

# Package ‘TOAST’

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**Type** Package

**Title** Tools for the analysis of heterogeneous tissues

**Version** 1.18.0

**Description** This package is devoted to analyzing high-throughput data (e.g. gene expression microarray, DNA methylation microarray, RNA-seq) from complex tissues. Current functionalities include 1. detect cell-type specific or cross-cell type differential signals 2. tree-based differential analysis 3. improve variable selection in reference-free deconvolution 4. partial reference-free deconvolution with prior knowledge.

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**License** GPL-2

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**biocViews** DNAMethylation, GeneExpression, DifferentialExpression, DifferentialMethylation, Microarray, GeneTarget, Epigenetics, MethylationArray

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assignCellType	<i>Align cell types when reference proportions are known</i>
----------------	--

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### Description

Align target proportions with reference proportions by pearson correlation coefficients.

### Usage

```
assignCellType(input,reference)
```

### Arguments

input	Input proportiona matrix of dimension N by K.
reference	Reference proportion matrix of dimension N by K.

### Value

The aligned proportion matrix, following the cell type ordering of reference proportion matrix.

### Author(s)

Ziyi Li <zli16@mdanderson.org>

## References

Ziyi Li, Zhijin Wu, Peng Jin, Hao Wu. "Dissecting differential signals in high-throughput data from complex tissues."

## Examples

```
## generate estimated proportion matrix
estProp <- matrix(abs(runif(50*4,0,1)), 50, 4)
estProp <- sweep(estProp, 1, rowSums(estProp), "/")

## generate reference proportion matrix
refProp <- matrix(abs(runif(50*4,0,1)), 50, 4)
refProp <- sweep(refProp, 1, rowSums(refProp), "/")

estProp_aligned = assignCellType(input = estProp,
reference = refProp)
```

---

beta\_emp

*Simulated methylation 450K array data with related*

---

## Description

This dataset is a list containing two matrices, one of which is methylation 450K array data of 3000 CpG sites on 50 samples, the other is methylation 450K array data of 3000 matched CpG sites on three immune cell types. The first dataset is generated by simulation. It originally has 459226 features and 50 samples. We reduce it to 3000 CpGs by random selection.

## Usage

```
data("beta_emp")
```

## Format

The format is: List of 2 \$ Y.raw: num [1:3000, 1:50] 0.7661 0.0968 0.8882 0.0286 0.6956 ... ..- attr(\*, "dimnames")=List of 2 .. ..\$: chr [1:3000] "cg08752431" "cg14555682" "cg23086843" "cg20308511" ... ..\$: NULL \$ ref.m: num [1:3000, 1:3] 0.7712 0.0996 0.9065 0.037 0.7242 ... ..- attr(\*, "dimnames")=List of 2 .. ..\$: chr [1:3000] "cg08752431" "cg14555682" "cg23086843" "cg20308511" ... ..\$: chr [1:3] "CD4T" "CD8T" "BCell"

## Examples

```
data(beta_emp)
```

---

CBS_PBMC_array	<i>An example dataset for partial reference-free cell composition estimation from tissue gene expression</i>
----------------	--

---

### Description

The dataset contains 511 microarray gene expressions for 20 PBMC samples (mixed\_all) and PBMC microarray reference for the matched 511 genes from 5 immune cell types (LM\_5). It also contains the true cell compositions from cell sorting experiment (trueProp) and prior knowledge of cell compositions for 5 cell types in PBMC (prior\_alpha and prior\_sigma).

### Usage

```
data("CBS_PBMC_array")
```

### References

Newman, Aaron M., et al. "Robust enumeration of cell subsets from tissue expression profiles." *Nature methods* 12.5 (2015): 453.

Rahmani, Elior, et al. "BayesCCE: a Bayesian framework for estimating cell-type composition from DNA methylation without the need for methylation reference." *Genome biology* 19.1 (2018): 141.

### Examples

```
data("CBS_PBMC_array")
CBS_PBMC_array$mixed_all[1:5,1:5]
head(CBS_PBMC_array$LM_5,3)
head(CBS_PBMC_array$trueProp,3)
CBS_PBMC_array$prior_alpha
CBS_PBMC_array$prior_sigma
```

---

cedar	<i>Testing cell type specific differential signals for specified phenotype by considering DE/DM state correlation between cell types.</i>
-------	---

---

### Description

This function provides posterior probability of whether a feature is DE/DM in certain cell type given observed bulk data.

**Usage**

```
cedar(Y_raw, prop, design.1, design.2=NULL, factor.to.test=NULL,
      pval = NULL, p.adj = NULL, tree = NULL, p.matrix.input = NULL,
      de.state = NULL, cutoff.tree = c('fdr', 0.01),
      cutoff.prior.prob = c('pval', 0.01),
      similarity.function = NULL, parallel.core = NULL, corr.fig = FALSE,
      run.time = TRUE, tree.type = c('single','full'))
```

**Arguments**

Y_raw	matrix of observed bulk data, with rows representing features and columns representing samples
prop	matrix of cell type composition of samples, with rows representing samples and columns representing cell types
design.1	covariates with cell type specific effect, with rows representing samples and columns representing covariates
design.2	covariates without cell type specific effect, with rows representing samples and columns representing covariates
factor.to.test	A phenotype name, e.g. "disease", or a vector of contrast terms, e.g. c("disease", "case", "control").
pval	matrix of p-values, with rows representing features and columns representing cell types. colnames must be same as input of prop
p.adj	matrix of adjusted p-values, with rows representing features and columns representing cell types. colnames must be same as input of prop
tree	tree structure between cell types, a matrix with row representing layers and column representing cell types (column name is required)
p.matrix.input	prior probability on each node of the tree structure. only work when tree structure has been specified. the dimension must be same as tree input.
de.state	DE/DM state of each feature in each cell type, with row representing features and column representing cell types (1:DE/DM, 0:non-DE/DM)
cutoff.tree	cut off used to define DE state to estimate tree could be 'fdr' or 'pval' default it 'fdr'=0.01. suggest to start with restrictive cut off and change to relative loose value when the restrictive cut off is failed
cutoff.prior.prob	cut off used to define DE state to estimate prior probs of nodes on tree could be 'fdr' or 'pval' default it 'fdr'=0.01. suggest to start with restrictive cut off and change to relative loose value when the restrictive cut off is failed
similarity.function	custom function used to calculate similarity between cell types that used for tree structure estimation. the input of the custom is assumed to be a matrix of log transformed p-value. dimension is: selected gene number * cell number
parallel.core	number of cores for parallel running, default is NULL
corr.fig	a boolean value, whether to plot correlation between cell types use function plotCorr()
run.time	a boolean value, whether to report running time in seconds
tree.type	tree type for inference, default is c('single','full')

**Value**

A list

<code>toast_res</code>	If <code>pval</code> is NULL, then TOAST result by function <code>csTest()</code> is returned
<code>tree_res</code>	matrix of posterior probability of each feature for each cell type
<code>fig</code>	If <code>corr.fig = TRUE</code> , then figure show DE/DM state correlation between cell types will be returned
<code>time_used</code>	If <code>run.time = TRUE</code> , then running time (seconds) of CeDAR will be returned

**Author(s)**

Luxiao Chen <luxiao.chen@emory.edu>

**Examples**

```

N <- 300 # simulation a dataset with 300 samples
K <- 3 # 3 cell types
P <- 500 # 500 features

### simulate proportion matrix
Prop <- matrix(runif(N*K, 10,60), ncol=K)
Prop <- sweep(Prop, 1, rowSums(Prop), FUN="/")
colnames(Prop) <- c("Neuron", "Astrocyte", "Microglia")

### simulate phenotype names
design <- data.frame(disease=factor(sample(0:1,size = N,replace=TRUE)),
                    age=round(runif(N, 30,50)),
                    race=factor(sample(1:3, size = N,replace=TRUE)))
Y <- matrix(rnorm(N*P, N, P), ncol = N)
rownames(Y) <- paste0('gene',1:nrow(Y))
d1 <- data.frame('disease' = factor(sample(0:1,size = N,replace=TRUE)))

### Only provide bulk data, proportion
res <- cedar(Y_raw = Y, prop = Prop,
            design.1 = design[,1:2],
            design.2 = design[,3],
            factor.to.test = 'disease',
            cutoff.tree = c('pval',0.1),
            corr.fig = TRUE,
            cutoff.prior.prob = c('pval',0.1) )

### result of toast (independent test)
str(res$toast_res)
### posterior probability of DE/DM of cedar with single layer tree structure
head(res$tree_res$single$pp)
### posterior probability of DE/DM of cedar with multiple layer tree structure
head(res$tree_res$full$pp)
### estimated tree structure of three cell types
head(res$tree_res$full$tree_structure)
### scatter plot of -log10(pval) showing DE/DM state correlation between cell types
res$fig

```

```

### Using custom similarity function to estimate tree structure
### In CeDAR, the input is assumed to be a matrix of log transformed p-values
### with row representing genes and columns representing cell types

sim.fun <- function(log.pval){
  similarity.res <- sqrt((1 - cor(log.pval, method = 'spearman'))/2)
  return(similarity.res)
}

res <- cedar(Y_raw = Y, prop = Prop,
            design.1 = design[,1:2],
            design.2 = design[,3],
            factor.to.test = 'disease',
            cutoff.tree = c('pval',0.1),
            similarity.function = sim.fun,
            corr.fig = FALSE,
            cutoff.prior.prob = c('pval',0.1) )

### posterior probability of DE/DM of cedar with multiple layer tree structure
head(res$tree_res$full$pp)

### Using custom tree structure as input
### cell type 1 and cell type 3 are more similar
tree.input <- rbind(c(1,1,1),c(1,2,1),c(1,2,3))
### If column name is provided for the matrix; make sure it is same as variable Prop
colnames(tree.input) <- c("Neuron", "Astrocyte", "Microglia")

res <- cedar(Y_raw = Y, prop = Prop,
            design.1 = design[,1:2],
            design.2 = design[,3],
            factor.to.test = 'disease',
            cutoff.tree = c('pval',0.1),
            tree = tree.input,
            corr.fig = FALSE,
            cutoff.prior.prob = c('pval',0.1) )

### posterior probability of DE/DM of cedar with multiple layer tree structure
head(res$tree_res$custom$pp)

### Using custom tree structure and prior probability of each node as input
### cell type 1 and cell type 3 are more similar
tree.input <- rbind(c(1,1,1),c(1,2,1),c(1,2,3))
colnames(tree.input) <- c("Neuron", "Astrocyte", "Microglia")

p.matrix.input <- rbind(c(0.2,0.2,0.2), c(0.5,0.25,0.5), c(0.5,1,0.5))
# marginally, each cell type has 0.05 (cell 1: 0.2 * 0.5 * 0.5, cell 2: 0.2 * 0.25 * 1)
# probability to be DE for a randomly picked gene
# there will be about 50% DE genes in cell type 1 overlapped with cell type 3;

```

```
# while there will be about 25% DE genes in cell type 1 overlaped with cell type 2

res <- cedar(Y_raw = Y, prop = Prop,
            design.1 = design[,1:2],
            design.2 = design[,3],
            factor.to.test = 'disease',
            cutoff.tree = c('pval',0.1),
            tree = tree.input,
            p.matrix.input = p.matrix.input,
            corr.fig = FALSE,
            cutoff.prior.prob = c('pval',0.1) )

### posterior probability of DE/DM of cedar with muliple layer tree structure
head(res$tree_res$custom$pp)
```

---

ChooseMarker	<i>Choose cell type-specific markers from pure cell type profiles or single cell data</i>
--------------	---

---

## Description

Choose cell type-specific markers from pure cell type profiles generated by microarray or RNA-seq, or from single cell RNA-seq data by differential analysis.

## Usage

```
ChooseMarker(pure_all, CellType, nMarkCT = 10, chooseSig = FALSE, verbose = TRUE)
```

## Arguments

pure_all	Input pure cell type profile matrix or single cell data matrix. Rows are for genes, columns are for cell types or cells.
CellType	A list object consisting of cell type information for columns in pure_all. Each element is a cell type, and contains the corresponding column number in pure_all matrix. For example, CellType = list(BCell = 1:3, CD4T = 4:5).
nMarkCT	Number of markers chosen per cell type. Default is 10.
chooseSig	A boolean variable representing whether to consider the significance of selected markers. When chooseSig = FALSE, all nMarkerCT number of markers will be chosen per cell type. Otherwise the non-significant (p value > 0.05) markers will be filtered out.
verbose	A boolean variable of whether to output messages.

## Details

Here we provide more details for CellType variable. This variable should be a list, with each element being the corresponding column numbers in pure\_all for each cell type. For example, suppose pure\_all is a 1000 by 300 matrix with row being genes and column being cells (or cell types). The first 1 to 100 columns are cell A, 101 to 200 columns are cell B, and 201 to 300 columns are cell C. Then CellType should be assigned as CellType = list(A = 1:100, B = 101:200, C = 201:300). If pure\_all only has three columns for three cell types A, B and C, then CellType = list(A = 1, B = 2, C = 3).

## Value

A list variable, including the selected variables for all cell types.

## Author(s)

Ziyi Li <zli16@mdanderson.org>

## References

Ziyi Li, Zhenxing Guo, Ying Cheng, Peng Jin, Hao Wu. "Robust partial reference-free cell composition estimation from tissue expression profiles."

## Examples

```
## randomly simulate pure cell type profiles
pure_all <- matrix(abs(rnorm(1000*9)), 1000, 9)
CellType <- list(CellA = 1:3,
                 CellB = 4:6,
                 CellC = 7:9)

## choose significant markers
SelMarker <- ChooseMarker(pure_all, CellType,
                           nMarkCT = 30,
                           chooseSig = TRUE,
                           verbose = FALSE)
```

---

csDeconv

*Improve reference-free deconvolution using cross-cell type differential analysis*

---

## Description

This function improve the feature selection in reference-free deconvolution through cross-cell type differential analysis

## Usage

```
csDeconv(Y_raw, K, FUN, nMarker = 1000,
         InitMarker = NULL, TotalIter = 30, bound_negative = FALSE)
```

**Arguments**

Y_raw	A $G \times N$ matrix, $G$ is the number of features, $N$ is the number of subjects; or a SummarizedExperiment object.
K	The number of cell types. Need to be specified a priori.
FUN	The reference-free deconvolution function, this function should take Y_raw and K, and the return values should be a $N$ by $K$ proportion matrix. $N$ is the number of samples and $K$ is the number of cell types. Default function is a wrapper of the RefFreeCellMix() function from CRAN package RefFreeEWAS.
nMarker	The number of markers used in the deconvolution. Default is 1000.
InitMarker	A vector of length $L$ to represent the selection of initial markers. $L$ should be equal or smaller than $G$ . If $G$ is large, it is recommended that $L$ is much smaller than $G$ . If not specified, the most variable nMarker features will be used.
TotalIter	The total number of iterations of applying cross-cell type differential analysis. Default is 30.
bound_negative	Whether to bound all negative parameter estimators to zero.

**Value**

allProp	A list of estimated proportions from all iterations.
allRMSE	A vector of root mean squared errors (RMSE) from all iterations.
estProp	A $N \times K$ matrix representing the mixture proportions of $K$ cell types in $N$ subjects, chosen from allProp with the smallest RMSE.
updatedInx	Selected variable index from the algorithm.

**Author(s)**

Ziyi Li <zli16@mdanderson.org>

**References**

Ziyi Li and Hao Wu. "Improving reference-free cell composition estimation by cross-cell type differential analysis".

**Examples**

```
Y_raw <- abs(matrix(runif(10000*20, 0,1),10000,20))
K <- 3

## wrap your reference-free
## deconvolution method into a function
## this function should take Y and K as input
## and output a N by K proportion matrix
## here we use RefFreeCellMix() as an example
outT <- csDeconv(Y_raw, K)

RefFreeCellMix_wrapper <- function(Y, K){
  outY = myRefFreeCellMix(Y,
```

```

    mu0=myRefFreeCellMixInitialize(Y,
    K = K)
    Prop0 = outY$Omega
    return(Prop0)
}

outT <- csDeconv(Y_raw, K,
                FUN = RefFreeCellMix_wrapper)

```

---

csTest                      *Testing differential signals for specified phenotype and cell type(s).*

---

## Description

This function conducts statistical tests for specified phenotype and cell type(s).

## Usage

```

csTest(fitted_model, coef = NULL, cell_type = NULL,
       contrast_matrix = NULL, var_shrinkage = TRUE,
       verbose = TRUE, sort = TRUE)

```

## Arguments

fitted_model	The output from fitModel() function.
coef	A phenotype name, e.g. "disease", or a vector of contrast terms, e.g. c("disease", "case", "control").
cell_type	A cell type name, e.g. "celltype1", or "neuron". If cell_type is NULL or specified as "ALL", compound effect of coef in all cell types will be tested.
contrast_matrix	If contrast_matrix is specified, coef and cell_type will be ignored! A matrix (or a vector) to specify contrast, e.g., cmat <- matrix(0, 2, 6); cmat[1,3] <- 1; cmat[2,4] <- 1 is to test whether the 3rd parameter and 4th parameter are zero simultaneously i.e. beta3 = beta4 = 0.
var_shrinkage	Whether to apply shrinkage on estimated MSE or not. Applying shrinkage helps remove extremely small variance estimation and stabilize statistics.
verbose	A boolean parameter. Testing information will be printed if verbose = TRUE.
sort	A boolean parameter. The output results will be sorted by p value if sort = TRUE.

## Value

A matrix including the results from testing the phenotype in specified cell type(s).

## Author(s)

Ziyi Li <zli16@mdanderson.org>

## References

Ziyi Li, Zhijin Wu, Peng Jin, Hao Wu. "Dissecting differential signals in high-throughput data from complex tissues."

## Examples

```

N <- 300 # simulation a dataset with 300 samples
K <- 3 # 3 cell types
P <- 500 # 500 features

### simulate proportion matrix
Prop <- matrix(runif(N*K, 10,60), ncol=K)
Prop <- sweep(Prop, 1, rowSums(Prop), FUN="/")
colnames(Prop) <- c("Neuron", "Astrocyte", "Microglia")

### simulate phenotype names
design <- data.frame(disease=factor(sample(0:1,
                                     size = N,replace=TRUE)),
                   age=round(runif(N, 30,50)),
                   race=factor(sample(1:3, size = N,replace=TRUE)))
Y <- matrix(rnorm(N*P, N, P), ncol = N)

### generate design matrix and fit model
Design_out <- makeDesign(design, Prop)
fitted_model <- fitModel(Design_out, Y)

### check the names of cell types and phenotypes
fitted_model$all_cell_types
fitted_model$all_coefs

### detect age effect in neuron
test <- csTest(fitted_model, coef = "age",
              cell_type = "Neuron", contrast_matrix = NULL)

## coef can be specified in different ways:
#### jointly test a phenotype:
test <- csTest(fitted_model, coef = "age",
              cell_type = "joint", contrast_matrix = NULL)

#### if I do not specify cell_type
test <- csTest(fitted_model, coef = "age",
              cell_type = NULL, contrast_matrix = NULL)
## this is exactly the same as
test <- csTest(fitted_model, coef = "age",
              contrast_matrix = NULL)

#### other examples
test <- csTest(fitted_model, coef = "race",
              cell_type = "Astrocyte", contrast_matrix = NULL)
test <- csTest(fitted_model, coef = "age",
              cell_type = "Microglia", contrast_matrix = NULL)

```

```
#### specify contrast levels
test <- csTest(fitted_model, coef = c("race", 3, 2),
cell_type = "Neuron", contrast_matrix = NULL)
#### specify contrast levels in all cell types
test <- csTest(fitted_model, coef = c("race", 3, 2),
cell_type = "joint", contrast_matrix = NULL)

#### csTest can tolerate different ways of specifying contrast level
#### note race=1 is used as reference when fitting model
#### we can here specify race=2 as reference
test <- csTest(fitted_model, coef = c("race", 1, 2),
cell_type = "Neuron", contrast_matrix = NULL)
## get exactly the same results as
test <- csTest(fitted_model, coef = c("race", 2, 1),
cell_type = "Neuron", contrast_matrix = NULL)

#### specify a contrast matrix:
cmatrix = rep(0,15)
cmatrix[c(4,5)] = c(1,-1)
test <- csTest(fitted_model, coef = NULL,
cell_type = NULL, contrast_matrix = cmatrix)
#### specific a contrast matrix with two rows:
cmatrix = matrix(rep(0,30),2,15)
cmatrix[1,4] = 1
cmatrix[2,5] = 1
test <- csTest(fitted_model, coef = NULL,
contrast_matrix = cmatrix)
```

---

DEVarSelect

*Feature selection for reference-free deconvolution using cross-cell type differential analysis*


---

## Description

This function selects cross-cell type differential features for reference-free deconvolution.

## Usage

```
DEVarSelect(Y_raw, Prop0, nMarker, bound_negative)
```

## Arguments

Y_raw	A data matrix containing P features and N samples; or a SummarizedExperiment object.
Prop0	A N by K proportion matrix with K as number of cell types.
nMarker	Number of markers selected.
bound_negative	Whether to bound all negative parameter estimators to zero.

**Value**

Selected markers using cross-cell type differential analysis.

**Author(s)**

Ziyi Li <zli16@mdanderson.org>

**References**

Ziyi Li, Zhijin Wu, Peng Jin, Hao Wu. "Dissecting differential signals in high-throughput data from complex tissues."

**Examples**

```
Y_raw <- matrix(runif(5000*20, 0, 1), 5000, 20)
tmp <- matrix(runif(20*4), 20, 4)
Prop0 <- sweep(tmp, 1, rowSums(tmp), "/")
varlist <- DEVarSelect(Y_raw, Prop0,
                      nMarker=1000,
                      bound_negative=FALSE)
```

---

findRefinx

*findRefinx*

---

**Description**

Find index for marker genes with largest coefficient of variation based on raw data.

**Usage**

```
findRefinx(rawdata, nmarker=1000, sortBy = "var")
```

**Arguments**

rawdata	A data matrix with rows representing features and columns representing samples; or a SummarizedExperiment object.
nmarker	Desired number of markers after selection. Default is 1000.
sortBy	Desired method to select features. "var" represents selecting by largest variance. "cv" represents selecting by largest coefficients of variation. Default is "var".

**Value**

A vector of index for the selected markers.

**Author(s)**

Ziyi Li <zli16@mdanderson.org>

**References**

Ziyi Li, Zhijin Wu, Peng Jin, Hao Wu. "Dissecting differential signals in high-throughput data from complex tissues."

**Examples**

```
Y_raw <- matrix(runif(5000*20, 0, 1), 5000, 20)
idx <- findRefinx(Y_raw, nmarker=500)
idx2 <- findRefinx(Y_raw, nmarker=500, sortBy = "cv")
```

---

fitModel

*Fit model with proportions and phenotypes.*


---

**Description**

This function receives design matrix from makeDesign() and fits the model including all cell types and phenotypes.

**Usage**

```
fitModel(Design_out, Y)
```

**Arguments**

Design_out	The output from function makeDesign().
Y	A G*N matrix, G is the number of features, N is the number of subjects; or a SummarizedExperiment object.

**Value**

Design_out	The input Design_out object.
N	Number of samples from matrix Y.
coefs	Estimated coefficients (beta) in the model.
coefs_var	Estimated variance of the coefficients (beta variance) in the model.
Y	Observation Y matrix.
Ypred	Predicted Y from the fitted model.
all_coefs	The names of all phenotypes.
all_cell_types	The names of all cell types.
MSE	Estimated mean squared error.
model_names	The names of all terms in the fitted model.

**Author(s)**

Ziyi Li <zli16@mdanderson.org>

## References

Ziyi Li, Zhijin Wu, Peng Jin, Hao Wu. "Dissecting differential signals in high-throughput data from complex tissues."

## Examples

```
N = 300 # simulation a dataset with 300 samples
K = 3 # 3 cell types
P <- 500 # 500 features

### simulate proportion matrix
Prop = matrix(runif(N*K, 10,60), ncol=K)
Prop = sweep(Prop, 1, rowSums(Prop), FUN="/")
colnames(Prop) = c("Neuron", "Astrocyte", "Microglia")
Y <- matrix(rnorm(N*P, N, P), ncol = N)

### simulate phenotype names
design <- data.frame(disease=factor(sample(0:1,
                                     size = N,replace=TRUE)),
                   age=round(runif(N, 30,50)),
                   race=factor(sample(1:3, size = N,replace=TRUE)))
Design_out <- makeDesign(design, Prop)

### fit model
fitted_model <- fitModel(Design_out, Y)
```

---

GetPrior

*Get prior knowledge for supported tissue types*

---

## Description

Users can use this function to get priors provided in this package. Users can also directly specify tissue type in MDeconv function.

## Usage

```
GetPrior(alpha = NULL, sigma = NULL)
```

## Arguments

alpha	A string chosen from "human pbmc", "human liver", "human brain", "human pancreas", "human skin", or a numeric vector for manually specified alpha.
sigma	Keep it as NULL for supported tissues. Otherwise a numeric vector for manually specified sigma.

**Value**

alpha_prior	Prior knowledge for the mean of proportions.
sigma_prior	Prior knowledge for the sigma of proportions.

**Author(s)**

Ziyi Li <zli16@mdanderson.org>

**References**

Ziyi Li, Zhenxing Guo, Ying Cheng, Peng Jin, Hao Wu. "Robust partial reference-free cell composition estimation from tissue expression profiles."

**Examples**

```
GetPrior("human pbmc")
GetPrior("human liver")
GetPrior("human brain")
GetPrior("human skin")
GetPrior("human pancreas")
```

---

makeDesign

*Generate design matrix from input phenotypes and proportions.*


---

**Description**

This function generate design matrix and make preparations for following fitModel and csTest.

**Usage**

```
makeDesign(design, Prop)
```

**Arguments**

design	A N by P phenotype matrix, with rows as samples and columns as phenotypes (e.g. age, gender, disease, etc.).
Prop	A N by K proportion matrix, with rows as samples and columns as cell types

**Value**

design_matrix	A comprehensive design matrix incorporated phenotype and proportion information.
Prop	The input proportion matrix.
design	The input design/phenotype matrix.
all_coefs	The names of all phenotypes.
all_cell_types	The names of all cell types.
formula	The formula of the tested model, including all phenotypes, cell types and interaction terms.

**Author(s)**

Ziyi Li <zli16@mdanderson.org>

**References**

Ziyi Li, Zhijin Wu, Peng Jin, Hao Wu. "Dissecting differential signals in high-throughput data from complex tissues."

**Examples**

```
N = 300 # simulation a dataset with 300 samples
K = 3 # 3 cell types

### simulate proportion matrix
Prop = matrix(runif(N*K, 10,60), ncol=K)
Prop = sweep(Prop, 1, rowSums(Prop), FUN="/")
colnames(Prop) = c("Neuron", "Astrocyte", "Microglia")

### simulate phenotype names
design <- data.frame(disease=factor(sample(0:1, size = N,replace=TRUE)),
                    age=round(runif(N, 30,50)),
                    race=factor(sample(1:3, size = N,replace=TRUE)))
Design_out <- makeDesign(design, Prop)
```

---

MDeconv

---

*Estimate cell compositions via partial reference-free deconvolution.*


---

**Description**

This function is the wrapper for TOAST/-P (partial reference-free deconvolution without prior) and TOAST/+P (with prior). It guides cell composition estimation through extra biological information, including cell type specific markers and prior knowledge of compositions.

**Usage**

```
MDeconv(Ymat, SelMarker,
        alpha = NULL, sigma = NULL,
        epsilon = 0.001, maxIter = 1000,
        verbose = TRUE)
```

**Arguments**

Ymat	A gene expression data matrix from complex tissues. Rows represent genes and columns represent samples. Row names (e.g. gene symbol or ID) are needed. There is no need to filter data matrix by marker genes.
SelMarker	A list variable with each element includes cell type-specific markers for this cell type. The marker list can be selected from pure cell type profiles or single cell data using ChooseMarker(). It can also be easily created manually. Please see details section below and example for more information.

alpha	A vector including the prior mean for all cell types.
sigma	A vector including the prior standard deviation for all cell types.
epsilon	A numeric variable to control the level of convergence. With a large epsilon, it converges quickly and the algorithm may not converge well. With a small epsilon, it converges slower and the algorithm may converge "too much". The default value is 1e-3, which we find is a reasonable threshold.
maxIter	Number of maximum iterations.
verbose	A boolean variable of whether to output messages.

### Details

More about SelMarker: in addition to selecting markers using ChooseMarker(), we can manually specific marker list. For example, if we know there are two cell type-specific markers for cell type A "Gene1" and "Gene2", and two cell type-specific markers for cell type B "Gene3" and "Gene4", we can create marker list by SelMarker = list(CellA = c("Gene1", "Gene2"), CellB = c("Gene3", "Gene4")).

One thing to note is that, the genes in marker list should have matches in row names of Ymat. If not, the unmatched markers will be removed during analysis.

### Value

A list including

H	Estimated proportion matrix, rows for cell types and columns for samples.
W	Estimated pure cell type profile matrix, rows for genes and columns for cell types

### Author(s)

Ziyi Li <zli16@mdanderson.org>

### References

Ziyi Li, Zhenxing Guo, Ying Cheng, Peng Jin, Hao Wu. "Robust partial reference-free cell composition estimation from tissue expression profiles."

### Examples

```
# simulate mixed data from complex tissue
# without prior
Wmat <- matrix(abs(rnorm(60*3, 4, 4)), 60, 3)
SelMarker <- list(CellA = 1:20,
                  CellB = 21:40,
                  CellC = 41:60)
for(i in 1:3) {
  Wmat[SelMarker[[i]], i] <- abs(rnorm(20, 50, 10))
}
Hmat <- matrix(runif(3*25), 3, 25)
Hmat <- sweep(Hmat, 2, colSums(Hmat), "/")
```

```

Ymat <- Wmat %*% Hmat + abs(rnorm(60*25))
rownames(Ymat) <- 1:60

# deconvolution with TOAST/-P (TOAST without prior)
res <- MDeconv(Ymat, SelMarker, verbose = FALSE)
print(dim(Ymat))
cor(t(res$H), t(Hmat))

# suppose we observe the proportions
# for the same tissue from another study
alpha_prior <- rep(0.33, 3)
sigma_prior <- rep(1, 3)

# deconvolution with TOAST/+P (TOAST with prior)
res2 <- MDeconv(Ymat, SelMarker,
                alpha = alpha_prior, sigma = sigma_prior,
                verbose = FALSE)
cor(t(res2$H), t(Hmat))

```

---

myprojectMix

*Replicate the function myprojectMix() from RefFreeEWAS package*


---

## Description

Replicate the function myprojectMix() from RefFreeEWAS package (<https://cran.r-project.org/web/packages/RefFreeEWAS/>) as that package is not in CRAN anymore

## Usage

```
myprojectMix(Y, Xmat, nonnegative=TRUE, sumLessThanOne=TRUE, lessThanOne=!sumLessThanOne)
```

## Arguments

Y	Matrix (m CpGs x n Subjects) of DNA methylation beta values
Xmat	Matrix (m CpGs x K cell types) of cell-type specific methylomes
nonnegative	All coefficients $\geq 0$ ?
sumLessThanOne	Coefficient rows should sum to less than one?
lessThanOne	Every value should be less than one (but possibly sum to value greater than one)?

## Details

Function for projecting methylation values (Y) onto space of methylomes (Xmat), with various constraints. This is the reference-based method described in Houseman et al. (2012) and also appearing in the minfi package.

## Value

Projection coefficients resulting from constrained projection

---

myRefFreeCellMix      *Replicate the function RefFreeCellMix() from RefFreeEWAS package*

---

### Description

Replicate the function RefFreeCellMix() from RefFreeEWAS package (<https://cran.r-project.org/web/packages/RefFreeEWAS/>) as that package is not in CRAN anymore

### Usage

```
myRefFreeCellMix(Y, mu0=NULL, K=NULL, iters=10, Yfinal=NULL, verbose=TRUE)
```

### Arguments

Y	Matrix (m CpGs x n Subjects) of DNA methylation beta values
mu0	Matrix (m CpGs x K cell types) of *initial* cell-type specific methylomes
K	Number of cell types (ignored if mu0 is provided)
iters	Number of iterations to execute
Yfinal	Matrix (m* CpGs x n Subjects) of DNA methylation beta values on which to base final methylomes
verbose	Report summary of errors after each iteration?

### Details

Reference-free decomposition of DNA methylation matrix into cell-type distributions and cell-type methylomes,  $Y = Mu \Omega^T$ . Either an initial estimate of Mu must be provided, or else the number of cell types K, in which case RefFreeCellMixInitialize will be used to initialize. Note that the decomposition will be based on Y, but Yfinal (=Y by default) will be used to determine the final value of Mu based on the last iterated value of Omega.

### Value

Object of S3 class RefFreeCellMix, containing the last iteration of Mu and Omega.

### Author(s)

E. Andres Houseman

### References

Houseman, E. Andres, Kile, Molly L., Christiani, David C., et al. Reference-free deconvolution of DNA methylation data and mediation by cell composition effects. BMC bioinformatics, 2016, vol. 17, no 1, p. 259.

---

myRefFreeCellMixInitialize

*Replicate the function RefFreeCellMixInitialize() from RefFreeEWAS package*

---

### Description

Replicate the function RefFreeCellMixInitialize() from RefFreeEWAS package (<https://cran.r-project.org/web/packages/RefFreeEWAS/>) as that package is not in CRAN anymore

### Usage

```
myRefFreeCellMixInitialize(Y,K=2,Y.Distance=NULL, Y.Cluster=NULL,
  largeOK=FALSE, dist.method = "euclidean", ...)
```

### Arguments

Y	Matrix (m CpGs x n Subjects) of DNA methylation beta values
K	Number of cell types
Y.Distance	Distance matrix (object of class "dist") to use for clustering.
Y.Cluster	Hierarchical clustering object (from hclust function)
largeOK	OK to calculate distance matrix for large number of subjects? (See details.)
dist.method	Method for calculating distance matrix
...	Additional parameters for hclust function

### Details

Initializes the methylome matrix "Mu" for RefFreeCellMix by computing the mean methylation (from Y) over K clusters of Y, determined by the Y.Cluster object. If Y.Cluster object does not exist, it will be created from Y.Distance (using additional clustering parameters if supplied). If Y.Distance does not exist, it will be created from t(Y). As a protection against attempting to fit a very large distance matrix, the program will stop if the number of columns of Y is > 2500, unless largeOK is explicitly set to TRUE.

### Value

An m x K matrix of mean methylation values.

### Author(s)

E. Andres Houseman

---

`plotCorr`*Show DE/DM state correlation between cell types*

---

**Description**

This function generates  $-\log_{10}$  transformed p-values for each pair of cell types and calculate corresponding Pearson correlation and odds ratio - a modification of function 'ggpairs' in package 'GGally'.

**Usage**

```
plotCorr(pval, de.state=NULL, pval.thres=NULL, fdr.thres=NULL, p.size = 0.2,
         p.color = grDevices::adjustcolor("black", alpha.f = 0.2),
         fig.margin = c(1,1,1,1),
         fig.margin.unit = 'in', line.type = 'dashed', line.color = 'blue')
```

**Arguments**

<code>pval</code>	matrix of p-values, with rows representing features and columns representing cell types.
<code>de.state</code>	(optional) matrix of DE/DM states (1: DE/DM, 0:non-DE/DM), with rows representing features and columns representing cell types.
<code>pval.thres</code>	threshold of p-value to define DE/DM, required if <code>de.state</code> not provided.
<code>fdr.thres</code>	threshold of FDR to define DE/DM, required if <code>de.state</code> not provided and FDR is preferred than p-value.
<code>p.size</code>	point size for scatter plot
<code>p.color</code>	point color for scatter plot
<code>fig.margin</code>	figure margin
<code>fig.margin.unit</code>	unit of figure margin
<code>line.type</code>	line type in scatter plot
<code>line.color</code>	line color in scatter plot

**Value**

A figure contains scatter plots, Pearson correlation and odds ratio of  $-\log_{10}$  transformed p-values for each pair of cell types.

**Author(s)**

Luxiao Chen <luxiao.chen@emory.edu>

**Examples**

```
pval.1 <- runif(1000,0,1)
pval.2 <- pval.1 + rnorm(1000,0,0.01)
pval.2[pval.2 < 0] =0
pval.3 <- runif(1000,0,1)

pval.input <- data.frame('cell.1'=pval.1,
                        'cell.2'=pval.2,
                        'cell.3'=pval.3)

plotCorr(pval = pval.input, pval.thres = 0.05)
```

---

RA\_100samples

*An example dataset for cellular proportion estimation and multiple factor design*

---

**Description**

The dataset contains normalized beta values for 3000 CpGs from 100 samples (50 Rheumatoid arthritis patients and 50 controls) and their phenotypes (disease status, age, and gender). The dataset also contains a sub-setted blood reference matrix for the matched 3000 CpGs. This data was obtained and processed based on GSE42861.

**Usage**

```
data("RA_100samples")
```

**References**

Liu Y, Aryee MJ, Padyukov L, Fallin MD et al. Epigenome-wide association data implicate DNA methylation as an intermediary of genetic risk in rheumatoid arthritis. *Nat Biotechnol* 2013 Feb;31(2):142-7. PMID: 23334450

**Examples**

```
data(RA_100samples)
RA_100samples$Y_raw[1:5,1:5]
head(RA_100samples$Pheno)
head(RA_100samples$Blood_ref)
```

---

Tsisal *Complete Deconvolution of DNA methylation data based on TOAST and SISAL*

---

### Description

A function to conduct complete reference-free deconvolution on DNA methylation data. If a full reference or a partial reference panel is provided, this function also automatically annotate the solved proportions to known cell types.

### Usage

```
Tsisal(Y_raw, K = NULL, knowRef = NULL, possibleCellNumber = 3:15)
```

### Arguments

**Y\_raw** The DNA methylation 450K array data from complex tissues, rows for CpG sites and columns for samples.

**K** The number of pure cell types, we allow users to pre-specify or use our method to estimate.

**knowRef** The external reference panel for cell type label assignment.

**possibleCellNumber** Range of possible number of cell types. Default is 3:15.

### Value

**estProp** Estimated proportions.

**selMarker** Selected cell type-specific markers.

**K** Optional number of cell types.

### Author(s)

Weiwei Zhang <wwzhangly@163.com>

### References

Complete deconvolution of DNA methylation signals from complex tissues: a geometric approach. Weiwei Zhang, Hao Wu and Ziyi Li.

### Examples

```
### generate a simulation data
knowRef <- matrix(runif(5000*5), 5000, 5)
colnames(knowRef) <- paste0("CellType", 1:5)
Y_raw <- matrix(runif(5000*20), 5000, 20)
rownames(Y_raw) <- paste0("CpG", 1:5000)
colnames(Y_raw) <- paste0("Sample", 1:20)
```

```
Tsisal(Y_raw = Y_raw, K = 5, knowRef = knowRef)

## if cell type number is unknown
# Tsisal(Y.raw = Y_raw, K = NULL, knowRef = knowRef, possibleCellNumber = 4:10)
```

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