | 0 | | |
| 887 | | | 0 | | |
| 911 | | | 0 | | |
| 979 | | | 0 | | |